



## Review Article

**A NATURAL FEED ADDITIVE PHYTOBIOTIC, POMEGRANATE  
(*PUNICA GRANATUM* L.), AND THE HEALTH STATUS OF POULTRY**

Wafaa A. Abd El-Ghany

*Poultry Diseases Department, Faculty of Veterinary Medicine,  
Cairo University, Giza 12211, Egypt*

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**ABSTRACT**

The addition of antibiotic growth promoters in poultry diets results in a development of resistant bacterial strains and accumulation of drug residues in the meat and eggs. The new trend in poultry industry is the dietary addition of natural feed additives including phytobiotics. Pomegranate (*Punica granatum* L.) is a natural cheap feed source that has been extensively used in the livestock production. In poultry production system, pomegranate by-products such as peel powder or extract, seed oil, or juice showed high nutritional values, several health benefits, and good economic profits. Pomegranate displays a growth promoting effect and an enhancement of carcass traits of broilers, along with an improvement of the egg production traits parameters. Moreover, dietary pomegranate by-products showed a potential antioxidant and antimicrobial effects on the treated birds. Modulation of both humeral and cell mediated immune response, hypo-lipidemia, as well as enhancement of liver functions have been proved following dietary treatment with different pomegranate by-products. Therefore, this review article was designed to present the different effects of dietary pomegranate by-products on the production indices of broilers and layers, the antioxidant, antimicrobial, and immune status, as well as the blood parameters.

**Key words:** antioxidant, chicken, immunity, performance, *Punica granatum* L.**INTRODUCTION**

The emergence of anti-microbial growth promoter's resistance becomes an important threat that affects poultry industry worldwide. Besides, accumulation of drug residues in the livestock edible tissues has an adverse effect on the human's health. Accordingly, the international authorities banned the use of growth promoter's antibiotics since 2006. Nowadays, there is a great demand for searching natural feed additives alternatives. Dietary supplementations of poultry with probiotics, prebiotics, parabiotics, postbiotics, microalgae,

and immunoglobulins preparations have been developed to improve the feed utilization efficiency and to maintain the general health conditions (1, 2, 3, 4, 5). The phytobiotics include a large variety of plant-derived products such as essential oils, extracts, herbs, and oleoresin which have positive impacts on the host's productivity and the final product quality (6, 7, 8). Moreover, several types of phytobiotics have been effectively used in poultry industry as growth promoters, antimicrobials, and immunomodulators (9, 10, 11, 12).

Pomegranate with a scientific Latin name (*Punica granatum* L., family Punicaceae) is commonly called in Latin as "ponus" and "granatus". It is a native fruit crop that has been widely used as one of the eldest known drug medicines. It has been mentioned in the ancient Egyptian herbs papyrus written in about 1550 BC (13). Moreover, the medicinal properties of pomegranate have been reported in China, Greece, India, Iran, South Africa, and Mediterranean countries such as Turkey, Tunisia,

*Corresponding author:* Prof. Dr. Wafaa A. Abd El-Ghany, PhD  
*E-mail address:* wafaa.soliman@cu.edu.eg  
*Present address:* Poultry Diseases Department, Faculty of Veterinary  
Medicine, Cairo University, Giza 12211, Egypt  
*Phone:* +0201224407992

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Morocco, and Spain (14, 15). Pomegranate has long been considered as a fruit with several health benefits (16). This fruit contains internal membranes which represent about 26-30% of its weight (17). It has been found that the edible parts of the plant involve 78% juice and 22% by-products. The by-products such as seed, husk, peel, etc. have several bioactive polyphenols compounds (18). The pomegranate peel (PP) by-product is rich in numerous high molecular weights polyphenols phytochemicals including phenolic acids (gallic, ellagic, caffeic, citric, tartaric, malic, and ascorbic), flavonoid (flavonols as catechin, epicatechin, and galloctechin as well as anthocyanins), hydrolyzable tannin (ellagitannins, punicalin, punicalagin, pedunculagin, and gallotannins), and condensed tannins (19). Polyunsaturated fatty acids, vitamins, and minerals are other components of pomegranate (20). The components of pomegranate seed oil are cis-9, trans-11, cis-13 octadecatrienoic (punicic) acid, and one conjugated linolenic acid (21). Therefore, pomegranate displayed numerous nutritional values and health-raising effects which can be used as a cheap alternative feed source in poultry diets (22, 23).

Besides, pomegranate and its by-product showed antioxidant, antimicrobial, anti-carcinogenic, anti-inflammatory, and growth promoting effects (24, 25, 26). In poultry, the PP, PP powder (PPP), PP extract (PPE), and pomace of PP have a great influence on the growth performance parameters. The improved production indices of broilers and layers (27, 28) as well as the enhanced immune response (22) of birds fed on pomegranate-derived additives have been described. The biological activities of PP as antibacterial and antioxidant have been investigated (29, 30, 31). Besides, the protective effect of PPP against pulmonary hypertension and mortality of broiler chickens has been demonstrated (32, 33). It is interesting to note that PPE showed an *in-vitro* antiviral potential, as its polyphenols content could prevent the entry of SARS-CoV-2 into the host cells (34).

According to the above mentioned, this review article aimed to describe the different effects of dietary pomegranate by-products on the production indices of broilers and layers, the antioxidant, antimicrobial, and immune status, as well as the blood parameters modifications.

## PRODUCTION INDICES

### *Broilers*

The improvement of the feed intake (FI), body weight (BW), BW gain (BWG), feed efficiency, feed conversion ratio (FCR), and carcass and organ parameters have been documented in birds fed on PPP and PPE including broiler chickens (35, 36, 37, 38, 39). Table 1 shows the different effects of PPP on the production indices of poultry (40, 41, 42, 43, 44, 45, 46, 47).

The potential growth-enhancement effect of pomegranate may be owed to the presence of essential oils which inhibit the pathogenic intestinal bacteria, enhance the intestinal flora, improve the nutrients digestion and energy utilization, and promote growth parameters. Moreover, PPP and PPE have different types of alkaloids such as flavonoids, glycosides, and tannins that improve the digestion through decreasing the intestinal motility and thus fluid accumulation (48). It has been reported that PP-containing proanthocyanidin could increase the pancreatic and the intestinal digestible enzymes, enhance the nutrient's absorption, and consequently prevent the harmful effect of free radicals on enterocytes (49). Furthermore, PPE is able to prevent the activity of pancreatic lipase and inhibits the intestinal fat absorption excreted in droppings (29). Addition of PPE of 2 mL/L dose in drinking water for broiler chickens increased the blood anti-radical activity and enhanced the intestinal microbial ecology via enhancing the lactic acid bacteria (50). The antioxidant and antimicrobial properties of *Punica granatum* fruit peel have a positive impact on the growth indices of birds (51). Saeed et al. (52) reported that *Punica granatum* L. by-product could modulate both the gut microbiota and immune function of broiler chickens and decrease the odorous gas emissions from droppings. Improvement of the villus length and villus length to villus depth ratio as well as increasing of the intestinal lactic acid bacteria have been found in broiler chickens and laying quails fed on PPE (50). The improvement in the nutrient's availability and intestinal absorption resulted in an enhancement of the bird's performance (44).

**Table 1.** The different effects of PPP on the production indices of poultry

| Dose of PPP/diet            | Species                       | Findings  | Reference |
|-----------------------------|-------------------------------|---|-----------|
| 15,000 mg/kg                | Broiler chicken               | No improvement of BW<br>Decreased ascites-related mortality<br>Decreased lipid oxidation of meat under refrigerated storage   | (40)      |
| 5%, 10%, and 15%            | Laying hens                   | High egg production<br>No effect on FI, egg weight, egg mass, and FCR   | (67)      |
| 0.5 and 1.0%                | Broiler chicken               | Decreased FI<br>Improved FCR  | (21)      |
| Up to 2%                    |                               | Enhance the quality of the breast and thigh muscles   |           |
| 0.5 and 1.0%                | Broiler chicken               | Reduced the FI and FCR  | (41)      |
| 0.01%                       | Broiler chicken               | Improved FI, BWG, and carcass weight percentage   | (35)      |
| 4.5-7.5%                    | Japanese laying quail         | Increased egg production, egg weight, and egg mass<br>No effect on the growth performance   | (68)      |
| 0.02%                       | Broiler chicken               | Impaired BWG and FI<br>Improved quality index of breast muscles   | (42)      |
| 2%                          | Broiler chicken               | Decreased liver, gizzard, and edible giblet proportion<br>Improved thigh muscles palatability   | (60)      |
| 0.05%                       | Broiler chicken               | Increased BW, BWG, and FCR  | (29)      |
| 3 g/kg                      | Broiler chicken               | Decreased growth performance parameters   | (43)      |
| 1%                          | Japanese laying quail         | No effect on egg production, specific gravity, and eggshell thickness   | (69)      |
| 1% and 1.5%                 | Japanese quail                | Improved carcass quality  | (61)      |
| 250 and 500 mg/kg           | Broiler chicken               | No effect on BWG, FCR, and carcass traits   | (44)      |
| 250, 450, and 650 mg/kg     | Heat stressed broiler chicken | Increased BWG and FCR<br>Decreased FI<br>No effect on relative weights of carcass, breast, thigh, abdominal fat, liver, pancreas, gallbladder, heart, gizzard, bursa of Fabricius, and spleen<br>Improved carcass quality during refrigerator storage | (38)      |
| 7.5 and 10 g/kg             | Broiler chicken               | Decreased FI<br>Improved FCR<br>Reduced heart weight and the right-to-total ventricular weight ratio  | (33)      |
| 2 or 4 g/kg                 | Broiler chicken               | Improved average final BW and daily BWG   | (45)      |
| 4%                          | Laying hens                   | Increased egg production  | (27)      |
| 6%                          | Japanese quails               | Increased final BW and BWG<br>Decreased total FI  | (46)      |
| 0.04%                       | Heat-stressed laying quails   | Improved egg quality  | (70)      |
| 0.25, 0.50, 1.00, and 1.50% | Broiler chicken               | Improved BW, BWG, FI, and FCR<br>Increased weights of spleen, thymus glands, and bursa of Fabricius<br>Reduced abdominal fat  | (26)      |
| 5-10 gm                     | Broiler chicken               | No effect on BW and FCR   | (25)      |
| 8%                          | Broiler chicken               | Decreased BWG, FI, FCR, and production index  | (22)      |
| 8%                          | Broiler chicken               | Decreased BWG and FI  | (23)      |
| 4%                          | Broiler chicken               | Improved gut morphology<br>Improved meat quality  |           |
| 0%, 5%, and 10%             | Laying hens                   | No significant effect on BW, Haugh unit, egg breaking strength, and yolk color of eggs  | (71)      |
| 2, 3, and 4 gm/kg           | Broiler chicken               | Improved BW and BWG<br>No effect on FCR<br>Improved all carcass characteristics, except gizzard and abdominal fat ratio were not affected   | (47)      |

