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# STUDYING OF BISPHENOL A LEVELS IN SOME CANNED FOOD, FEED AND BABY BOTTLES IN EGYPTIAN MARKETS

Mohyelden A Osman<sup>1</sup>, Ghada I Mahmoud<sup>1</sup>, Mohamed H Elgammal<sup>2</sup>, Randa S Hasan<sup>2,\*</sup>

<sup>1</sup>Biochemistry Department, Faculty of Agriculture, Cairo University P. Box 12613, Gamma St, Giza, Egypt

<sup>2</sup>Regional Centre for Food and Feed (RCFF), Agricultural Research Centre (ARC) P. Box 588, Orman, Giza, Egypt

## ABSTRACT

Bisphenol A (BPA) is an endocrine disrupter which causes negative effects on human health. The aim of the present study is to determine BPA levels leaching of several kinds of canned foods, beverages, baby bottles and feed in local Markets by using gas chromatography-mass spectrum (GC/MS/MS). Furthermore, estimate a relationship between BPA migrations from polycarbonate baby bottles, temperature, and effect of numbers of use. Results showed that in more than 130 different samples taken from Egyptian markets, the highest mean BPA level was in chicken cocktail sausaged (710.59 ppb) which was given highly significant results with regard to all other food samples result, whilst the lowest was for tomato ketchup at (5.75 ppb). BPA levels were ranged between 31.02 and 724.06 for canned meat, 5.57 and 233.78 for vegetables and fruits samples and 30.89 and 312.08 for milk samples. It was found that the levels of BPA ranged between (233.65 to 933.75 ppb) in feed additives. The leaching of BPA from baby bottles to milk was found to be highest contents (123.53 ppb) compare with other baby drinks. The effect of temperature and numbers of use on BPA migration from baby bottles to milk results indicated a positive relationship between them, whereas an increasing of temperatures and numbers of use increased BPA migration levels in milk. The concentration of BPA in milk has reached 1046.79 ppb when baby bottles were used 100 times at 90 °C.

## KEYWORDS:

Baby bottles, Bisphenol A, Feed, Food, Numbers of use, Temperature.

## INTRODUCTION

Years ago endocrine disruptors have caught the attention of the world because of their possible harmful effects on human health. Bisphenol A (BPA) is a substance of worry since it is an endocrine disruptor and has been related to different negative health effects [1].

BPA is an organic synthetic compound consists of two phenol rings associated with a methyl connect, with have two methyl functional groups joined to the bridge. BPA made by mixing acetone and phenol is utilized to produce epoxy resins and polycarbonate (PC) plastics. Epoxy resins are utilized to lining metal cans to avoid touch between the inside can surface and the food, for prevent can erosion and rustiness [2]. BPA can be leaching when canned food is heated to typical can processing temperatures [3]. BPA can also be leaching from deficiently polymerized epoxy resins.

These protective layer are like utilized for metal covering in cap for food in glass bottles [2]. Because of a deficient polymerization operation, remains of BPA monomer in PC container and covering can be leaching into food, mostly throughout storage and manufacturing at elevated temperatures [4].

Polycarbonate plastics are utilized in food and drink canned and reusable jugs. The carbonate linkages are relatively steady however can be hydrolyzed at high temperature and unbiased to antacid pH, bringing about the arrival of BPA [5]. Various variables including pH, salt, oil, and glucose have been appeared to impact the movement of BPA from the lacquer and plastic infant bottle to the continuer content [6, 7].

The major risk of BPA is food exposition for kids and elderly [8]. BPA caused harmful health effect so should domination leaching from container coatings by setting assizes by various agencies. The leaching limit for BPA in food was confirmed by [9] at 600 ng/g whilst the tolerable daily intake (TDI) and maximal reference dose were confirmed at (50 and 25 µg/kg body weight/day) by [10, 11]. As of late, another preparatory 10 time bring down TDI of 5 µg/kg b.w. /day was prescribed by [12].

Inside mice, low measurements presentation has been related to early maturity [13], increased nervous action [14], and changed maternal conduct [15]. Exposition to BPA through different life stages has additionally been related to specific diseases, including mammary gland and prostate tumors; diabetes; and immune system modification [16]. Rising urinary BPA levels have been significantly connected with male sexual dysfunction as depicted by

seven files, including reduced sexual lust, erectile, and ejaculatory issues [17, 18]. Many studied on animal indicated BPA to be a regenerative, formative, and systemic toxicant, and lowly estrogenic [19]. Woodruff et al., [20] Examined the number of chemicals to which pregnant ladies in the U.S. are uncovered discovered BPA in 96% of studied ladies. Moreover, drinking water and different drinks from plastic containers made with BPA raised the urinary concentration of the dangerous compound by about 70% [21].

The point of this survey was to determine BPA levels leaching in several kinds of canned foods, feed, beverages and drinks in baby bottles in Egyptian Markets. Furthermore, estimate a relationship between BPA migrations from polycarbonate baby bottles, temperature, and effect of numbers of use.

