

# SYNTHESIS OF A HIGH-PERFORMANCE ANTIMICROBIAL O-QUATERNIZED ALGINATE – A PROMISING POTENTIAL ANTIMICROBIAL AGENT

MAHMOUD H. ABU ELELLA,\* MARWA M. ABDEL-AZIZ\*\* and NAHED A. ABD EL-GHANY\*

\*Chemistry Department, Faculty of Science, Cairo University, Giza, 12613, Egypt

\*\*Regional Center for Mycology and Biotechnology, Al-Azhar University, Cairo, 11651, Egypt

✉ Corresponding author: N. A. Abd El-Ghany, dr\_nahed\_055@yahoo.com

Received October 1, 2020

Three novel biologically active quaternized sodium alginates were synthesized *via* the reaction of sodium alginate (SA) with 3-chloro-2-hydroxypropyl trimethylammonium chloride, at room temperature for different time intervals (1, 3 and 6 h), to produce quaternized sodium alginates designated as QSA1, QSA3 and QSA6. The percentage degree of quaternization (DQ%) significantly increased with increasing the reaction time. Images from FTIR, <sup>1</sup>H-NMR, XRD and SEM have confirmed the chemical structures of the QSA samples. Their antimicrobial activity was investigated against bacteria and fungi using XTT assay, and the results showed that all QSA samples displayed high growth inhibition capacity of the tested microorganisms, compared to zero inhibition for SA, as shown by their lower minimum inhibitory concentration (MIC). The QSA6 was the best antimicrobial composite, displaying the same MIC value as that of the used reference drugs. The developed composites were found to be safe on normal human fibroblast cells (WI-38 cell line), by evaluating them using cytotoxic activity measurement, which makes QSA a promising material in biomedical and food applications.

**Keywords:** quaternized sodium alginate, Quat188, antimicrobial activity, XTT assay, cytotoxicity

## INTRODUCTION

Microbial infection is considered as one of the most serious problems that threaten millions of people worldwide. It is caused by pathogenic microorganisms as *Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Salmonella enteritidis* as bacteria and *Aspergillus niger* as a fungus, leading to different illnesses, such as nausea, pneumonia, abdominal pain, enterocolitis, pyogenic liver abscess, hemorrhagic colitis, diarrhea, hemolytic uremic syndrome and vomiting. Consequently, on a global level, researchers have been doing their best to develop new effective antimicrobial systems.<sup>1-5</sup> In view of their important, effective and broad-spectrum activity against resistant microorganisms, polymers have recently attracted great interest.<sup>6-9</sup> In addition, antimicrobial polymers may be used in comprehensive preservation and packaging of food, and in the textile industry.<sup>10,11</sup> Consequently, the development of antimicrobial polymers is one of the main areas of antimicrobial drug delivery research with the objective to control microbial infection in biomedical devices,<sup>12</sup> wound healing,<sup>13</sup> food packaging,<sup>14</sup> textiles, cosmetics products<sup>15</sup> and water treatment systems.<sup>16</sup>

Due to their excellent properties, polysaccharides have been commonly used in a wide range of applications in the biomedical and food industries, as they have biocompatibility, biodegradability, low cost, and non-toxicity to human cells.<sup>17</sup> Among polysaccharides, alginates have attracted considerable interest in the development of biopolymers, because they represent sustainable polymers present in the brown algae (Phaeophyceae) cell walls.<sup>18</sup> Alginate is a linear anionic polysaccharide composed of  $\beta$ -D-mannuronate and  $\alpha$ -L-guluronate connected by glycosidic bonds (1, 4). It is not intrinsically antimicrobial in nature and thus most work related to the use of alginate-based materials with antibacterial properties depends on the incorporation of antibiotic segments into the alginate matrix, either chemically by the grafting technique, or physically by ionic interaction.<sup>19</sup>

It is well known that quaternary ammonium salts are cationic compounds that contain alkyl groups and have a strong antimicrobial activity,<sup>20-23</sup> primarily due to their ability to promote ionic exchange between the membrane cell and the positively charged species, resulting in microbial cell membrane damage, resulting in intracellular components leakage and cell death. Polysaccharides as chitosan,<sup>24</sup> cellulose<sup>25</sup> and starch,<sup>26</sup> have been quaternized with N-(3-chloro-2-hydroxypropyl) trimethylammonium chloride and other quaternized agents to produce quaternized materials with

strong antimicrobial activity, which have been used in numerous biomedical and food applications.<sup>27-29</sup> Furthermore, sodium alginate was blended with quaternized poly(4-vinyl-N-carboxymethylpyridine),<sup>30</sup> quaternized carboxymethyl chitosan/clay nanocomposite<sup>31</sup> and quaternized chitosan; the blending process greatly improved the physical, chemical and antimicrobial properties of alginate. No studies have been made on direct quaternization of alginate *via* introducing the quaternary ammonium group on the alginate backbone so far.

Therefore, in the present study, our aim was to prepare quaternized sodium alginates (QSAs) *via* the reaction of sodium alginate with N-(3-chloro-2-hydroxypropyl) trimethylammonium chloride at room temperature to be used as active antimicrobial agents. The chemical structures of the prepared QSAs were verified by FTIR, <sup>1</sup>H-NMR, SEM, and XRD analysis. The effect of changing the reaction time on the degree of quaternization (DQ%) was studied. In addition, the antimicrobial activities of the prepared QSAs were examined against various pathogenic microorganisms: *Staphylococcus aureus* (*S. aureus*) as a Gram-positive bacterium, *Pseudomonas aeruginosa* (*P. aeruginosa*) and *Salmonella enteritidis* (*S. enteritidis*) as Gram-negative bacteria, and *Aspergillus niger* (*A. niger*) as a fungus. The cytotoxicity of the modified alginate on the human cell line was also evaluated.