On the other side, PPP or PPE supplementation has no effect on growth parameters in broiler chickens (53, 54). This discrepancy may be attributed to the fact that high doses of polyphenols (polyphenolic hydroxyl groups) in pomegranate by-products can bind to the endogenous proteins (protein carbonyl groups) in the intestinal luminal tract, thus leading to reduction in protein and amino acids digestibility (55). Similarly, Goni et al. (56) demonstrated that polyphenol contents of PP diminished the growth performance in broilers. The previous studies have revealed that tannins in PP adversely affects the physiological functions of the body in terms of improper nitrogen balances, suppressed growth, reduced intestinal absorption of amino acids and sugars, diminished immune response, and improved protein catabolism (57). Tannins also could bind to cholesterol and bile salts, and so, reduce the fat digestibility. Additionally, tannins may interfere with protein and calcium digestibility (58), which consequently induce adverse influences on the growth and other traits. Besides, the high quantity of tannins and crude fiber in PP could decrease the gastrointestinal motility and then reduce the bird's growth performance (22). It has been found that digestive enzymes such as trypsin, lipase, and amylase could bound to tannins and reduce the digestibility of nutrients (59). High levels of dietary PPP (8%) decreased the intestinal crypt depth and the villi/crypts ratio in the intestine of broiler chickens (23).

#### *Carcass traits*

Table 1 reveals the different influences of PPP on the carcass and organs in broilers (60, 61). Previous studies revealed significant improvement of carcass traits following supplementation with PPP (37, 43, 45, 62). An increase in the proportion of muscles resulted in more profitability in the broiler's industry (63). A dietary supplementation of broiler chickens with PPP induced a positive effect on the antioxidant status and consequently the meat quality (62, 64). Szymczyk and Szczurek (65) indicated that increasing the levels of pomegranate seed oil in broiler chickens feed resulted in an increase in the polyunsaturated acids, a decrease in the monounsaturated acids proportions, as well as an increase in the proportion of conjugated linoleic acid of the breast muscles. The improved quality of breast muscles may be attributed to increasing the total phenol and flavonoid contents following dietary inoculation of PPE (37). In the same context, dietary addition of PPE up to 300 mg/kg significantly increased the

phenolic content and antioxidant effect in the breast meat of broiler chickens (66).

In contrast, no improvement of the meat quality has been also reported after dietary supplementation with pomegranate by-product (22). Another study of Abdel Baset et al. (39) proved that broiler chickens fed on diets containing PPP showed reduced liver/carcass weight ratio.

#### *Layers*

The effects of PPP on the egg production parameters are shown in Table 1 (67, 68, 69, 70, 71). Most studies revealed an improvement in the production parameters of layer hens due to feeding on pomegranate (72). However, no effect of dietary supplementation with pomegranate by-products on the specific gravity and eggshell thickness of laying hens (71) and quails (69) has been reported. Similarly, Sharma et al. (73) did not find any effect of supplying layer hens with a blend of active ingredients from thyme, garlic, and pomegranate on the eggshell breaking strength. Kostogryś et al. (74) observed a dose-dependent increase in FI, egg production, and egg mass of layer hen's supplemented with pomegranate seed oil. This may be related to the antioxidant effect of pomegranate, or the improvement of intestinal enzymatic activity, and the consequent enhancement of digestibility and absorption of specific nutrients required for egg quantity and quality.

## ANTIOXIDANT ACTIVITY

A dietary supplementation of broiler chickens with PPP induced different effects on the antioxidant status of poultry (Table 2). Antioxidants in PPP could improve the feed efficiency via reducing the oxidative damage, increasing the gastro-intestinal enzymatic activity, and therefore enhancing nutrient's digestibility and absorption (36). The potential antioxidant character of pomegranate by-products may be due to their rich contents of ellagitannins (75). The polyphenols compounds in pomegranate such as tannins, anthocyanins, ponica, lagnine catechin, epicatechin, galocatechin, gallic, ellagic, caffeic, ellagitannins, and gallotannins could enhance the antioxidant activity in broilers and layers (76, 77, 78). Ahmed et al. (21) demonstrated that dietary inoculation with pomegranate improved the protein and fatty acid profile, but reduced the thiobarbituric acid reactive substances values in breast meat of broilers. Following absorption of

polyphenols-associated pomegranate, they bound by erythrocytes, which leads to an increase in the antioxidant blood capacity (79). Additionally, polyphenols and anthocyanins contents of PP could scavenge the hydroxyl and superoxide anion free radicals (43). The level of serum malondialdehyde (MDA) is an indication of lipid peroxidation and oxidative damage caused by reactive oxygen species (ROS). Also, catalase enzyme (CAT) inhibits the lipid peroxidation of hydrogen and peroxide toxicity. The antioxidant enzymes are able to neutralize the free radicals of different types of oxygen. The high concentration of CAT and superoxide dismutase (SOD) could improve the protection of cell membranes against oxidative stress and ROS (62). Pomegranate by-products could inhibit the peroxidation of lipid and induce over-expression of antioxidant genes including CAT and SOD. Dietary pomegranate could decrease the protein and DNA damage and possibly inhibit the ROS production (80), thus protecting the liver against oxidative stress and damage (44).

The variations in the antioxidant capacity of pomegranate depend on the type of its by-product (seed pulp, powder, extract, juice, etc.) and on the processing method (81). For instance, the presence of different amounts of phenolics which acts as hydrogen donors and reducing agents mediates the variation in pomegranate antioxidant activity (82). The results of Benchagra et al. (83) showed that PPP is rich in flavonoids and anthocyanin contents which have free radical scavenging and reducing power,  $\beta$ -carotene bleaching, and hydrogen peroxide scavenging activities. PPP has the ability to inhibit the conjugation and formation of diene and to reduce the  $\alpha$ -tocopherol disappearance induced by copper-sulfate-mediated low-density lipoproteins (LDL) peroxidation. In addition, PPE may modulate the activation of mitogen-activated protein kinase and nuclear factor kappa B in the 12-O-tetradecanoylphorbol13-acetate induced skin carcinogenesis model (84). PPE showed the highest removal activity of 2,2-diphenyl-1-picrylhydrazyl and 2,2'-azinobis [3-ethylbenzothiazoline-6-sulfonic acid] when compared to some other antioxidant fruits (85).

### ANTIMICROBIAL ACTIVITY

The various antimicrobial effects of PPP are illustrated in Table 2 (86, 87). Different *in-vitro* reports have shown the antibacterial and the antifungal influences of pomegranate (88, 89). The antibacterial

potential of pomegranate against Gram negative and Gram positive bacteria such as *Salmonella* spp., *Escherichia coli* (*E. coli*), *Staphylococcus aureus* (*S. aureus*), *Clostridium perfringens* (*C. perfringens*), *Listeria monocytogenes* (*L. monocytogenes*), *Bacillus cereus*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia*, and *Helicobacter pylori* has been proven (90, 91, 92, 93, 94, 95, 96). Moreover, the antifungal effect of pomegranate against some fungi such as *Aspergillus niger* has been demonstrated (97, 98). The antimicrobial activity of the phenolic and tannin-rich fractions of the PPP and seed extracts has been demonstrated (93).