## MATERIALS AND METHODS

**Materials and reagents.** Bisphenol A standard (minimum purity  $\geq 99\%$ ) was purchased from Sigma–Aldrich (UK), n-Heptane (Analytical reagent grade, Fisher chemical), Acetonitrile (J. T. Baker, HPLC grade), Anhydrous sodium sulphate (Sigma-Aldrich), Potassium carbonate (Sigma-Aldrich), Acetic acid anhydrous (Honeywell, 99.8%) and distilled water.

**Samples. (1) Canned Food samples.** Different types of market packing food, three cans for each sample which include (Beef, sausaged, luncheon, tuna, Beans, Mushroom, Sweetcorn, Tomato ketchup, Tomato paste, oil, pickles, fruit, milk, and Beverages) were purchased from different local markets in Egypt.

**(2) Feed samples.** Seven feed additives samples (a mixture of amino acid, vitamin and essential oil addition in the diet for animal) were purchased from different countries in different types of plastic bottles.

**(3) Water samples.** Reusable water bottles. Four brands of mineral water bottles have been chosen.

**(4) Drinks in baby bottles.** Different liquids given to baby in a baby bottle i.e. anise, milk, and water.

**Determination of Bisphenol. A Sample preparation.** The whole contents of a can were fully blended utilizing homogenizer. Twenty gram (g) of subsample was taken. All content canned samples were extracted by adding n-heptane and acetonitrile (20 ml + 20 ml) into glass measure utilizing a homogenizer for five min. The extract was remained

for 15 min then filtrated by utilizing Double rings paper and a Buchner flask. The heptane was taken away, utilizing glass syringe then make another extraction on residue sample in filter paper by adding new acetonitrile 20 ml utilizing the Omni blender homogenizer. Then filtered another extraction as before and removed heptane. The twice acetonitrile was collective and added anhydrous sodium sulphate (30 g, 1 min), then poured the mixture into cylinder 100 ml. Canned samples which content little lipids, such as meat, fish, oil, vegetables, fruits and drinks heptane was decanted. The mixture was steamed to around 5 ml, then dilution by water to 50 ml and put in separating funnel 300 ml. Added potassium carbonate and methanol {10 ml (72% w/v), 10 ml} respectively, then added 10 ml of acidic anhydride to the mixture whirled slowly. Then left the mixture for 15 min and extracted by n-heptane 5 ml. The heptane solvent was gathered and determine by GC/MS/MS.

Baby formula samples were extracted by weighted ten g of sample and added acetonitrile 20 ml into glass beaker 50 ml and shaking it for 5 min. Baby formulae samples were extracted by weighted ten g of sample and added acetonitrile 20 ml into glass beaker 50 ml and shaking it for 5 min. The mixture was permitted to settle, then filtered an acetonitrile by a Double rings paper. The remains of a sample were extracted again by added acetonitrile 10 ml, then shaken and filtered. The remains of sample was extracted again by added acetonitrile 10 ml, then shaken and filtered. The two mixture were collective and steamed such as mentioned above. Drinks samples should open to permit gas to go out before extraction. Fifteen ml of the sample was put into separation funnel 250 ml and extracted as previously mentioned. The experimental of leaching BPA into water samples from the bottles added boiling water to all bottles and left one day [22].

**Effect of temperature and numbers of use on PC baby bottles.** The PC baby bottles were brought with various brand from a local pharmacy in Egypt and taken three bottles from each brand. All bottles were rinsed with water. Baby bottles are always washed up and sterilized before being used and filling with water, anise, and milk to each bottle to its capacity and left one hour. Then tested the effect of different temperature and times of use it.

**Condition of GC/MS/MS.** The determination of the bisphenol A was done utilizing a gas chromatography (Agilent Technologies 7890A) connected by a mass-selective detector (MSD, Agilent 7000). Which content 5% - phenyl methyl polysiloxane (HP-5ms, Agilent) and column (capillary, 30 m  $\times$  0.25 mm i. d. and 0.25  $\mu$ m film thickness Helium was used as carrier gas by velocity (one ml/ min). The program of the oven was (120 °C / 1 min, 1.5 min at a rate of 30 °C/ min then 300 °C /1.7 min at a rate of 35 °C/min). The temperature of injector and