## EXPERIMENTAL

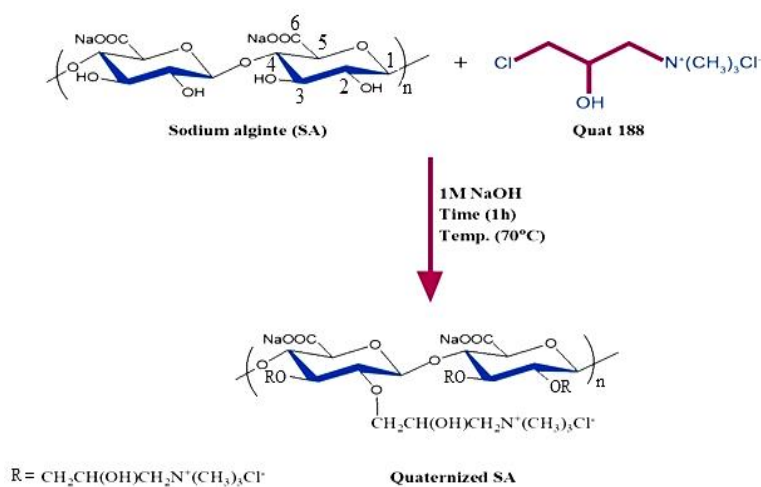
### Materials

Sodium alginate (SA) was purchased from Alpha-Chemika (India). Quaternary ammonium salt, Quat 188, (N-(3-chloro-2-hydroxypropyl)-trimethyl ammonium chloride solution, 60 wt% in H<sub>2</sub>O) was supplied from Sigma-Aldrich, Germany. We purchased regenerated cellophane dialysis tubes, with molecular weight cut off 12-14 x 10<sup>3</sup> g mol<sup>-1</sup> from Serva Electrophoresis, Heidelberg, Germany. The tested pathogens: *Staphylococcus aureus* (ATCC 25923) as a Gram-positive bacterium, *Pseudomonas aeruginosa* (ATCC 15692) and *Salmonella enteritidis* (ATCC 13076) as Gram-negative bacteria, and *Aspergillus niger* (ATCC 9142) as a fungus, were supplied by Regional Center for Mycology and Biotechnology of Azhar University, Egypt.

### Methods

#### Synthesis of O-quaternized sodium alginate (QSA)

A predetermined amount of 1 g of SA (0.005 mol) was dissolved in 10 mL of distilled water and the pH of the solution was adjusted to 11 with 1M NaOH. An equal weight ratio (0.005 mol) of N-(3-chloro-2-hydroxypropyl) trimethylammonium chloride (Quat188) was gradually added to the above solution. The pH of the solution changed to 8 after the addition of Quat188. The mixture was well stirred under nitrogen atmosphere at 70 °C for different periods of time: 1, 3 and 6 h (Scheme 1). After the mentioned time, the solution was neutralized by 1M HCl. The product was purified using a dialysis membrane against distilled water for three days, in which, water was changed once daily. After three days, the resulted product was precipitated with cold ethanol and collected by centrifugation (3,000 rpm for 10 min) and then drying at 60 °C to constant weight. Finally, the degree of quaternization on SA chains was calculated from <sup>1</sup>H-NMR spectroscopy.<sup>32</sup>



Scheme 1: Synthesis of quaternized sodium alginate composite (QSA)

### **XTT assay for determination of minimum inhibitory concentration (MIC)**

Minimum inhibitory concentration is known as the lowest antimicrobial agent concentration that inhibits a microorganism's growth during overnight incubation. The MIC of all grafted samples was determined by XTT assay (Cell Proliferation Kit II), using a microdilution method previously mentioned by Tunney *et al.*,<sup>33</sup> against different pathogenic microorganisms, namely *Staphylococcus aureus* (*S. aureus*) as Gram-positive bacteria, *Pseudomonas aeruginosa* (*P. aeruginosa*) and *Salmonella enteritidis* (*S. enteritidis*) as Gram-negative bacteria and *Aspergillus niger* (*A. niger*) as a fungus. The microbial inoculates were prepared and the suspensions were adjusted to 10<sup>6</sup> CFU/mL. After that, the tested samples (SA and QSA1, QSA3 and QSA6) and the reference drugs – Ciprofloxacin, as a broad spectrum antibacterial drug, and Amphotericin B, as an antifungal one – were prepared. Then, two-fold serial dilution of the stock samples (125, 62.5, 31.25, 15.63, 7.81, 3.90, 1.95, 0.98, 0.49 and 0.24 µg/mL) was performed in 96 well micro-plates, in which, 40 µL of the growth medium brain heart infusion, 10 µL of inoculums and 50 µL of both tested sample solution and standard drugs were added. DMSO was used as a negative control. For antibacterial activity and antifungal activity assessment, the plates were incubated at 37 °C for 24 h, and at 25 °C for 48 h, respectively. After the time of incubation, tetrazolium salt (2,3-bis[2-methoxy-4-nitro-5-sulfophenyl]-2H-tetrazolium-5-carboxanilide) (XTT), 40 µL, was added to the wells, then incubated in the dark at 37 °C for 1 h. After incubation, the colorimetric change in the XTT reduction assay was measured at 492 nm, using a micro-titer plate reader (Tecan Sunrise absorbance reader; Tecan UK, Reading, UK). Finally, the percentage of growth inhibition was determined according to Equation (1) below:

$$\text{Inhibition percentage} = (1 - \text{Measured sample absorption} / \text{Control absorbance}) \times 100 \quad (1)$$

The results are expressed as the mean value of three reproducible experiments.