The high content of phenolic acid, flavonoid, and hydrolysable tannins in PP is the main reason for its antimicrobial activity (99). The early study of Scalbert (100) showed that tannins inhibited the extracellular microbial enzymes and eliminated the substrates required for microbial multiplication, resulting in distraction of the membrane structure and function. Besides, tannins lowered the harmful gut microbial populations and inhibited their metabolism by preventing the oxidative phosphorylation process (101). The phenolic compounds could adhere to the pathogenic bacteria by the 'lectin-receptor' mechanism, inhibit the incorporation of pathogenic microorganism to the mucosa, and decrease the gut pathogenic bacterial count (102). Ferreira et al. (103) reported on the antimicrobial effects of tannin rich plant extract against  $\text{NH}_3$  producing *E. coli*, *Klebsiella* spp., and *Pseudomonas* spp. Further, the dissipation of the intestinal pH along with the antimicrobial activity of phenolic tannins may have been responsible for reducing the intestinal *E. coli* and *Salmonella* spp. count (41). The recent study of Tang et al. (104) revealed that the dietary ellagic acid could ameliorate *C. perfringens*-induced subclinical necrotic enteritis in broiler chickens. The authors owed this effect of ellagic acid to the regulation of the jejunal inflammation signaling pathways, reduction of the intestinal oxidative stress, balancing of the cecal microbiota, and inhibition of the intestinal barrier damage (104). Increasing in the intestinal *Lactobacillus* spp. count is usually associated with decreasing in the gut pathogenic bacteria. It seems that that PP could increase the count of the intestinal beneficial *Lactobacillus* spp. by reducing the pathogenic bacteria, and consequently improving the digestion and absorption of broiler chickens (105). In the same line, the *in-vitro* and *in-vivo* anti-parasitic properties of PP have been documented (106, 107, 108, 109, 110, 111, 112).



Phenolics, anthocyanins, hydrolysable tannins, and flavonoids are important anti-parasitic components of PP, flower, seed, and leaf (113).

On the other side, PPE has been widely used as a good food preserver due to its strong antimicrobial effect. A high concentration of PPE could inhibit the growth of bacteria causing spoilage and oxidative rancidity in chicken's products (86). In the same context, chicken meat products contain PPE showed an increase in their shelf life by 2-3 weeks during storage in refrigerator. Dipping of chicken's carcass in pomegranate juice decreased the microbial growth and population under refrigeration condition (114). Juneja et al. (94) demonstrated that PPP could enhance the heat inactivation of *E. coli* in ground chickens. Moreover, the earlier study of Hayrapetyan et al. (115) showed an inhibition of *L. monocytogenes* by PPE in meat paté at different temperatures.

### IMMUNO-MODULATION

It has been shown that pomegranate by-products could improve the immune response (36, 37, 38, 52). The immuno-potentiating effect of PPP is listed in Table 2 (116). For example, Saleh (59) found high concentrations of immunoglobulin (Ig) M and Ig G in broilers chickens supplemented by PP. Recently, Elsebai et al. (26) demonstrated an increase in weights of spleen, thymus glands, and bursa of Fabricius of broiler chickens as a result of supplementation with PPP at different dietary levels. On the other side, Rao et al. (44) reported that feeding of broiler chickens on PP significantly enhanced the antibody titers against Newcastle disease virus (NDV) vaccine, while the cell-mediated immune response to phyto-haemagglutinin phosphate was not significantly affected. Pomegranate seeds are a rich source of polyunsaturated fatty acid, especially  $\alpha$ -linolenic acid and linoleic acid (117, 118), which are responsible for increasing the weight of immune organs such as spleen and bursa of Fabricius (119).

### BLOOD PARAMETERS MODIFICATION

Different concentrations of PPP induced an enhancement in some hematological and biochemical parameters in the treated poultry species. The lowering effect of pomegranate on the lipid metabolism is shown in Table 2 (120). The hypo-cholesterolemic and hypo-lipidemic effects as well as interference with lipid

digestion, absorption, and metabolism have been demonstrated following supplementation with pomegranate (120). Pomegranate by-products have numerous key proteins and genes with a cholesterol-lowering effect (121). Supplementation of chicken's diet with pomegranate seed oil induced positive effects on the lipid profile of meat and eggs as well as their cholesterol content (65). Aviram and Fuhrman (122) also reported that PPE reduced the oxidation of LDL. In the same line, the concentration of cholesterol and LDL as well as the lipid peroxidation were decreased in broiler chickens fed on PPP (43). The anti-hyperlipidemic effect of PP was mediated by the down-regulation of peroxisome proliferator activated receptor alpha (33). Moreover, PPE reduced the total cholesterol and LDL through modulating of the activity of HMG-CoA reductase (123). It has been found that Ellagic could improve the cholesterol catabolism via increasing the production of bile acid in the hepatocytes (124). The *in-vitro* study of Zhao et al. (125) demonstrated that polyphenol in pomegranate significantly increased the expression of mRNA in the hepatic cells.

Many previous reports showed that dietary pomegranate by-products lowered the levels of some liver function enzymes such as aspartate transferase (AST) and alanine transaminase (ALT) in broiler chickens (22, 26, 39) and Japanese quails (68). Similar results were obtained by Sharifian et al. (38) in heat-stressed broiler chickens. The depressant effect of pomegranate on the liver enzymes is related to its phenolic constituent.

Regarding the effect of pomegranate on the other hematological parameters, it has been shown that PPP increased the count of red blood cells (RBCs) count (126). Similarly, PPP at dietary levels of 0.25, 0.50, 1.00, and 1.50% increased the count of RBCs and white blood cells, the hemoglobin concentration, and the packed cell values (26). However, addition of 8% PP to boiler's diet lowered the RBCs count, the hemoglobin concentration, and the packed cell value (22). The recent study of Gultepe et al. (71) revealed that pomegranate juice in 5%, and 10% water solution didn't induce significant effects on the lipid profile, liver enzyme, and most of the hematological parameters. The PPE significantly increased the RBCs count as well as the hemoglobin and hematocrit levels which indicated the increased erythropoiesis process, however, no significant effect on glucose, cholesterol, triglycerides, high density lipoprotein (HDL), and LDL levels had been noticed (127). The oral daily gavage with

**Table 2.** The different effects of PPP on the antioxidant, antimicrobial, immune status, and lipid metabolism of poultry

| Dose of PPP/diet        | Species                        | Findings   | Reference |
|-------------------------|--------------------------------|--|-----------|
| 0.01%                   | Broiler chickens               | Reduced <i>Bacillus cereus</i> and <i>S. aureus</i> in the retail meat products  | (86)      |
| 0.05%                   | Broiler chickens               | Decreased serum total cholesterol and LDL  | (120)     |
| 0.5 and 1.0%            | Broiler chickens               | Increased weights of the spleen and bursa of Fabricius<br>Increased serum concentration of IgA and IgG<br>Increased intestinal <i>Bacillus</i> spp.<br>Reduced <i>E. coli</i> and <i>Salmonella</i> spp.                 | (41)      |
| 20 g/kg                 | Broiler chickens               | Enhanced immune function<br>Proliferation of intestinal beneficial bacteria<br>Reduced pathogenic bacteria   | (36)      |
| 0.02%                   | Broiler chickens               | High serum SOD and glutathione peroxidation<br>No effect on glutamic oxaloacetic transaminase and glutamic pyruvic transaminase enzymes<br>Reduced lipid peroxidation of breast meat<br>Enhanced humoral immune response | (42)      |
| 100 mg/kg               | Heat-stressed broiler chickens | No significant increase in antibody titer to NDV vaccine<br>Reduced cholesterol level  | (116)     |
| 250 and 500 mg/kg       | Broiler chickens               | Increased glutathione peroxidation and SOD in liver<br>Reduced lipid peroxidation  | (44)      |
| 250, 450, and 650 mg/kg | Heat-stressed broiler chickens | Decreased MDA in the breast muscle<br>Increased water holding capacity of breast meat during storage<br>Increased IgG and anti-sheep RBCs titers<br>Reduced cholesterol and LDL<br>Increased HDL                         | (38)      |
| 7.5 and 10 g/kg         | Broiler chickens               | Upregulation of hepatic CAT and SOD<br>Reduced MD, triglycerides, cholesterol, and abdominal fat deposition<br>Increased nitric oxide  | (33)      |
| 4%                      | Laying hens                    | Reduced MDA<br>Enhanced antioxidative enzymes and total antioxidant blood capacity   | (27)      |
| 5-10 gm                 | Broiler chickens               | No effect on glutathione peroxidase, SOD, and CAT<br>Decreased MDA in breast meat<br>Reduced cecal <i>C. perfringens</i> count   | (25)      |
| 4% and 8%               | Broiler chickens               | Increased total antioxidant capacity, SOD, and glutathione peroxidase<br>Decreased MDA and LDL in meat<br>Increased IgG, IgM, and total immunoglobulins  | (22)      |
| 3% and 9%               | Quails                         | Improved antioxidant activity and greater expression of hepatic growth hormone receptors gene<br>Increased immune cells count  | (46)      |
| 4 g/kg                  | Broiler chickens               | Increased IgM  | (39)      |
| 4%                      | Broiler chickens               | Decreased peroxidase value in meat<br>High intestinal <i>Lactobacillus</i> count<br>Low intestinal coliform populations  | (23)      |
| 5% and 10%              | Layer hens                     | Decreased IgG  | (71)      |
| 400 ppm                 | Broiler chickens               | Decreases intestinal lesions<br>Decrease <i>Eimeria</i> oocysts count  | (87)      |

0.8 mL/kg pomegranate seed oil in rats resulted in a significant elevation of creatinine, urea, LDL, triglyceride, and glucose levels in serum and increasing of urine markers and MDA in tissue homogenates, but a

significant reduction in total thiol content and serum HDL (128). The study of Attia et al. (129) revealed that administration of pomegranate juice (3 mL/kg/day) and peel water extract (200 mg/kg/day) resulted in a significant reduction in serum glucose,

triglycerides, total cholesterol, LDL, urea, uric acid, and creatinine, however, HDL was elevated in diabetic and hypercholesterolemic rats. Besides, a significant increase in white blood cells and RBCs were detected in hypercholesterolemic rats treated with a pomegranate juice (129).