**TABLE 1**  
**Bisphenol A Levels in food from local market**

Products	Brand	PH	Pack type	BPA Conc. (ppb)	Average range
<b>Meat</b>					
Corned beef	A	6.1	Can	255.78 <sup>c</sup> ± 0.39	(255.10 - 255.78)
Corned beef	B	6.1	Can	84.49 <sup>mm</sup> ± 1.12	(82.46 - 86.31)
Chicken cocktail sausaged	A	6.4	Can	710.59 <sup>a</sup> ± 7.97	(696.51 - 724.06)
Chicken luncheon meat	B	5.3	Can	32.52 <sup>a</sup> ± 0.87	(31.02 - 34.02)
Beef luncheon meat		5.9	Can	75.50 <sup>p</sup> ± 0.88	(74.56 - 77.26)
<b>Mean</b>				231.77 ± 125.41	(31.02 - 724.06)
<b>Fish</b>					
Light meat tuna with chili (shredded)	A	6.5	Can	94.11 <sup>k</sup> ± 1.51	(91.35 - 96.56)
Dark shredded tuna meat skipjack	B	6.1	Can	60.80 <sup>o</sup> ± 1.77	(57.68 - 63.79)
Light meat tuna (fancy solid)	C	5.8	Can	146.49 <sup>hi</sup> ± 2.80	(141.35 - 150.96)
Dark shredded tuna	D		Can	167.94 <sup>g</sup> ± 0.32	(167.41 - 168.52)
<b>Mean</b>				117.34 ± 28.21	(57.68 - 168.52)
<b>Vegetables and Fruits</b>					
<b>Beans</b>					
Plain Mesdames fava beans	A	6.2	Can	49.59 <sup>p</sup> ± 0.85	(48.66 - 51.28)
Fava beans with Salsa	C	5.8	Can	114.14 <sup>j</sup> ± 1.32	(112.22 - 116.67)
Bean in plastic bag	D	6.6	Plastic	14.90 <sup>uv</sup> ± 0.18	(15.20 - 14.91)
White kidney beans	A	6.2	Can	164.77 <sup>e</sup> ± 1.12	(162.54 - 165.89)
Red kidney beans dark	B	5.8	Can	60.66 <sup>o</sup> ± 0.89	(59.01 - 60.92)
<b>Mushroom</b>					
Mushroom	A	5.7	Can	23.48 <sup>st</sup> ± 0.11	(23.3 - 23.69)
Mushroom	B	5.7	Can	17.61 <sup>tu</sup> ± 0.21	(17.18 - 17.82)
<b>Sweet corn</b>					
Whole kernel golden sweet corn	A	6.5	Can	6.14 <sup>w</sup> ± 0.23	(5.68 - 6.38)
Kernel sweet corn	B	5.7	Can	7.46 <sup>w</sup> ± 0.13	(7.24 - 7.70)
<b>Tomato</b>					
Tomato paste	A	4.1	Can	83.20 <sup>mn</sup> ± 1.07	(81.91 - 85.33)
Tomato paste	B	4.0	Can	19.62 <sup>tu</sup> ± 0.23	(19.29 - 20.06)
Tomato ketchup	A	3.7	Plastic	8.69 <sup>vw</sup> ± 0.09	(8.51 - 8.81)
Tomato ketchup	B	3.6	Plastic	9.75 <sup>vw</sup> ± 0.52	(8.72 - 10.30)
Tomato ketchup	C	3.7	Plastic	5.75 <sup>w</sup> ± 0.09	(5.57 - 5.88)
Local tomato sauce of koushari	D	4.3	Plastic	231.59 <sup>e</sup> ± 1.32	(229.21 - 233.78)
<b>Pickles</b>					
Pickles (olive)	A	3.4	Plastic	29.42 <sup>qr</sup> ± 0.39	(28.69 - 30.04)
Pickles (lemon)	B	2.3	Plastic	10.46 <sup>vw</sup> ± 0.37	(10.02 - 11.19)
<b>Oil</b>					
Local market oil	A		Plastic	79.44 <sup>mn</sup> ± 3.87	(72.1 - 85.22)
<b>Fruits</b>					
Peach (halves of peaches in light syrup)	A	3.2	Can	74.63 <sup>n</sup> ± 0.17	(74.31 - 74.89)
Peaches (halves in light syrup)	B	3.8	Can	22.24 <sup>st</sup> ± 0.99	(21.14 - 24.21)
<b>Mean of vegetable and fruits</b>				51.53 ± 12.79	(5.57 - 233.78)
<b>Milk</b>					
Cream cheese spread	A	6.5	Plastic	31.38 <sup>q</sup> ± 0.24	(30.89 - 31.62)
Cream	B	6.5	Can	219.85 <sup>f</sup> ± 0.80	(218.33 - 221.04)
Milk (infant milk formula)	A	5.9	Can	89.35 <sup>l</sup> ± 1.06	(87.34 - 90.95)
Milk (sweetened condensed milk)	B	6.6	Can	33.43 <sup>q</sup> ± 0.48	(32.71 - 34.35)
Yoghurt	A		Plastic	236.76 <sup>d</sup> ± 1.67	(234.95 - 240.1)
Dannet chocolate	A	6.7	Plastic	79.28 <sup>mn</sup> ± 0.64	(78.21 - 80.43)
Dannet vanilla	B	6.5	Plastic	311.74 <sup>b</sup> ± 0.17	(311.50 - 312.08)
<b>Mean</b>				129.38 ± 55.30	(30.89 - 312.08)
<b>Beverages</b>					
Orange juice	A	3.9	Plastic	149.36 <sup>b</sup> ± 3.67	(142.2 - 154.34)
Soft drink Orange juice	B	3.3	Plastic	35.53 <sup>q</sup> ± 0.15	(35.36 - 35.83)
Guava juice	A	4.0	Plastic	17.06 <sup>mn</sup> ± 0.26	(16.72 - 17.58)
Guava juice	B	4.0	Plastic	32.63 <sup>q</sup> ± 0.28	(32.24 - 33.17)
Nectar mango	A	3.7	Plastic	25.79 <sup>rs</sup> ± 0.27	(25.36 - 26.28)
Mango float	B	3.6	Can	14.50 <sup>uv</sup> ± 0.28	(13.96 - 14.87)
<b>Mean</b>				45.81 ± 29.71	(13.96 - 154.34)
<b>L.S.D 0.05</b>				4.6280	