### **Cytotoxicity evaluation using viability assay**

For the cytotoxicity assay, in 100 µL of growth medium, normal human lung fibroblast cells (WI-38 cell line) were seeded at a cell concentration of 1x10<sup>4</sup> cells per well in a 96-well plate. Serial doubling dilutions of the chemical compound tested were added to the convergent cell monolayers dispensed in the 96-well, flat-bottomed microliter plates (Falcon, NJ, USA) with a multichannel pipette. For each test sample concentration, three wells were used, the control cells were incubated without a test sample, with or without DMSO. The little percentage of DMSO present in the wells (maximum 0.1%) was found not to affect the experiment. The yield of viable cells was determined by the colorimetric method after incubation of the cells. All experiments were performed in triplicates and a calculation was made of the cell cytotoxic effect of each tested compound. The optical density was determined using the micro-plate reader (SunRise, TECAN, Inc, USA) to assess the number of viable cells and the viability percentage was calculated as [(ODt/ODc)] x100 per cent, where ODt is the mean optical density of the wells treated with the sample, and ODc is the mean optical density of untreated cells. The cytotoxic concentration (CC50), the concentration needed to cause toxic effects in 50% of intact cells, was determined for each concentration from graphic plots of the dose response curve, using of Graphpad Prism (San Diego, CA, USA) software.<sup>34</sup>

### **Measurements**

FTIR spectra of Quat, SA and QSA composites were acquired on a JR-Affinity<sup>-1</sup> Shimadzu Fourier Transform Spectrophotometer, using KBr pellets within the wavenumber range of 4000-400 cm<sup>-1</sup> at 25 °C.

<sup>1</sup>H-NMR spectra of Quat, SA and QSA composites were analyzed by a Varian Mercury VX-300 NMR Spectrometer and the spectra were operated at 300 MHz in D<sub>2</sub>O as solvent.

X-ray diffractograms of SA and QSA samples were recorded on an X-ray powder diffractometer (Philips Xpert MPD Pro), using Ni-filter and CuK $\alpha$  radiation source (50 kV/40 mA). The relative intensity was recorded at a scan speed of 1 step s<sup>-1</sup> in the scattering range 2 $\theta$  from 5° to 60°.

Surface morphology images of the tested samples were examined with SEM (Quanta 250 FEG). The freeze-dried samples were loaded on the surface of an aluminum SEM specimen holder and sputter coated with gold, before photographed.

## **RESULTS AND DISCUSSION**

### **Characterization and structure confirmation of QSA composites**

QSA composites were prepared by the reaction of sodium alginate with *N*-(3-chloro-2-hydroxypropyl) trimethylammonium chloride at room temperature. It seemed that the quaternary alkyl moiety attacks the secondary OH groups, forming etherifying quaternized composite. The chemical structure of QSA composites was detected using FTIR and <sup>1</sup>H-NMR tools, which provided information about the newly added functional groups, while the morphology and surface topology were clarified through SEM images. The inner structure and the changes in the alginate crystallinity upon quaternization were studied by XRD analysis. In addition, the reaction time was varied to

investigate its effect on the percentage degree of quaternization, which increased significantly with increasing the reaction time up to 6 h, as calculated from  $^1\text{H-NMR}$  analysis.

### Fourier transform infrared (FTIR) spectroscopy

Figure 1 shows the FTIR spectra of SA, Quat188 and QSA6 (as representative example). The FTIR spectrum of SA presented the following common absorption bands: a broad band around  $3435\text{ cm}^{-1}$  related to the hydrogen bonded OH groups, the absorption band at  $2928\text{ cm}^{-1}$  referred to the stretching vibration of  $-\text{C-H}$  bonds. Strong absorption bands appeared at  $1616$  and  $1420\text{ cm}^{-1}$  attributed to asymmetrical and symmetrical vibrations of the carboxylate  $-\text{COO}^-$  groups, respectively. The peaks at  $1091$ ,  $1031\text{ cm}^{-1}$  are assigned to the stretching vibration of  $-\text{C-O-C-}$  and  $\text{C-OH}$  of the pyranose ring, respectively, the peak at  $944\text{ cm}^{-1}$  corresponds to the overlapped  $-\text{C-C-H}$ ,  $\text{C-OH}$  (bending) and  $\text{C-O}$  groups, the peak at  $880\text{ cm}^{-1}$  is related to anomeric  $\beta\text{-C1-H}$  deformation.<sup>35,36</sup>