## CONCLUSION

Pomegranate is a widely known plant that shows promising effects as a natural feed additive in poultry production system. The plant and its by-products showed positive influences on the production of poultry. More in-depth research work on the effects of pomegranate by-products on poultry health and immunity must be carried out.

## CONFLICT OF INTEREST

The author declares that they have no potential conflict of interest with respect to the authorship and/or publication of this article.

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## AUTHORS' CONTRIBUTIONS

WAA has collected and drafted the manuscript, formatted it, and approved the final manuscript.

## REFERENCES

- Abd El-Ghany, W.A. (2020). Paraprobiotics and postbiotics: contemporary and promising natural antibiotics alternatives and their applications in the poultry field. *Open Vet J.* 10(3): 323-330. <https://doi.org/10.4314/ovj.v10i3.11> PMID:33282704 PMCID:PMC7703615
- Abd El-Ghany, W.A. (2020). Microalgae in poultry field: a comprehensive perspectives. *Adv Anim Vet Sci.* 8(9): 888-897. <https://doi.org/10.17582/journal.aavs/2020/8.9.888.897>
- Abd El-Ghany, W.A. (2021). Uses of immunoglobulins as an antimicrobials alternative in Veterinary Medicine. *World Vet J.* 11(1): 16-22. <https://doi.org/10.54203/scil.2021.wvj2>
- Abd El-Ghany, W.A., Abdel-Latif, M.A., Hosny, F., Alatfehy, N.M., Noreldin, A.E., Quesnell, R.R., Chapman, R., et al. (2022). Comparative efficacy of postbiotic, probiotic, and antibiotic against necrotic enteritis in broiler chickens. *Poult Sci.* 101(8): 101988. <https://doi.org/10.1016/j.psj.2022.101988> PMID:35809347 PMCID:PMC9272375
- Abd El Ghany, W.A., Fouad, H., Quesnell, R., Sakai, L. (2022). The effect of a postbiotic produced by stabilized non-viable Lactobacilli on the health, growth performance, immunity, and gut status of colisepticaemic broiler chickens. *Trop Anim Health Prod.* 54(5): 286. <https://doi.org/10.1007/s11250-022-03300-w> PMID:36083376 PMCID:PMC9463281
- Hady, M.M., Zaki, M.M., Abd El-Ghany, W.A., Korany, R.M.S. (2016). Assessment of the broilers performance, gut healthiness and carcass characteristics in response to dietary inclusion of dried coriander, turmeric and thyme. *Int J Environ Agric Res.* 2(6): 153-159.
- Zaki, M.M., Abd El-Ghany, W.A., Hady, M.M., Korany, R.M.S. (2016). Effect of certain phytobiotics on the immune response of Newcastle disease vaccinated broiler chickens. *Asian J Poult Sci.* 10(3): 134-140. <https://doi.org/10.3923/ajpsaj.2016.134.140>
- Abd El-Ghany, W.A. (2020). Phytobiotics in poultry industry as growth promoters, antimicrobials and immunomodulators - a review. *J World Poult Res.* 10(4): 571-579. <https://doi.org/10.36380/jwpr.2020.65>
- Abd El-Ghany, W.A., Eraky, R. (2020). Influence of dietary Moringa oleifera on broilers performance, intestinal microbial population and humoral immune competence. *J Hellenic Vet Med Soc.* 70(4): 1805-1810. <https://doi.org/10.12681/jhvms.22224>
- Abd El-Ghany, W.A. (2020). A Review on the use of Azolla species in poultry production. *J World Poult Res.* 10(2): 378-384. <https://doi.org/10.36380/jwpr.2020.44>
- Abd El-Ghany, W.A. (2022). The potential uses of silymarin, a milk thistle (*Silybum marianum*) derivative, in poultry production system. *Online J Anim Feed Res.* 12(1): 46-52. <https://doi.org/10.51227/ojaf.2022.7>
- Abd El-Ghany, W.A., Babazadeh, D. (2022). Betaine: a potential nutritional metabolite in the poultry industry. *Animals* 12(19): 2624. <https://doi.org/10.3390/ani12192624> PMID:36230366 PMCID:PMC9559486



13. Ross, I.A. (1999). Medicinal plants of world. Totowa, New Jersey: Humana press
14. Ercisli, S., Gadze, J., Agar, G., Yildirim, N., Hizarci, Y. (2011). Genetic relationships among wild pomegranate (*Punica granatum*) genotypes from Coruh Valley in Turkey. *Genet Mol Res.* 10(1): 459-464.  
<https://doi.org/10.4238/vol10-1gmr1155>  
PMid:21425096
15. Pienaar, L. (2021). The Economic contribution of South Africa's pomegranate industry. South Africa: Western Cape Department of Agriculture
16. Sharma, J., Ramchandra, K.K., Sharma, D., Meshram, D.T., Maity, A.N.N. (2014). Pomegranate: cultivation marketing and utilization.
17. Ismail, T., Sestili, P., Akhar, S. (2012). Pomegranate peel and fruit extracts: a review of potential anti-inflammatory and anti-infective effects. *J Ethnopharmacol.* 143(2): 397-405.  
<https://doi.org/10.1016/j.jep.2012.07.004>  
PMid:22820239
18. Naveena, B.M., Sen, A.R., Vaithyanathan, S., Babji, Y., Kondaiah, N. (2008). Comparative efficacy of pomegranate juice, pomegranate rind powder extract and BHT as antioxidants in cooked chicken patties. *Meat Sci.* 80(4): 1304-1308.  
<https://doi.org/10.1016/j.meatsci.2008.06.005>  
PMid:22063872
19. Kandyliis, P., Kokkinomagoulos, E. (2020). Food applications and potential health benefits of pomegranate and its derivatives. *Foods.* 9(2): 122.  
<https://doi.org/10.3390/foods9020122>  
PMid:31979390 PMCid:PMC7074153
20. Akhtar, S., Ismail, T., Fraternal, D., Sestili, P. (2015). Pomegranate peel and peel extracts: Chemistry and food features. *Food Chem.* 174, 417-425.  
<https://doi.org/10.1016/j.foodchem.2014.11.035>  
PMid:25529700
21. Ahmed, S.T., Islam, M.M., Bostami, A.B., Mun, H.S., Kim, Y.J., Yang, C.J. (2015). Meat composition, fatty acid profile and oxidative stability of meat from broilers supplemented with pomegranate (*Punica granatum* L.) by products. *Food Chem.* 188(1-2): 481-488.  
<https://doi.org/10.1016/j.foodchem.2015.04.140>  
PMid:26041221
22. Ghasemi-Sadabadi, M., Ebrahimnezhad, Y., Maheri-Sis, N., Ghalehkandi, J.G., Shaddel-Teli, A. (2021). Immune response and antioxidant status of broilers as influenced by oxidized vegetable oil and pomegranate peel. *J Anim Sci Technol.* 63(5): 1034-1063.  
<https://doi.org/10.5187/jast.2021.e99>  
PMid:34796346 PMCid:PMC8564296
23. Ghasemi Sadabadi, M., Ebrahimnezhad, Y., Maheri-Sis, N., Shaddel-Teli, A., Ghalehkandi J.G., Veldkamp, T. (2022). Effects of supplementation of pomegranate processing by products and waste cooking oils as alternative feed resources in broiler nutrition. *Sci Rep.* 12, 21216.  
<https://doi.org/10.1038/s41598-022-25761-7>  
PMid:36481691 PMCid:PMC9731951
24. Reddy, M.K., Gupta, S.K., Jacob, M.R., Khan, S.I., Ferreira, D. (2007). Antioxidant, antimalarial and antimicrobial activities of tannin-rich fractions, ellagitannins and phenolic acids from *Punica granatum* L. *Planta Med.* 73(5): 461-467.  
<https://doi.org/10.1055/s-2007-967167>  
PMid:17566148
25. Ghasemi-Sadabadi, M., Ebrahimnezhad, Y., Maheri-Sis, N., Shaddel Teli, A., Ghiasi Ghalehkandi, J., Veldkamp, T. (2021). Supplementation of pomegranate processing waste and waste soybean cooking oil as an alternative feed resource with vitamin E in broiler nutrition: Effects on productive performance, meat quality and meat fatty acid composition. *Arch Anim Nutr.* 75(5): 355-375.  
<https://doi.org/10.1080/1745039X.2021.1965414>  
PMid:34461782
26. Elsebai, A., El-Tahawy, W.S., El-Attar, F.M., ELnaggar, A.S. (2022). Effect of dietary inclusion of pomegranate peel powder (*Punica granatum*) on growth performance and some physiological parameters of broiler chicks. *J Agric Environ Sci.* 21(2): 19-44.  
<https://doi.org/10.21608/jaesj.2022.159602.1017>
27. Eid, Y., Kirrella, A.A., Tolba, A., El-Deeb, M., Sayed, S., El-Sawy, H.B., Shukry, M., Dawood, M.A.O. (2021). Dietary pomegranate by-product alleviated the oxidative stress induced by dexamethasone in laying hens in the pre-peak period. *Animals* 11(4): 1022.  
<https://doi.org/10.3390/ani11041022>  
PMid:33916329 PMCid:PMC8066172
28. Vasilopoulos, S., Dokou, S., Papadopoulos, G.A., Savvidou, S., Christaki, S., Kyriakoudi, A., Dots, V., et al. (2022). Dietary supplementation with pomegranate and onion aqueous and cyclodextrin encapsulated extracts affects broiler performance parameters, welfare and meat characteristics. *Poultry* 1(2): 74-93.  
<https://doi.org/10.3390/poultry1020008>
29. Kishawy, A.T., Omar, A.E., Gomaa, A.M. (2018). Growth performance and immunity of broilers fed rancid oil diets that supplemented with pomegranate peel extract and sage oil. *Jpn J Vet Res.* 64(Suppl 2): S31-S38.