- Each value represents the mean ± S.E (Standard Error) and mean of three replicates.

- Values in the same column with the same letter are not significant at p ≤ 0.05.

- Brand (A, B, C and D) means different company.

- PET (Polyethylene Terephthalate).

detector were 250 °C. The mode of injection was (split to split, rate 1: 20) and one µl of volume. The mass has parameter ionization potential and interface temperature (70 eV, 280 °C). The system of selected ion monitoring (SIM) was utilized (213 and 228 m/z). Quantitative results were acquired utilizing the GC-MS outcomes by comparing with external standards.

**Statistical analysis.** Statistical analysis (standard error “SE”) was done by [23]. LSD (Least significant difference) analysis was utilized to contrasts any significant differences between the means of treatment [24]. All static analysis was done by Costat program.

## RESULTS AND DISCUSSION

**Canned food results.** BPA contents in food samples under study are appeared in Table 1. It was seen that chicken cocktail sausaged (710.59 ppb) which have highest average BPA concentration result contrasted with other samples results, trailed by danned vanilla (311.74 ppb); whilst the lowest were for tomato ketchup at (5.75ppb).

Our results agree with [25] indicated for BPA levels in canned corned beef and chicken ranged (17 - 602 ppb). Also, [22] reported for infant formula and milk (9 - 384 ppb) which agree with present results.

The appearance of BPA in canned food due to that the canned foods were sterilized during the production process. Goodson et al. [26] reported that storage factor of the can such as time, temperature and state of the can may affect the levels of BPA in food.

Besides to food in paper containers, the release of BPA into food due to mostly reused this container and can lining materials [27]. BPA will probably migrate into oily, fatty foods or high protein contents food such as meat, fish and coconut cream [28].

**Results of baby bottles samples.** Table 2 gives the summarized results about BPA levels in two different types baby bottles filling with various kinds of liquid. These data indicate the levels of BPA at PC baby feeding bottles and water drinking bottles. As can be seen, the highest mean BPA level values was in milk in baby bottle brand (A) (123.53 ppb) which was a highly significant result with respect to all drinks samples results, trailed by anise in baby bottle brand (B) (114.94 ppb); whilst the lowest in water in bottle (3.88 ppb).

From above result, it could make seen that the BPA was leaching from the PC bottles to the drinking water they contain. The highest level of BPA in milk due to milk content amines like putrescine and 1,4-diaminobutane which two amines that produce from degradation of protein in milk that simulants migration of BPA and that has been a statement by [29, 30].

The high level may be due to the exposure of baby bottles to the high temperature of the anise or milk while that is not happening in water samples.

**Results of feed additives samples.** The concentrations of BPA in feed additives are shown in Table 3. It was noted that the levels of BPA ranged between (233.65 to 933.75 ppb) in feed additives bottles which made from PC sample (A, B and C), which agree with [31, 25] which reported that levels of BPA in canned pet foods ranged (179 and 602 ppb). While the levels of BPA ranged between (2.03 to 4.24 ppb) in feed additives bottles which made from HDPE samples (D, E, F, and G).

BPA contents in packing material, like as, HDPE, Tetra Pak, glass, and PET were very smaller than PC canned foods and drinks. This showed that low-level presence of BPA through food processing and before to filling into the container which as reported by [32]. As the presences of BPA in feed additives probably will be found in animal milk which could be risked on human as a result.

**TABLE 2**  
**Bisphenol A Levels in two baby bottles after 1h from filling**

Products	Brand	PH	Pack type	BPA Conc. (ppb)	Range
Anise in baby bottle	A	8.50	PC	6.53 <sup>ef</sup> ± 0.18	( 6.21 - 6.84)
Anise in baby bottle	B	8.50	PC	114.94 <sup>b</sup> ± 1.17	( 112.68 - 116.56)
Milk in baby bottle	A	7.00	PC	123.53 <sup>a</sup> ± 1.27	( 121.28 - 125.66)
Milk in baby bottle	B	7.00	PC	70.27 <sup>c</sup> ± 0.28	( 69.83 - 70.78)
Water in baby bottle	A	8.50	PC	4.60 <sup>fg</sup> ± 0.39	( 3.83 - 5.13)
Water in baby bottle	B	8.60	PC	16.55 <sup>d</sup> ± 0.42	( 16.13 - 17.39)
Water in bottle	A	8.70	PC	5.38 <sup>efg</sup> ± 0.46	( 4.47 - 5.93)
Water in bottle	B	8.80	PC	3.88 <sup>g</sup> ± 0.08	( 3.79 - 4.04)
Water in bottle (reused mineral water bottles)	C	8.70	PET	7.05 <sup>e</sup> ± 0.18	( 6.69 - 7.23)
Water in bottle (water bottle)	D	8.30	PET	4.48 <sup>fg</sup> ± 0.16	( 4.26 - 4.80)
<b>Mean</b>				35.72 ± 27.99	( 3.88 - 123.53)
<b>L.S.D 0.05</b>				1.7883	