On the other hand, the spectrum of Quat displayed vibration bands at the following wavenumbers:  $3406\text{ cm}^{-1}$  assigned to hydrogen bonded OH groups,  $2958\text{ cm}^{-1}$  corresponding to  $\text{C-H}$  stretching vibration,  $1482$  and  $1426\text{ cm}^{-1}$  related to bending vibration of the methyl ( $-\text{CH}_3$ ) and methylene ( $-\text{CH}_2-$ ) groups, respectively. The observed peak at  $1090\text{ cm}^{-1}$  is signed to  $\text{C-O}$  bonds, the weak absorption band at  $1302\text{ cm}^{-1}$  refers to OH bending vibration. The vibrations at  $781$ ,  $747$  and  $717\text{ cm}^{-1}$  are related to  $\text{N-Cl}$  bonds, that at  $687\text{ cm}^{-1}$  is attributed to out-of-plane  $\text{C-H}$  bending.<sup>37</sup>

The structure of quaternized sodium alginate QSA6 (as representative example) was significantly confirmed by its FTIR spectrum, which showed the following characteristic peaks: (1) the broad peak at  $1090\text{ cm}^{-1}$  resulting from the additional ether linkage  $-\text{C-O-C}$  between alginate and the quaternary alkyl moiety; (2) the absence of peaks at  $781$ ,  $747$  and  $717\text{ cm}^{-1}$  corresponding to  $\text{N-Cl}$  bonds of Quat; (3) there was no change in the position of the peak at  $1616\text{ cm}^{-1}$ , corresponding to the carboxylate group of the  $-\text{COO}^-$ , which confirmed the attack of the quaternized group on the secondary OH groups of alginate. Thus, the aforementioned data support the quaternization process and the formation of the QSA composite.

It is worth mentioning that the degree of quaternization of sodium alginate increased with the increase in the reaction time, as evidenced by the FTIR spectra provided in Figure 1 (b). Accordingly, the FTIR spectra of QSA1, QSA3 and QSA6 showed a significant increase in the intensity of the peaks relating to the quaternized moiety around  $1482$ ,  $1426$  and  $1090\text{ cm}^{-1}$ , corresponding to the  $-\text{CH}_3$  group,  $-\text{CH}_2$  group and the etheric  $-\text{C-O-C}$ -bond, respectively, with the increase in the reaction time from QSA1 to QSA6.