30. Akarca, G., Başpınar, E. (2019). Determination of pomegranate peel and seed extracted in different solvents for antimicrobial effect. *Turk J Agric Food Sci Technol.* 7(1): 46-53.
31. Benchagra, L., Berrougui, H., Islam, M.O., Ramchoun, M., Boulbaroud, S., Hajjaji, A., Fulop, T., et al. (2021). Antioxidant effect of Moroccan pomegranate (*Punica granatum* l. sefri variety) extracts rich in punicalagin against the oxidative stress process. *Foods* 10(9): 2219.  
<https://doi.org/10.3390/foods10092219>  
PMid:34574329 PMCID:PMC8469689
32. Ahmadipour, B., Pat, S., Khajali, F. (2018). The protective effect of pomegranate peel powder on pulmonary hypertension in broiler chickens. *JSM Biomarkers.* 4(1): 1013.
33. Ahmadipour, B., Pat, S., Abaszadeh, S., Hassanpour, H., Khajali, F. (2021). Pomegranate peel as a phytogetic in broiler chickens: Influence upon antioxidant, lipogenesis and hypotensive response. *Vet Med Sci.* 7(5): 1907-1913.  
<https://doi.org/10.1002/vms3.556>  
PMid:34132060 PMCID:PMC8464295
34. Tito, A., Colantuono, A., Pirone, L., Pedone, E., Intartaglia, D., Giamundo, G., Conte, I., et al. (2021). Pomegranate peel extract as an inhibitor of SARS-CoV-2 spike binding to human ACE2 receptor (in vitro): a promising source of novel antiviral drugs. *Front Chem.* 9, 638187.  
<https://doi.org/10.3389/fchem.2021.638187>  
PMid:33996744 PMCID:PMC8114579
35. Hamady, G.A., Abdel-Moneim, M.A., El-Chaghaby, G.A., Abd-El-Ghany, Z.M., Hassanin, M.S. (2015). Effect of pomegranate peel extract as natural growth promoter on the productive performance and intestinal microbiota of broiler chickens. *Afr J Agric Sci Technol.* 3(12): 514-519.
36. Rezvani, M.R., Rahimi, S. (2017). Effects of adding pomegranate peel extract and commercial antioxidant to diets on performance, nutrient digestibility, gastrointestinal micro flora and antibody titer of broilers. *J Vet Res.* 72(2): 147-156.
37. Kishawy, A.T., Amer, S.A., Abd El-Hack, M.E., Saadeldin, I.M., Swelum, A.A. (2019). The impact of dietary linseed oil and pomegranate peel extract on broiler growth, carcass traits, serum lipid profile, and meat fatty acid, phenol, and flavonoid contents. *Asian-Australas J Anim Sci.* 32(8): 1161-1171.  
<https://doi.org/10.5713/ajas.18.0522>  
PMid:30744351 PMCID:PMC6599952
38. Sharifian, M., Hosseini-Vashan, S.J., Nasri, M.F., Perai, A.H. (2019). Pomegranate peel extract for broiler chickens under heat stress: Its influence on growth performance, carcass traits, blood metabolites, immunity, jejunal morphology, and meat quality. *Livest Sci.* 227, 22-28.  
<https://doi.org/10.1016/j.livsci.2019.06.021>
39. Abdel Baset, S., Ashour, E.A., Abd El-Hack, M.E., El-Mekkawy, M.M. (2022). Effect of different levels of pomegranate peel powder and probiotic supplementation on growth, carcass traits, blood serum metabolites, antioxidant status and meat quality of broilers. *Anim Biotechnol.* 33(4): 690-700.  
<https://doi.org/10.1080/10495398.2020.1825965>  
PMid:33000991
40. Rajani, J., Karimi Torshizi, M.A., Rahimi, S. (2011). Control of ascites mortality and improved performance and meat shelf-life in broilers using feed adjuncts with presumed antioxidant activity. *Anim Feed Sci Tech.* 170(3-4): 239-245.  
<https://doi.org/10.1016/j.anifeedsci.2011.09.001>
41. Ahmed, S.T., Yang, C.J. (2017). Effects of dietary *Punica granatum* L. by-products on performance, immunity, intestinal and fecal microbiology, and odorous gas emissions from excreta in broilers. *J Poult Sci.* 54(2): 157-166.  
<https://doi.org/10.2141/jpsa.0160116>  
PMid:32908421 PMCID:PMC7477121
42. Saleh, H., Golian, A., Kermanshahi, H., Mirakzchi, M.T. (2017). Effects of dietary  $\alpha$ -tocopherol acetate, pomegranate peel, and pomegranate peel extract on phenolic content, fatty acid composition, and meat quality of broiler chickens. *J Appl Anim Res.* 45(1): 629-636.  
<https://doi.org/10.1080/09712119.2016.1248841>
43. Saleh, H., Golian, A., Kermanshahi, H., Mirakzchi, M.T. (2018). Antioxidant status and thigh meat quality of broiler chickens fed diet supplemented with  $\alpha$ -tocopherolacetate, pomegranate pomace and pomegranate pomace extract. *Ital J Anim Sci.* 17(2): 386-395.  
<https://doi.org/10.1080/1828051X.2017.1362966>
44. Rao, S.V.R., Raju, M.V.L.N., Prakash, B., Rajkumar, U., Reddy, E.P.K. (2019). Effect of supplementing moringa (*Moringa oleifera*) leaf meal and pomegranate (*Punica granatum*) peel meal on performance, carcass attributes, immune and antioxidant responses in broiler chickens. *Anim Prod Sci.* 59(2): 288-294.  
<https://doi.org/10.1071/AN17390>