- Each value represents the mean ± S.E (Standard Error) and mean of three replicates.

-Values in the same column with the same letter are not significant at p≤0.05.

- Brand (A, B, C and D) means different company.

- PET (Polyethylene Terephthalate), PC (Polycarbonate).

**TABLE 3**  
**Bisphenol A levels in Feed additives**

Products	Brand	PH	Pack type	BPA Conc. (ppb)	Range
Feed additives	A	3.80	PC	933.75 <sup>a</sup> ± 2.49	( 929.03 - 937.49)
Feed additives	B	7.70	PC	414.48 <sup>b</sup> ± 2.51	( 410.45 - 419.07)
Feed additives	C	6.30	PC	271.09 <sup>c</sup> ± 3.93	( 265.86 - 278.78)
Feed additives	D	6.80	HDPE	7.05 <sup>d</sup> ± 0.38	( 6.41- 7.71)
Feed additives	E	2.70	HDPE	4.24 <sup>d</sup> ± 0.28	( 3.69 - 4.62)
Feed additives	F	4.70	HDPE	2.91 <sup>d</sup> ± 0.02	( 2.87 - 2.95)
Feed additives	G	2.30	HDPE	2.03 <sup>d</sup> ± 0.04	( 1.96 - 2.07)
<b>Mean</b>				233.65 ± 202.33	( 2.87 - 937.49)
<b>L.S.D 0.05</b>				14.1729	

- Each value represents the mean ± S.E (Standard Error) and mean of three replicates.
- Values in the same column with the same letter are not significant at  $p \leq 0.05$ .
- Brand (A, B, C, D and G) means different company.
- HDPE (High density polyethylene), PC (Polycarbonate).

**TABLE 4**  
**Bisphenol A levels in milk from PC baby bottles as affected by PC source, temperature and numbers of use**

Numbers of use	Type	Temperature / Conc. (ppb)			Range
		25 °C	60 °C	90°C	
<b>New bottles</b>	A	48.50 <sup>e</sup> ± 0.89	65.62 <sup>h</sup> ± 0.40	105.90 <sup>i</sup> ± 0.33	65.62 <sup>h</sup> ± 0.40
	B	38.04 <sup>h</sup> ± 0.28	165.02 <sup>g</sup> ± 0.42	222.90 <sup>g</sup> ± 1.47	165.02 <sup>g</sup> ± 0.42
	C	67.61 <sup>f</sup> ± 1.09	161.94 <sup>g</sup> ± 1.35	205.71 <sup>h</sup> ± 1.33	161.94 <sup>g</sup> ± 1.35
<b>Mean</b>		51.38 ± 8.66	130.86 ± 32.63	178.17 ± 63.48	
<b>50</b>	A	137.80 <sup>e</sup> ± 0.39	190.28 <sup>f</sup> ± 4.07	399.38 <sup>f</sup> ± 2.01	190.28 <sup>f</sup> ± 4.07
	B	291.57 <sup>d</sup> ± 1.14	436.40 <sup>d</sup> ± 4.33	623.81 <sup>d</sup> ± 3.02	436.40 <sup>d</sup> ± 4.33
	C	132.54 <sup>e</sup> ± 1.36	265.70 <sup>e</sup> ± 2.00	524.85 <sup>e</sup> ± 3.50	265.70 <sup>e</sup> ± 2.00
<b>Mean</b>		187.30 ± 52.16	297.46 ± 72.80	516.01 ± 64.94	
<b>100</b>	A	405.01 <sup>c</sup> ± 0.78	527.37 <sup>c</sup> ± 1.75	842.15 <sup>c</sup> ± 1.31	527.374 <sup>c</sup> ± 1.75
	B	614.04 <sup>a</sup> ± 3.98	918.25 <sup>a</sup> ± 3.72	1022.92 <sup>b</sup> ± 2.05	918.25 <sup>a</sup> ± 3.72
	C	471.32 <sup>b</sup> ± 4.32	883.53 <sup>b</sup> ± 3.61	1046.79 <sup>a</sup> ± 4.81	883.53 <sup>b</sup> ± 3.61
<b>Mean</b>		497.12 ± 61.06	776.38 ± 124.19	970.62 ± 64.61	
<b>L.S.D 0.05</b>		6.2920	8.3568	7.5659	

- Each value represents the mean ± S.E (Standard Error) and mean of three replicates.
- Values in the same column with the same letter are not significant at  $p \leq 0.05$ .
- Brand (A, B and C) means different company.
- PC (Polycarbonate).