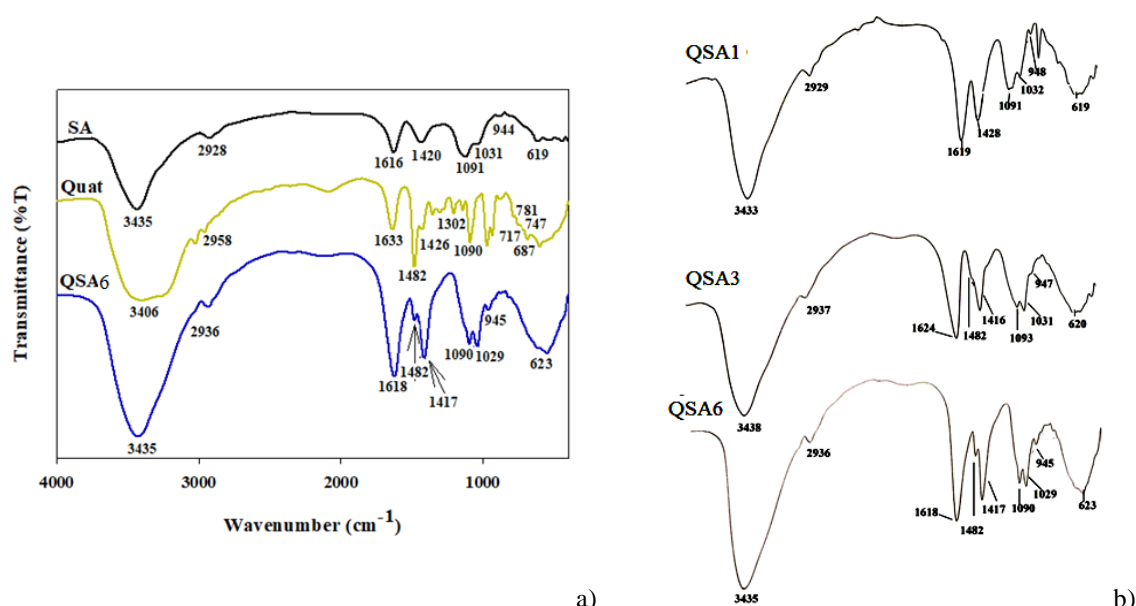


Figure 1: FTIR spectra of (a) Quat, SA and QSA6, (b) QSA1, QSA3 and QSA6

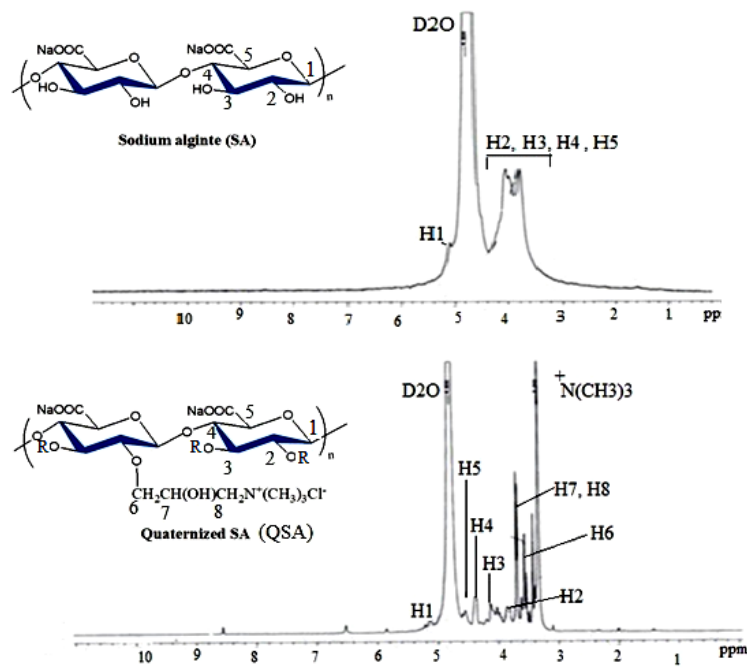


Figure 2:  $^1\text{H-NMR}$  spectra of SA and QSA6

### Proton nuclear magnetic resonance ( $^1\text{H-NMR}$ )

Other evidence of the formation of the new QSA composite was elucidated from the  $^1\text{H-NMR}$  analysis. Figure 2 illustrates the  $^1\text{H-NMR}$  spectra of both SA and QSA6 (as representative example). Thus, the SA spectrum shows the following common signals: a multiplet at  $\delta = 3.8\text{--}4.1$  ppm, corresponding to the protons of C2, C3, C4 and C5 (H2, H3, H4 and H5) of the pyranose ring; a singlet at  $\delta = 5.1$  ppm, related to the anomeric proton (H1) of the glucopyranose unit.<sup>38</sup> Meanwhile, the spectrum of QSA6 displays the same signals of SA and, in addition, a sharp singlet at  $\delta = 3.4$  ppm, assigned to the quaternary amino methyl groups ( $\text{N}^+(\text{CH}_3)_3$ ) and other signals at  $\delta = 3.5$  and 3.7 ppm, attributed to  $-\text{CH}$  and  $\text{CH}_2$  groups (H6, H7 and H8) of the quaternary ammonium salt moiety, which confirm the achievement of the quaternization process. Furthermore, a sharp signal observed at  $\delta = 4.8$  ppm is related to the solvent  $\text{D}_2\text{O}$ .

### Determination of the percentage degree of quaternization from $^1\text{H-NMR}$ measurements

The quaternization of SA proceeded by binding of the quaternized alkyl moiety with the secondary OH of SA to produce the QSA composite. The percentage degree of SA quaternization (DQ%) was measured kinetically at various time intervals (1, 3 and 6 h) in order to determine the optimal time that yielded a greater degree of quaternization. Based on the reaction time, three quaternized samples QSA1, QSA3 and QSA6 were prepared, and the DQ% was determined after each reaction time from the integration under the proper signals in  $^1\text{H-NMR}$  spectra, according to the following equation:

$$\text{DQ}\% = \left( \frac{[\text{N}^+(\text{CH}_3)_3]}{[\text{H}]} \right) \times 1/9 \times 100 \quad (2)$$

where  $[\text{N}^+(\text{CH}_3)_3]$  is an integral part of the trimethylamine group and  $[\text{H}]$  is an integral part of C1 protons of the pyranose ring (Fig. 3a).<sup>38</sup> The results confirmed that the DQ% increased as the reaction time increased. It was observed that the percentage of quaternization reached 63.8% after 1 h of the reaction, rose to 70.6% after 3 h, and further increased to 85.0% after 6 h of reaction time. The correlation between DQ% and the reaction time was depicted in (Fig. 3b). The data indicate that the reaction time is an important factor influencing the phase of quaternization.

### Powder X-ray diffraction (XRD)

In order to show the changes in the inner structure of sodium alginate after incorporation of the quaternary ammonium moiety, the XRD diffraction patterns of SA and QSA composites were analyzed. Figure 4 illustrates the XRD pattern of alginate, displaying a crystalline structure shown by the two crystalline peaks at  $2\theta = 13.7^\circ$  and  $23.0^\circ$ ,<sup>39,40</sup> as a result of the large number of internal hydrogen bonding and the ordering of the chains. These crystalline peaks entirely disappeared after the

quaternization process and were replaced by an amorphous structure clearly observed in the XRD patterns of all the QSA composites. It seems that the incorporated quaternary ammonium moiety substantially breaks the intramolecular and intermolecular hydrogen bonds between the alginate chains, thus increasing the intermolecular distance between the neighboring chains, leading to a more open and amorphous structure. It became clear that, as the degree of quaternization rose, the amorphous fraction increased, depending on the time of the quaternization process.