45. Akuru, E.A., Mpendulo, C.T., Oyeagu, C.E., Nantapo, C.W.T. (2021). Pomegranate (*Punica granatum* L.) peel powder meal supplementation in broilers: effect on growth performance, digestibility, carcass and organ weights, serum and some meat antioxidant enzyme biomarkers. *Ital J Anim Sci.* 20(1): 119-131.  
<https://doi.org/10.1080/1828051X.2020.1870877>
46. Kamel, E.R., Shafk, B.M., Mamdouh, M., Elrafaay, S., Abdelfattah, F.A.I. (2021). Response of two strains of growing Japanese quail (*Coturnix Coturnix Japonica*) to diet containing pomegranate peel powder. *Trop Anim Health Prod.* 53(6): 549.  
<https://doi.org/10.1007/s11250-021-02987-7>  
PMid:34782923
47. Mohamed, T.M., Sun, W., Bumbie, G.Z., Dosoky, W.M., Rao, Z., Hu, P., Wu, L., Tang, Z. (2022). Effect of dietary supplementation of *Bacillus subtilis* on growth performance, organ weight, digestive enzyme activities, and serum biochemical indices in broiler. *Animals* 12(12): 1558.  
<https://doi.org/10.3390/ani12121558>  
PMid:35739895 PMCID:PMC9219452
48. Qnais, E.Y., Elokda, A.S., Abu Ghalyun, Y.Y., Abdulla, F.A. (2007). Antidiarrheal activity of the aqueous extract of *Punica granatum* (Pomegranate) peels. *Pharm Biol.* 45(9): 715-720.  
<https://doi.org/10.1080/13880200701575304>
49. Reddy, B.U., Mullick, R., Kumar, A., Sudha, G., Srinivasan, N., Das, S. (2014). Small molecule inhibitors of HCV replication from pomegranate. *Sci Rep.* 4, 5411.  
<https://doi.org/10.1038/srep05411>  
PMid:24958333 PMCID:PMC4067622
50. Perricone, V., Comi, M., Giromini, C., Rebucci, R., Agazzi, A., Savoini, G., Bontempo, V. (2020). Green tea and pomegranate extract administered during critical moments of the production cycle improves blood antiradical activity and alters cecal microbial ecology of broiler chickens. *Animals (Basel).* 10(5): 785.  
<https://doi.org/10.3390/ani10050785>  
PMid:32366030 PMCID:PMC7277556
51. Middha, S.K., Usha, T., Pande, V. (2013). HPLC evaluation of phenolic profile, nutritive content, and antioxidant capacity of extracts obtained from *Punica granatum* fruit peel. *Adv Pharmacol Sci.* 2013, 296236.  
<https://doi.org/10.1155/2013/296236>  
PMid:23983682 PMCID:PMC3747345
52. Saeed, M., Naveed, M., Bibi, J., Kamboh, A.A., Arain, M.A., Shah, Q.A., Alagawany, M., et al. (2018). The promising pharmacological effects and therapeutic/medicinal applications of *Punica Granatum* L. (Pomegranate) as a functional food in humans and animals: a review. *Recent Pat Inflamm Allergy Drug Discov.* 12(1): 24-38.  
<https://doi.org/10.2174/1872213X12666180221154713>  
PMid:29473532
53. Cho, J.H., Kim, H.J., Kim, I.H. (2014). Effects of phytogetic feed additive on growth performance, digestibility, blood metabolites, intestinal microbiota, meat color and relative organ weight after oral challenge with *Clostridium perfringens* in broilers. *Livest Sci.* 160, 82-88.  
<https://doi.org/10.1016/j.livsci.2013.11.006>
54. Bostami, A.B.M.R., Ahmed, S.T., Islam, M.M. (2015). Growth performance, fecal noxious gas emission and economic efficacy in broilers fed fermented pomegranate byproducts as residue of fruit industry. *Int J Adv Res.* 3(3): 102-114.
55. Pascariu, S.M., Pop, I.M., Simeanu, D., Pavel, G., Solcan, C. (2017). Effects of wine by-products on growth performance, complete blood count and total antioxidant status in broilers. *Rev Bras Cienc Avic.* 19(2): 191-202.  
<https://doi.org/10.1590/1806-9061-2016-0305>
56. Goni, I., Brenes, A., Centeno, C., Viveros, A., Saura-Calixto, F., Rebolé, A., Arija, I., Estevez, R. (2007). Effect of dietary grape pomace and vitamin E on growth performance, nutrient digestibility, and susceptibility to meat lipid oxidation in chickens. *Poult Sci.* 86(3): 508-516.  
<https://doi.org/10.1093/ps/86.3.508>  
PMid:17297163
57. Marzo, F., Urdaneta, E., Santidrian, S. (2002). Liver proteolytic activity in tannic acid-fed birds. *Poult Sci.* 81(1): 92-94.  
<https://doi.org/10.1093/ps/81.1.92>  
PMid:11885906
58. Bravo, L., Abia, R., Eastwood, M.A., Saura-Calixtol, F. (1994). Degradation of polyphenols (catechin and tannic acid) in the rat intestinal tract. Effect on colic fermentation and faecal output. *Br J Nutr.* 71(6): 933-946.  
<https://doi.org/10.1079/BJN19940197>  
PMid:8031740
59. Saleh, H. (2015). Effects of natural antioxidant on the immune response, antioxidant enzymes and hematological broilers chickens. *Iran Vet J.* 11(3): 67-79.

60. Banaszkiwicz, T., Białek, A., Tokarz, A., Kaszperuk, K. (2018). Effect of dietary grape and pomegranate seed oil on the post-slaughter value and physicochemical properties of muscles of broiler chickens. *Acta Sci Pol Technol Aliment.* 17(3): 199-209. <https://doi.org/10.17306/J.AFS.0563> PMID:30269459
61. Hamad, K.M., Kareem, K.Y. (2019). Growth performance, carcass quality, and economics of production of Japanese quails fed with pomegranate peel powder. *Int J Eng Technol Manag Appl Sci.* 10(14): 10A14L.
62. Akuru, E.A., Oyeagu, C.E., Mpendulo, T.C., Rautenbach, F., Oguntibeju, O.O. (2020). Effect of pomegranate (*Punica granatum* L) peel powder meal dietary supplementation on antioxidant status and quality of breast meat in broilers. *Heliyon* 6(12): e05709. <https://doi.org/10.1016/j.heliyon.2020.e05709> PMID:33364487 PMCid:PMC7750561
63. Murugesan, G.R., Syed, B., Haldar, S., Pender, C. (2015). Corrigendum: phytogenic feed additives as an alternative to antibiotic growth promoters in broiler chickens. *Front Vet Sci.* 2, 21. <https://doi.org/10.3389/fvets.2015.00037>
64. Al-Shammari, K.I.A., Batkowska, J., Zamil, S.J. (2019). Role of pomegranate peels and black pepper powder and their mixture in alleviating the oxidative stress in broiler chickens. *Int J Poult Sci.* 18(3): 122-128. <https://doi.org/10.3923/ijps.2019.122.128>
65. Szymczyk, B., Szczurek, W. (2016). Effect of dietary pomegranate seed oil and linseed oil on broiler chickens performance and meat fatty acid profile. *J Anim Feed Sci.* 25(1): 37-44. <https://doi.org/10.22358/jafs/65585/2016>
66. Rafiei, F., Khajali, F. (2021). Flavonoid antioxidants in chicken meat production: Potential application and future trends. *World Poult Sci J.* 77(2): 347-361. <https://doi.org/10.1080/00439339.2021.1891401>
67. Saki, A.A., Rabet, M., Zamani, P., Yousefi, A. (2014). The effects of different levels of pomegranate seed pulp with multi-enzyme on performance, egg quality and serum antioxidant in laying hens. *Iran J Appl Anim Sci.* 4(4): 803-808.
68. Abbas, R.J., Al-Salhi, K.C.K., Al-Hummod, S.K. (2017). The effect of using different levels of pomegranate (*Punica granatum*) peel powder on productive and physiological performance of Japanese quail (*Coturnix coturnix japonica*). *Livest Res Rural Dev.* 29(12): 231.
69. Ghahtan, N., Kohanmoo, M.A., Habibi, H. (2019). Evaluation of dietary medicinal plants and algae in laying Japanese quails. *J World Poult Res.* 9(2): 82-88. <https://doi.org/10.36380/jwpr.2019.10>
70. Mutlu, S.I., Güler, T. (2021). The effect of ellagic acid on performance, digestibility, egg quality, cecal bacterial flora, antioxidant activity, and some blood parameters in laying quails reared at different temperatures. *Turk J Vet Anim Sci.* 45(1): 101-112. <https://doi.org/10.3906/vet-2002-58>
71. Gulpe, E.E., Iqbal, A., Cetingul, I.S., Uyarlar, C., Ozcinar, U., Bayram, I. (2022). Effects of pomegranate (*Punica granatum* L.) juice as a short-term water supplement during the peak production cycle in laying hens. *Ankara Univ Vet Fak Derg.* 69, 241-249. <https://doi.org/10.33988/auvfd.795175>
72. Iqbal, A., Bayram, I., Gulpe, E.E., Uyarlar, C., Ümit, Ö., Cetingul İ.S. (2021). The Effect of different level of Pomegranate molasses on performance, egg quality trait, serological and hematological parameters in older laying hens: the use of pomegranate molasses as growth promoter in poultry nutrition. *Progr Nutr.* 22(4): e2020052.
73. Sharma, M.K., Dinh, T., Adhikari, P.A. (2020). Production performance, egg quality, and small intestine histomorphology of the laying hens supplemented with phytogenic feed additive. *J Appl Poult Res.* 29(2): 362-371. <https://doi.org/10.1016/j.japr.2019.12.001>
74. Kostogrys, R.B., Filipiak-Florkiewicz, A., Dereń, K., Drahun, A., Czyżyńska-Cichoń, I., Cieślik, E., Szymczyk, B., Franczyk-Żarów, M. (2017). Effect of dietary pomegranate seed oil on laying hen performance and physicochemical properties of eggs. *Food Chem.* 221, 1096-1103. <https://doi.org/10.1016/j.foodchem.2016.11.051> PMID:27979064
75. Johanningsmeier, S.D., Harris, G.K. (2011). Pomegranate as a functional food and nutraceutical source. *Annu Rev Food Sci Technol.* 2: 181-201. <https://doi.org/10.1146/annurev-food-030810-153709> PMID:22129380
76. Seeram, N.P., Adams, L.S., Henning, S.M., Niu, Y., Zhang, Y., Nair, M.G., Heber, D. (2005). In vitro antiproliferative, apoptotic and antioxidant activities of punicalagin, ellagic acid and a total pomegranate tannin extract are enhanced in combination with other polyphenols as found in pomegranate juice. *J Nutr Biochem.* 16(6): 360-367. <https://doi.org/10.1016/j.jnutbio.2005.01.006> PMID:15936648