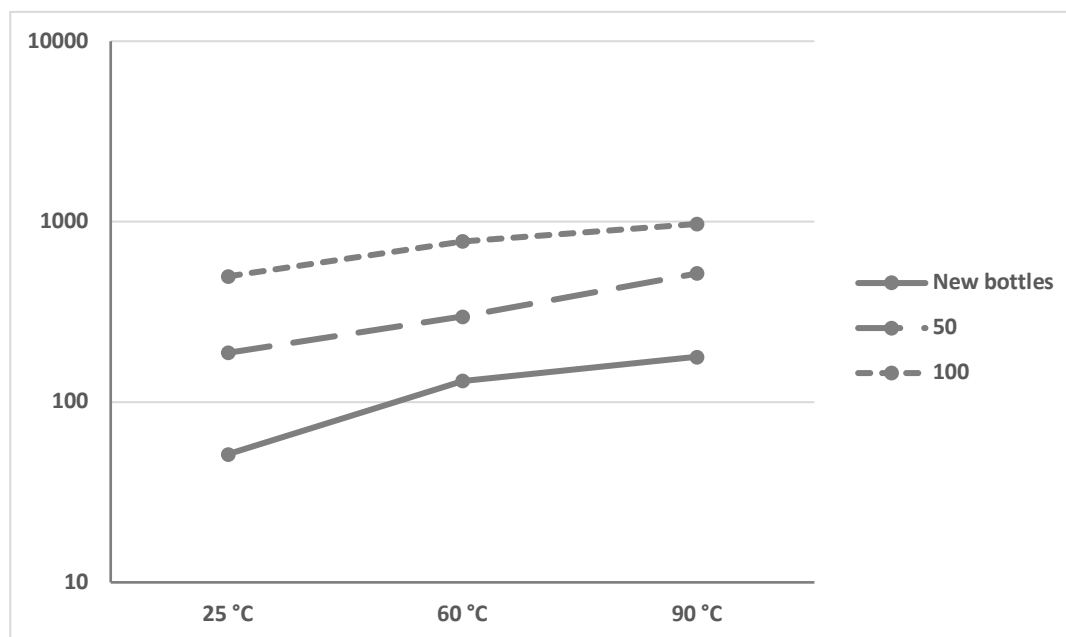
**Effect of temperature and numbers of use of PC baby bottles on BPA levels.** Table 4 including the numbers of use and temperature factors in following the leaching of baby food from baby bottles. The result clearly that there was an appositive relation between the release of BPA in milk, numbers of use and temperature. The BPA concentration was found to increase with raise the temperature of milk and increasing the numbers of using. Where found the higher concentration of BPA baby milk bottles when used 100 times at 90 °C (1046.79 ppb) and the lower concentration BPA when using new bottles at 25 °C (38.04 ppb). Fig. (1) Show that there was a linear relation between temperature, number of use and concentration of BPA.

Environment Canada [33] indicated that leaching from new bottles is very low, but can increase with repeated use, due to cleaning. Hoekstra and Simoneau [34] reported that the appearance of BPA into water due to increasing time and temperature. Le et al., [35] investigated that the concentration of

BPA in boiling water which leaching from new polycarbonate drinks bottles higher than water at room temperature by factor 15 - 55.

## CONCLUSION

Many various types of canned food and baby bottles could be a hazard factor for human and infant health because of the presence of variable levels of BPA. After analysis of more than 180 different sample which including meat, vegetable, baby bottles and feed additives. It was found different levels of BPA between (2.03 and 1046.79 ppb) which depend on their circumstances of production, transportation, storage, and migration process of the sample itself. Much care should be taken when chosen the material of baby bottles because of there was found that appositive relation between numbers of use and temperature with a concentration of BPA leaching in milk from PC infant bottles.



**FIGURE 1**

**Bisphenol A levels in milk from PC baby bottles as affected by PC source, temperature and numbers of use.**

To avoid exposure to contamination of food with BPA it was recommended to use more safety material in food packaging such as PET, glass, HDPE, Tetra Pack and Polypropylene (PP).