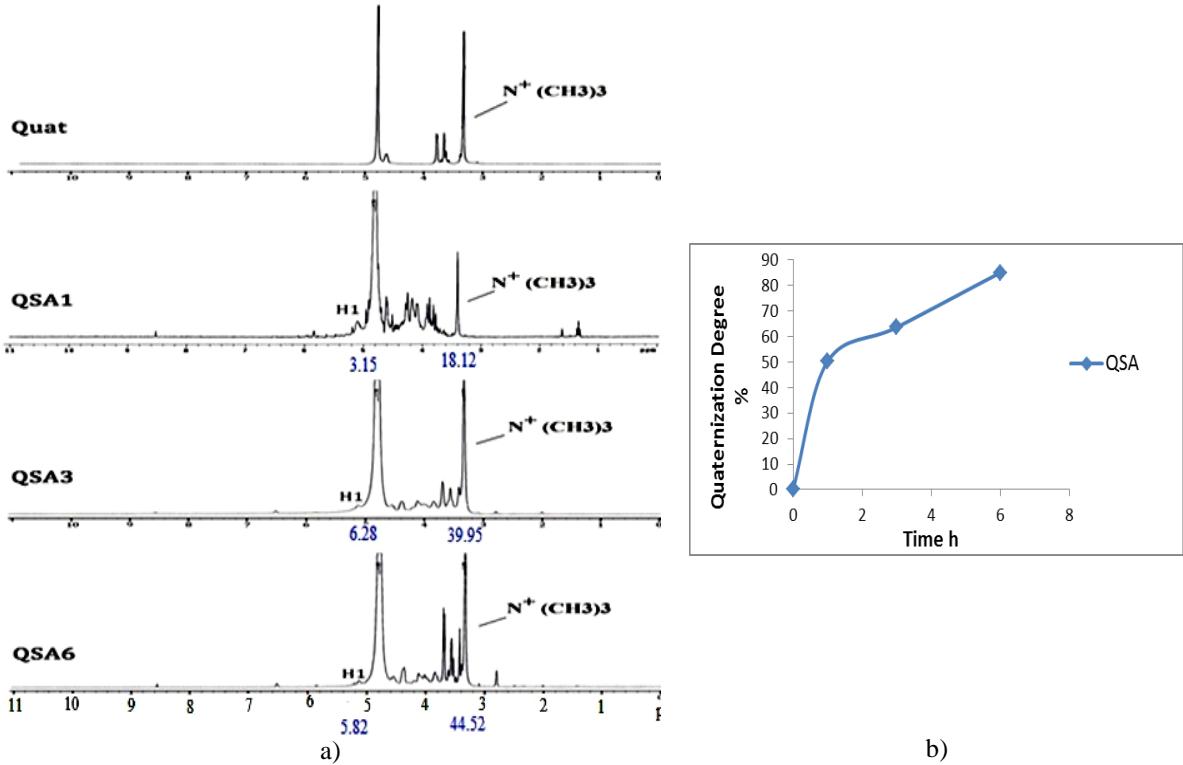


Figure 3: (a) Determination of quaternization degree at different time intervals; (b) Evolution of quaternization degree over time

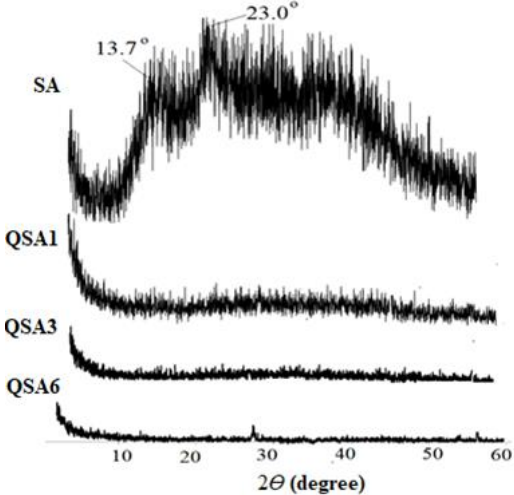


Figure 4: XRD patterns of SA and QSA1, QSA3 and QSA6 composites

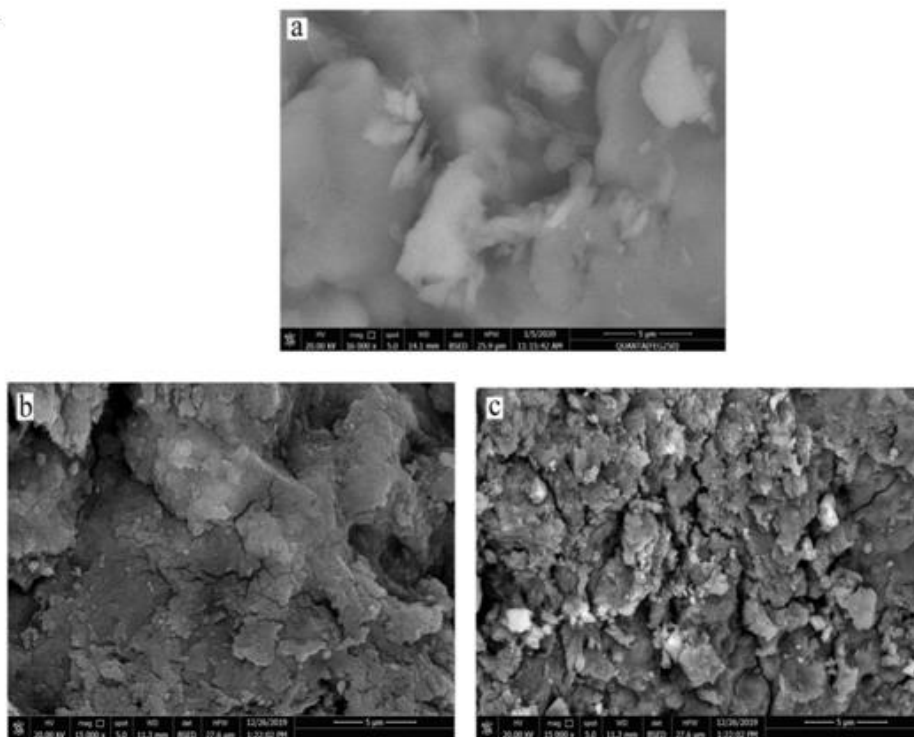


Figure 5: SEM images of (a) SA, (b) QSA1 and (c) QSA6, at 15000x magnification

### Scanning electron microscopy (SEM)

Scanning electron microscopy was used to study the changes that occurred in SA surface morphology after the quaternization process. From the SEM images, the SA surface appears smooth, with a small amount of lumps due to the presence of  $-COO-$  groups, while QSA images show a rough surface, with a lot of conglomerates due to the quaternary ammonium moiety incorporated onto the SA backbone. The SEM images illustrate that the degree of agglomeration increased with increasing the degree of quaternization from QSA1 to QSA6, as shown in Figure 5.