77. Singh, B., Singh, J.P., Kaur, A., Singh, N. (2018). Phenolic compounds as beneficial phytochemicals in pomegranate (*Punica granatum* L.) peel: a review. *Food Chem.* 261, 75-86.  
<https://doi.org/10.1016/j.foodchem.2018.04.039>  
PMid:29739608
78. Ghosh, S., Chatterjee, P.N., Maity, A., Mukherjee, J., Batabyal, S., Chatterjee, J.K. (2020). Effect of supplementing pomegranate peel infusion on body growth, feed efficiency, biochemical metabolites and antioxidant status of broiler chicken. *Trop Anim Health Prod.* 52(6): 3899-3905.  
<https://doi.org/10.1007/s11250-020-02352-0>  
PMid:32737663
79. Ginsburg, I., Kohen, R., Koren, E. (2011). Microbial and host cells acquire enhanced oxidant-scavenging abilities by binding polyphenols. *Arch Biochem Biophys.* 506(1): 12-23.  
<https://doi.org/10.1016/j.abb.2010.11.009>  
PMid:21081104
80. Faria, A., Monteiro, R., Mateus, N., Azevedo, I., Calhau, C. (2007). Effect of pomegranate (*Punica granatum*) juice intake on hepatic oxidative stress. *Eur J Nutr.* 46(5): 271-278.  
<https://doi.org/10.1007/s00394-007-0661-z>  
PMid:17514376
81. Gil, M.I., Tomás-Barberán, F.A., Hess-Pierce, B., Holcroft, D.M., Kader, A.A. (2000). Antioxidant activity of pomegranate juice and its relationship with phenolic composition and processing. *J Agric Food Chem.* 48(10): 4581-4589.  
<https://doi.org/10.1021/jf000404a>  
PMid:11052704
82. Gözlekçi, S., Saraçoğlu, O., Onursal, E., Ozgen, M. (2011). Total phenolic distribution of juice, peel, and seed extracts of four pomegranate cultivars. *Pharmacogn Mag.* 7(26): 161-164.  
<https://doi.org/10.4103/0973-1296.80681>  
PMid:21716925 PMCID:PMC3113357
83. Benchagra, L., Berrougui, H., Islam, M.O., Ramchoun, M., Boulbaroud, S., Hajjaji, A., Fulop, T., et al. (2021). Antioxidant effect of Moroccan pomegranate (*Punica granatum* L. Sefri Variety) extracts rich in punicalagin against the oxidative stress process. *Foods.* 10(9): 2219.  
<https://doi.org/10.3390/foods10092219>  
PMid:34574329 PMCID:PMC8469689
84. Afaq, F., Saleem, M., Krueger, C.G., Reed, J.D., Mukhtar, H. (2005). Anthocyanin and hydrolyzable tannin-rich pomegranate fruit extract modulates MAPK and NF- $\kappa$ B pathways and inhibits skin tumorigenesis in CD-1 mice. *Int J Cancer.* 113(3): 423-433.  
<https://doi.org/10.1002/ijc.20587>  
PMid:15455341
85. Singh, J.P., Kaur, A., Shevkani, K., Singh, N. (2016). Composition, bioactive compounds, and antioxidant activity of common Indian fruits and vegetables. *J Food Sci Technol.* 53(11): 4056-4066.  
<https://doi.org/10.1007/s13197-016-2412-8>  
PMid:28035161 PMCID:PMC5156649
86. Kanatt, S.R., Chander, R., Sharma, A. (2010). Antioxidant and antimicrobial activity of pomegranate peel extract improves the shelf life of chicken products. *Int J Food Sci Technol.* 45(2): 216-222.  
<https://doi.org/10.1111/j.1365-2621.2009.02124.x>
87. Khorrami, P., Gholami-Ahangaran, M., Moghtadaei-Khorasgani, E. (2022). The efficacy of pomegranate peel extract on *Eimeria* shedding and growth indices in experimental coccidiosis in broiler chickens. *Vet Med Sci.* 8(2): 635-641.  
<https://doi.org/10.1002/vms3.714>  
PMid:34981905 PMCID:PMC8959253
88. Dahham, S.S., Ali, M.N., Tabassum, H., Khan, M. (2010). Studies on antibacterial and antifungal activity of pomegranate (*Punica granatum* L.). *Am-Eurasian J Agric Environ Sci.* 9(3): 273-281.
89. Growther, L., Sukirtha, K., Savitha, N., Niren, A.S. (2012). Antibacterial activity of *Punica granatum* peel extracts against shiga toxin producing *E. coli*. *Int J Life Sci Biotechnol Pharma Res.* 1(4): 164-172.
90. Prashanth, D.J., Asha, M.K., Amit, A. (2001). Antibacterial activity of *Punica granatum*. *Fitoterapia* 72(2): 171-173.  
[https://doi.org/10.1016/S0367-326X\(00\)00270-7](https://doi.org/10.1016/S0367-326X(00)00270-7)  
PMid:11223228
91. Voravuthikunchai, S.P., Sririrak, T., Limsuwan, S., Supawita, T., Iida, T., Honda, T. (2005). Inhibitory effects of active compounds from *Punica granatum* pericarp on verocytotoxin production by enterohemorrhagic *Escherichia coli* O157:H7. *J Health Sci.* 51(5): 590-596.  
<https://doi.org/10.1248/jhs.51.590>
92. McCarrell, E.M., Gould, S.W., Fielder, M.D., Kelly, A.F., El Sankary, W., Naughton, D.P. (2008). Antimicrobial activities of pomegranate rind extracts: enhancement by addition of metal salts and vitamin C. *BMC Complement Altern Med.* 8, 64.  
<https://doi.org/10.1186/1472-6882-8-64>  
PMid:19077299 PMCID:PMC2628863
93. Nuamsetti, T., Dechayuenyong, P., Tantipaibulvut, S. (2012). Antibacterial activity of pomegranate fruit peels and arils. *Sci Asia.* 38, 319-322.  
<https://doi.org/10.2306/scienceasia1513-1874.2012.38.319>



94. Juneja, V.K., Cadavez, V., Gonzales-Barron, U., Mukhopadhyay, S., Friedman, M. (2016). Effect of pomegranate powder on the heat inactivation of *Escherichia coli* O104:H4 in ground chicken. *Food Control*. 70, 26-34.  
<https://doi.org/10.1016/j.foodcont.2016.05.027>
95. Pagliarulo, C., De Vito, V., Picariello, G., Colicchio, R., Pastore, G., Salvatore, P., Volpe, M.G. (2016). Inhibitory effect of pomegranate (*Punica granatum* L.) polyphenol extracts on the bacterial growth and survival of clinical isolates of pathogenic *Staphylococcus aureus* and *Escherichia coli*. *Food Chem*. 190, 824-831.  
<https://doi.org/10.1016/j.foodchem.2015.06.028>  
PMid:26213044
96. Wafa, B.A., Makni, M., Ammar, S., Khannous, L., Hassana, A.B., Bouaziz, M., Es-Safi, N.E., Gdoura, R. (2017). Antimicrobial effect of the Tunisian Nana variety *Punica granatum* L. extracts against *Salmonella enterica* (serovars Kentucky and Enteritidis) isolated from chicken meat and phenolic composition of its peel extract. *Int J Food Microbiol*. 241, 123-131.  
<https://doi.org/10.1016/j.ijfoodmicro.2016.10.007>  
PMid:27776287
97. Abdollahzadeh, S.H., Mashouf, R.Y., Mortazavi, H., Moghaddam, M.H., Roozbahani, N., Vahedi, M. (2011). Antibacterial and antifungal activities of *Punica granatum* peel extracts against oral pathogens. *J Dent (Tehran)* 8(1): 1-6.
98. Oraki, H.H., Demirci, A.S., Gümü, S.T. (2011). Antibacterial and antifungal activity of pomegranate (*Punica granatum* L. cv.) peel. *Elect J Env Agricult Food Chem*. 10(3): 1958-1969.
99. Singh, B., Singh, J.P., Kaur, A., Singh, N. (2019). Antimicrobial potential of pomegranate peel: a review. *Int J Food Sci Technol*. 54(4): 959-965.  
<https://doi.org/10.1111/ijfs.13964>
100. Scalbert, A. (1991). Antimicrobial properties of tannins. *Phytochemistry* 30(12): 3875-3883.  
[https://doi.org/10.1016/0031-9422\(91\)83426-L](https://doi.org/10.1016/0031-9422(91)83426-L)
101. Viuda-Martos, M., Fernández-López, J., Pérez-Álvarez, J.A. (2010). Pomegranate and its many functional components as related to human health: a review. *Compr Rev Food Sci Food Saf*. 9(6): 635-654.  
<https://doi.org/10.1111/j.1541-4337.2010.00131.x>  
PMid:33467822
102. Vidanarachchi, J.K., Mikkelsen, L., Sims, I., Iji, P., Choct, M. (2005). Phytobiotics: Alternatives to antibiotic growth promoters in monogastric animal feeds. *Recent Adv Anim Nutr Australia*. 15, 131-144.
103. Ferreira, S.B., Palmeira, J.D., Souza, J.H., Almeida, J.M., Figueiredo, M.C.P., Pequeno, A.S., Arruda, T.A., et al. (2010). Evaluation of the antimicrobial activity in vitro of the hydroalcoholic extract *Stryphnodendron adstringens* against of *Staphylococcus aureus* strains. *Braz J Clin Anal* 42, 27-31.
104. Tang, Y., Zhang, X., Wang, Y., Guo, Y., Zhu, P., Li, G., Zhang, J., et al. (2022). Correction: Dietary ellagic acid ameliorated *Clostridium perfringens*-induced subclinical necrotic enteritis in broilers via regulating inflammation and cecal microbiota. *J Anim Sci Biotechnol*. 13(1): 66.  
<https://doi.org/10.1186/s40104-022-00724-0>  
PMid:35562813 PMCid:PMC9103450
105. Sarica, S., Urkmez, D. (2016). The use of grape seed-, olive leaf-and pomegranate peel-extracts as alternative natural antimicrobial feed additives in broiler diets. *Europ Poult Sci*. 80, 1-13.  
<https://doi.org/10.1399/eps.2016.121>
106. Fahmy, Z.H., El-Shennawy, A.M., El-Komy, W., Ali, E., Hamid, S.S.A. (2009). Potential antiparasitic activity of pomegranate extracts against *Schistosomules schis* and mature worms of *Schistosoma mansoni*: in vitro and in vivo study. *Aust J Basic Appl Sci*. 3(4): 4634-4643.
107. El-Sherbini, G.T., Shoukry, N.M. (2012). In vitro effect of pomegranate peel extract on *Trichomonas tenax*. *Life Sci J*. 9(3): 791-797.
108. Al-Mathal, E.M., Alsalem, A.A. (2013). Pomegranate (*Punica granatum*) peel is effective in a murine model of experimental *Cryptosporidium parvum* ultrastructural studies of the ileum. *Exp Parasitol*. 134(4): 482-494.  
<https://doi.org/10.1016/j.exppara.2013.05.004>  
PMid:23684569
109. Boonmasawai, S., Sungpradit, S., Jirapattharasate, C., Nakthong, C., Piasai, L. (2013). Effects of alcoholic extract from pomegranate (*Punica granatum* L.) peels on gastrointestinal nematode egg counts in doe. *J Appl Anim Sci*. 6(2): 27-37.
110. Bunviboolvat, P., Taechaarpornkul, N., Saratham, J., Sungpradit, S., Jirapattharasate, C., Nakthong, C., Piasai, L., et al. (2013). Anthelmintic effects of ethanolic extracts from pomegranate peels, mangosteen peels and tamarind seeds on gastrointestinal nematode egg counts in lambs. *J Appl Anim Sci*. 6(2): 39-48.
111. Al-Megrin, W. (2016). Efficacy of pomegranate (*Punica granatum*) peel extract against *Hymenolepis nana* in infections mice. *Biosci Biotechnol Res Asia*. 13(1): 103-108.  
<https://doi.org/10.13005/bbra/2010>