## REFERENCES

- [1] Tsai, W.T. (2006) Human health risk on environmental exposure to bisphenol-A: a review. *Journal of Environmental Science and Health. Part C.* 24(2), 225-255.
- [2] Cao, X.L., Perez-Locas, C., Dufresne, G., Clement, G., Popovic, S., Beraldin, F., Dabeka, R. W. and Feeley, M. (2011) Concentrations of bisphenol A in the composite food samples from the 2008 Canadian total diet study in Quebec City and dietary intake estimates. *Food Additives and Contaminants.* 28(6), 791-798.
- [3] Kawamura, Y., Inoue, K., Nakazawa, H., Yamada, T. and Maitani, T. (2001) Cause of bisphenol A migration from cans for drinks and assessment of improved cans. *Shokuhin eiseigaku zasshi. Journal of the Food Hygienic Society of Japan.* 42(1), 13-17.
- [4] Noonan, G.O., Ackerman, L.K., and Begley, T.H. (2011) Concentration of bisphenol A in highly consumed canned foods on the US market. *Journal of agricultural and food chemistry.* 59(13), 7178-7185.
- [5] Ben-Jonathan, N. and Steinmetz, R. (1998) Xenoestrogens: the emerging story of bisphenol A. *Trends in Endocrinology and Metabolism.* 9(3), 124-128.
- [6] Takao, Y., Lee, H.C., Ishibashi, Y., Kohra, S., Tominaga, N. and Arizono, K. (1999) Fast Screening Method for Bisphenol A in Environmental Water and in Food by Solid-Phase Microextraction (SPME). (Proceedings of 24th Symposium on Toxicology and Environmental Health). *Journal of Health Science.* 45(1), P39-39.
- [7] Kang, J.H., Kito, K. and Kondo, F. (2003) Factors influencing the migration of bisphenol A from cans. *Journal of food protection.* 66(8), 1444-1447.
- [8] EFSA (2013) Draft Scientific Opinion on the risks to public health related to the presence of bisphenol A (BPA) in foodstuffs –Part: exposure assessment. EFSA Panel on food contact materials, enzymes, flavourings and processing aids. Draft for public consultation.
- [9] EC Directive (2004) EC Directive (2004)/19/EC of 1 March (2004) amending Directive (2002)/72/EC relating to plastic materials and articles intended to come into contact with foodstuffs. *Official Journal of the European Union,* 71, 8–21.
- [10] EFSA (2006) Opinion of the scientific panel on food additives, flavourings, processing aids and materials in contact with food on a request from the Commission related to 2, 2-bis (4-hydroxyphenyl) propane (Bisphenol A). *EFSA Journal* (2006). 428, 1-75.