### Antimicrobial activity of QSA composites

XTT is a negatively charged, tetrazolium salt that turns orange when it is reduced to a soluble formazan dye. This extracellular reduction is carried out by electron transport across the plasma membrane of a living cell. The amount of XTT reduced is reflective of the cellular metabolic activity. The absorbance can be measured and compared to the absorbance of a control solution of untreated cells to determine if cellular metabolic activity has increased or decreased.

For determining the antimicrobial activity of sodium alginate (SA) and QSA composites against four pathogenic microorganisms, including *Staphylococcus aureus* (*S. aureus*) as Gram-positive bacteria, *Pseudomonas aeruginosa* (*P. aeruginosa*) and *Salmonella enteritidis* (*S. enteritidis*) as Gram-negative bacteria and *Aspergillus niger* (*A. niger*) as a fungus, the XTT assay proceeded by measuring the minimum inhibitory concentration values (MICs) of the tested samples that inhibited the growth of the microorganisms. Ciprofloxacin and Amphotericin B were used as conventional drugs for bacteria and fungi, respectively.

### Antibacterial activity

Figure 6 shows the antimicrobial activity of SA and QSA composites against both Gram-positive and Gram-negative bacteria. The experimental data showed that SA did not have antimicrobial activity against the studied bacteria, there was no colorimetric shift of XTT, which is in agreement with the results of previously mentioned work,<sup>41</sup> while QSA composites exhibited higher antibacterial activity towards the tested bacteria. It seems that the quaternized moiety (3-chloro-2-hydroxy propyl trimethylammonium chloride), significantly strengthened the antibacterial potency of sodium alginates. The higher antibacterial activity of QSA resulted from its easy penetration into the bacterial cytoplasm and the interaction with the negatively charged cell membrane, leading to disruption of the

cell function and metabolic activity, as indicated by the XTT colorimetric shift. The three QSA composites exhibited inhibition potency for bacterial growth, as seen from their lower MIC values in Table 1. The data illustrate that the MIC values significantly decreased as the percentage of quaternization (DQ%) increased from QSA1 (DQ%, 63.8%) to QSA6 (DQ%, 85.0%). This may be associated with the increasing density of the positively charged species ( $N^+(CH_3)_3$ ) in the polymeric matrix. This resulted in an increase in the electrostatic interaction with the negatively charged bacterial cell membrane, thereby retarding the growth of bacteria. From Table 1, the MIC values measured for QSA1, QSA3 and QSA6 ranged from 1.95 to 31.25  $\mu\text{g/mL}$  against *S. aureus*, 0.98 to 7.8  $\mu\text{g/mL}$  against *S. enteritidis*, and 7.81 to 62.5  $\mu\text{g/mL}$  against *P. aeruginosa*, compared to the MIC values of 0.98 and 3.9  $\mu\text{g/mL}$  of the reference drug Ciprofloxacin against the same strains, respectively. Interestingly, the QSA6 sample (relative to QSA1, QSA3) exhibited the strongest antibacterial activity, very close and in many cases equal to that presented by the reference drug Ciprofloxacin, as inferred from their MIC values. They displayed MIC values equal to 1.95, 0.98 and 7.81  $\mu\text{g/mL}$  against the tested bacteria *S. aureus*, *S. enteritidis* and *P. aeruginosa*, compared to 0.98, 0.98 and 3.9  $\mu\text{g/mL}$  against the same bacterial strains, respectively. Although Gram-positive and Gram-negative bacteria have different cell wall structure,<sup>19</sup> the QSA composites showed stronger inhibition activity on both of them. This might be due to the presence of both hydrophilic and hydrophobic species in the QSA chains, which promote the penetration of the samples into the bacterial cells, leading to cell permeability disruption, in addition to binding with the cytoplasm DNA, preventing the formation of RNA, thus causing death in bacteria.<sup>42,43</sup>