112. Al-Megrin, W.A. (2017). In vivo study of pomegranate (*Punica granatum*) peel extract efficacy against *Giardia lamblia* in infected experimental mice. *Asian Pac J Trop Biomed.* 7(1): 59-63.  
<https://doi.org/10.1016/j.apjtb.2016.08.018>
113. Elfalleh, W., Hannachi, H., Tlili, N., Yahia, Y., Nasri, N., Ferchichi, A. (2012). Total phenolic contents and antioxidant activities of pomegranate peel, seed, leaf and flower. *J Med Plant Res.* 6(32): 4724-4730.  
<https://doi.org/10.5897/JMPR11.995>
114. Bazargani-Gilani, B., Aliakbarlu, J., Tajik, H. (2015). Effect of pomegranate juice dip ping and chitosan coating enriched with *Zataria multiflora* Boiss essential oil on the shelf-life of chicken meat during refrigerated storage. *Innov Food Sci Emerg Technol.* 29, 280-287.  
<https://doi.org/10.1016/j.ifset.2015.04.007>
115. Hayrapetyan, H., Hazeleger, W.C., Beumer, R.R. (2012). Inhibition of *Listeria monocytogenes* by pomegranate (*Punica granatum*) peel extract in meat paté at different temperatures. *Food Control.* 23(1): 66-72.  
<https://doi.org/10.1016/j.foodcont.2011.06.012>
116. Kumar, K.P., Reddy, V.R., Prakash, M.G. (2018). Effect of supplementing pomegranate (*Punica granatum*) peel extract on serum biochemical parameters and immune response in broilers during summer. *Pharma Innov J.* 7(3): 597-601.
117. Fadavi, A., Barzegar, M., Azizi, M.H. (2006). Determination of fatty acids and total lipid content in oilseed of 25 pomegranates varieties grown in Iran. *J Food Compos Anal.* 19(6-7): 676-680.  
<https://doi.org/10.1016/j.jfca.2004.09.002>
118. Melo, I.L.P., Carvalho, E.B.T., Mancini-Filho, J. (2014). Pomegranate seed oil (*Punica Granatum* L.): a source of punicic acid (Conjugated  $\alpha$ -Linolenic Acid). *J Hum Nutr Food Sci.* 2(1): 1024.
119. Wang, Y.W., Field, C.J., Sim, J.S. (2000). Dietary polyunsaturated fatty acids alter lymphocyte subset proportion and proliferation, serum immunoglobulin g concentration, and immune tissue development in chicks. *Poult Sci.* 79(12): 1741-1748.  
<https://doi.org/10.1093/ps/79.12.1741>  
PMid:11194036
120. Yaseen, A.T., El-Kholy, M.E.S.H., El-Razik, W.M.A., Soliman, M.H. (2016). Effect of using pomegranate peel extract as feed additive on performance, serum lipids and immunity of broiler chicks. *Zagazig Vet J.* 42(1): 87-92.  
<https://doi.org/10.21608/zvzj.2014.59473>
121. Hou, C., Zhang, W., Li, J., Du, L., Lv, O., Zhao, S., Li, J. (2019). Beneficial effects of pomegranate on lipid metabolism in metabolic disorders. *Mol Nutr Food Res.* 63(16): e1800773.  
<https://doi.org/10.1002/mnfr.201800773>  
PMid:30677224
122. Aviram, M., Fuhrman, B. (2001). Flavonoid antioxidants protect LDL from oxidation and attenuate atherosclerosis. *Curr Opin Lipidol.* 12(1): 41-48.  
<https://doi.org/10.1097/00041433-200102000-00008>  
PMid:11176202
123. Esmailizadeh, A., Tahbaz, F., Gaieni, I., Alavi-Majd, H., Azadbakht, L. (2006). Cholesterol-lowering effect of concentrated pomegranate juice consumption in type II diabetic patients with hyperlipidemia. *Int J Vitam Nutr Res.* 76(3): 147-151.  
<https://doi.org/10.1024/0300-9831.76.3.147>  
PMid:17048194
124. Lv, O., Wang, L., Li, J., Ma, Q., Zhao, W. (2016). Effects of pomegranate peel polyphenols on lipid accumulation and cholesterol metabolic transformation in L-02 human hepatic cells via the PPAR $\gamma$ -ABCA1/CYP7A1 pathway. *Food Funct.* 7(12): 4976-4983.  
<https://doi.org/10.1039/C6FO01261B>  
PMid:27845788
125. Zhao, W., Li, J., He, X., Lv, O., Cheng, Y., Liu, R. (2014). In vitro steatosis hepatic cell model to compare the lipid-lowering effects of pomegranate peel polyphenols with several other plant polyphenols as well as its related cholesterol efflux mechanisms. *Toxicol Rep.* 1, 945-954.  
<https://doi.org/10.1016/j.toxrep.2014.10.013>  
PMid:28962306 PMCid:PMC5598384
126. Wu, S., Tian, L. (2017). Diverse phytochemicals and bioactivities in the ancient fruit and modern functional food pomegranate (*Punica granatum*). *Molecules* 22(10): 1606.  
<https://doi.org/10.3390/molecules22101606>  
PMid:28946708 PMCid:PMC6151597
127. Manthou, E., Georgakouli, K., Deli, C.K., Sotiropoulos, A., Fatouros, I.G., Kouretas, D., Haroutounian, S., et al. (2017). Effect of pomegranate juice consumption on biochemical parameters and complete blood count. *Exp Ther Med.* 14(2): 1756-1762.  
<https://doi.org/10.3892/etm.2017.4690>  
PMid:28781633 PMCid:PMC5526177

128. Mollazadeh, H., Sadeghnia, H.R., Hoseini, A., Farzadnia, M., Boroushaki, M.T. (2016). Effects of pomegranate seed oil on oxidative stress markers, serum biochemical parameters and pathological findings in kidney and heart of streptozotocin-induced diabetic rats. *Ren Fail.* 38(8): 1256-1266. <https://doi.org/10.1080/0886022X.2016.1207053>  
PMid:27453190
129. Attia, N.Y.A., Abd El-Aleem, I.M., El-Tobgy, K.M.K., Mohamed, H.F. (2014). Biochemical studies on pomegranate. *Ann Agric Sci., Moshtohor* 52(4): 481-494. <https://doi.org/10.21608/assjm.2014.111897>