- [11] Health Canada (2008) Health risk assessment of bisphenol A from food packaging applications. Available from: [http://www.hc-sc.gc.ca/fn-an/alt\\_formats/hpfb-dgpsa/pdf/securit/bpa\\_hra-ers-eng.pdf](http://www.hc-sc.gc.ca/fn-an/alt_formats/hpfb-dgpsa/pdf/securit/bpa_hra-ers-eng.pdf).
- [12] EFSA (2014) Draft Scientific Opinion on the risks to public health related to the presence of bisphenol A (BPA) in foodstuffs. EFSA Panel on food contact materials, enzymes, flavourings and processing aids. Draft for public consultation, January 2014.
- [13] Howdeshell, K.L., Hotchkiss, A.K., Thayer, K.A., Vandenberg, J.G., and Vom Saal, F.S. (1999) Environmental toxins: exposure to bisphenol A advances puberty. *Nature*. 401(6755), 763.
- [14] Ryan, B.C. and Vandenberg, J.G. (2006) Developmental exposure to environmental estrogens alters anxiety and spatial memory in female mice. *Hormones and behaviour*. 50(1), 85-93.
- [15] Palanza, P.L., Howdeshell, K.L., Parmigiani, S. and vom Saal, F.S. (2002) Exposure to a low dose of bisphenol A during fetal life or in adulthood alters maternal behavior in mice. *Environmental health perspectives*. 110(Suppl 3), 415.
- [16] Vom Saal, F.S., Akingbemi, B.T., Belcher, S.M., Birnbaum, L.S., Crain, D.A., Eriksen, M., Farabollini, F., Guillette, L.J. Jr., Hauser, R., Heindel, J.J., Ho, S.M., Hunt, P.A., Iguchi, T., Jobling, S., Kanno, J., Keri, R.A., Knudsen, K.E., Laufer, H., LeBlanc, G.A., Marcus, M., McLachlan, J.A., Myers, J.P., Nadal, A., Newbold, R.R., Olea, N., Prins, G.S., Richter, C.A., Rubin, B.S., Sonnenschein, C., Soto, A.M., Talsness, C.E., Vandenberg, J.G., Vandenberg, L.N., Walser-Kuntz, D.R., Watson, C.S., Welshons, W.V., Wetherill, Y. and Zoeller, R.T. (2007) Chapel Hill bisphenol A expert panel consensus statement: integration of mechanisms, effects in animals and potential to impact human health at current levels of exposure. *Reproductive toxicology* (Elmsford, NY). 24(2), 131.
- [17] Li, D.K., Zhou, Z., Miao, M., He, Y., Qing, D., Wu, T., Wang, J., Weng, X., Ferber, J., Herrinton, J., Zhu, Q., Gao, J. and Yuan, J. (2010) Relationship between Urine Bisphenol-A Level and Declining Male Sexual Function. *Journal of andrology*. 31(5), 500-506.
- [18] Li, D., Zhou, Z., Qing, D., He, Y., Wu, T., Miao, M., Wang, J., Weng, X., Ferber, J., Herrinton, J., Zhu, Q., Gao, J., Checkoway, J. and Yuan, W. (2009) Occupational exposure to bisphenol-A (BPA) and the risk of self-reported male sexual dysfunction. *Human reproduction*. 25(2), 519-527.
- [19] EPA (2010) Bisphenol A Action Plan (CASRN 80-05-7). U.S. Environmental Protection Agency. [www.epa.gov/opptintr/existingchemicals/pubs/actionplans/bpa.html](http://www.epa.gov/opptintr/existingchemicals/pubs/actionplans/bpa.html).
- [20] Woodruff, T.J., Zota, A.R., and Schwartz, J.M. (2011) Environmental chemicals in pregnant women in the United States: NHANES 2003–2004. *Environmental health perspectives*. 119(6), 878.
- [21] Braun, J.M., Kalkbrenner, A.E., Calafat, A.M., Bernert, J.T., Ye, X., Silva, M.J., Barr, D.B., Sathyanarayana, S. and Lanphear, B.P. (2011) Variability and predictors of urinary bisphenol A concentrations during pregnancy. *Environmental health perspectives*. 119(1), 131.
- [22] Goodson, A., Summerfield, W. and Cooper, I. (2002) Survey of bisphenol A and bisphenol F in canned foods. *Food Additives and Contaminants*. 19(8), 796-802.
- [23] Fisher, R.A. (1970) *Statistical method for research workers*. Edinburgh Ed. 14, Oliver and Boyd, 140p.
- [24] Waller, R.A. and Duncan, D.B. (1969) A Bayes rule for symmetric multiple comparison problem. *Journal of the American Statistical Association*. 64, 1485 – 1503.
- [25] Imanaka, M., Sasaki, K., Nemoto, S., Ueda, E., Murakami, E., Miyata, D. and Tonogai, Y. (2001) Determination of bisphenol A in foods using GC/MS. *Shokuhin eiseigaku zasshi. Journal of the Food Hygienic Society of Japan*. 42(2), 71-78.
- [26] Goodson, A., Robin, H., Summerfield, W. and Cooper, I. (2004) Migration of bisphenol A from can coatings—effects of damage, storage conditions and heating. *Food additives and contaminants*. 21(10), 1015-1026.
- [27] Ozaki, A., Yamaguchi, Y., Fujita, T., Kuroda, K. and Endo, G. (2004) Chemical analysis and genotoxicological safety assessment of paper and paperboard used for food packaging. *Food and Chemical Toxicology*. 42(8), 1323-1337.
- [28] Thomson, B.M. and Grounds, P.R. (2005) Bisphenol A in canned foods in New Zealand: an exposure assessment. *Food additives and contaminants*. 22(1), 65-72.
- [29] Rykowska, I., Szymański, A. and Wasiak, W. (2005) Determination of bisphenol-A in drinking water using new SPE sorbents with chemically bonded ketoimine groups. *Polish journal of food and nutrition sciences*. 14(3), 237.
- [30] Maia, J., Cruz, J.M., Sendón, R., Bustos, J., Cirugeda, M.E., Sanchez, J.J., and Paseiro, P. (2010) Effect of amines in the release of bisphenol A from polycarbonate baby bottles. *Food research international*. 43(5), 1283-1288.
- [31] FSA (2001) Food Standards Agency, Survey of bisphenols in canned foods. Food surveillance sheet No. 13/01. United Kingdom.



- [32] Geens, T., Apelbaum, T.Z., Goeyens, L., Neels, H. and Covaci, A. (2010) Intake of bisphenol A from canned beverages and foods on the Belgian market. *Food Additives and Contaminants*. 27(11), 1627-1637.
- [33] Environment Canada. (2008) Draft Screening Assessment for The Challenge Phenol, 4, 4'-(1-methylethylidene) bis-(bisphenol A) Chemical Abstracts Service Registry Number 80-05-7. Accessed at: [http://www.ec.gc.ca/substances/ese/eng/challenge/batch2/batch2\\_80-05-7.cfm](http://www.ec.gc.ca/substances/ese/eng/challenge/batch2/batch2_80-05-7.cfm). Accessed: 14 October.
- [34] Hoekstra, E.J. and Simoneau, C. (2013) Release of bisphenol A from polycarbonate—a review. *Critical reviews in food science and nutrition*. 53(4), 386-402.
- [35] Le, H.H., Carlson, E.M., Chua, J.P. and Belcher, S.M. (2008) Bisphenol A is released from polycarbonate drinking bottles and mimics the neurotoxic actions of estrogen in developing cerebellar neurons. *Toxicology letters*. 176(2), 149-156.

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#### **CORRESPONDING AUTHOR**

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**Randa S Hasan**

Regional Centre for Food and Feed (RCFF),  
Agricultural Research Centre (ARC),  
Giza, Cairo – Egypt

e-mail: [randa.saad@post.agr.cu.edu.eg](mailto:randa.saad@post.agr.cu.edu.eg)