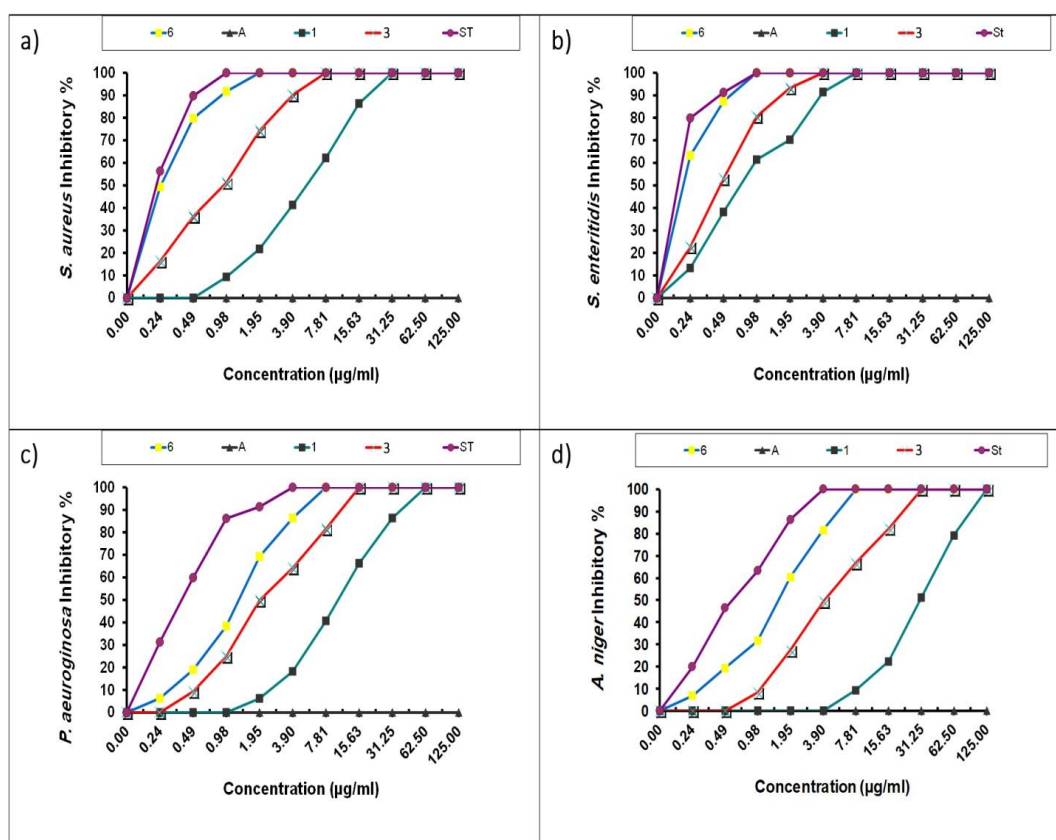


Figure 6: Microbial growth inhibition, %, at various concentrations of alginate (A) and its three quaternized compounds at three different times: 1 (QSA1), 3 (QSA3) and 6 h (QSA6) against a) *Staphylococcus aureus*, b) *Pseudomonas aeruginosa*, c) *Salmonella enteritidis* and d) *Aspergillus niger*, compared with standards including Ciprofloxacin as a broad spectrum antibacterial and Amphotericin B as an antifungal. The results are expressed as the mean value of three reproducible experiments



Table 1  
 Minimum inhibitory concentrations required to kill 50 (MIC 50) and 100% (MIC) of the tested pathogens

Pathogens	SA		QSA1		QSA3		QSA6		ST	
	MIC	MIC 50	MIC	MIC 50	MIC	MIC 50	MIC	MIC 50	MIC	MIC 50
Gram-positive bacterium	Ciprofloxacin									
<i>Staphylococcus aureus</i> ATCC 25923	NA	NA	31.25	5.5	7.81	0.94	1.95	0.25	0.98	0.24
Gram-negative bacteria	Ciprofloxacin									
<i>Salmonella enteritidis</i> ATCC 13076	NA	NA	7.81	0.74	3.9	0.46	0.98	0.24	0.98	0.24
<i>Pseudomonas aeruginosa</i> ATCC 15692	NA	NA	62.5	10.67	15.63	1.97	7.81	1.35	3.9	0.31
Fungus	Amphotericin B									
<i>Aspergillus niger</i> ATCC 9142	NA	NA	125	30.6	31.25	4.0	7.81	1.6	3.9	0.52

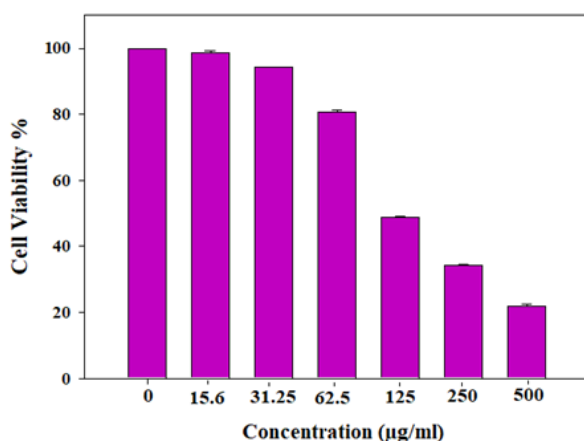


Figure 7: Cytotoxic effect of different concentrations of QSA6 composite on the viability of human lung fibroblast cells

### **Antifungal activity**

The antifungal experimental data clarified that SA did not have any inhibition activity on the growth of the tested fungus *A. niger*, while all the QSA composites exhibited higher potency in inhibition of the fungus growth, as shown in Figure 6. The measured MIC values of the QSA composites resulting in 100% inhibition ranged from 7.81 to 125 µg/mL, as the inhibition efficiency increased by increasing the DQ% from QSA1 to QSA6. The hypothesized mechanism for the antifungal activity of QSA is based on: (1) the electrostatic force interactions between the cationic groups on QSA chains and the electronegative charges on the fungus surfaces, leading to leakage of intracellular electrolytes and proteinaceous constituents; (2) QSA can penetrate inside the hyphae of *A. niger* and intervene with the activities of the enzyme that are responsible for fungus growth, similarly to the findings of previous work reported on a chitosan derivative.<sup>44</sup>

### **Estimation of cytotoxicity of QSA composite**

The cytotoxicity exhibited by QSA6 (as representative example) on normal human lung fibroblast cells (WI-38 cell line) was investigated utilizing different concentrations of the sample ranging between 0 and 500 µg/mL, as shown in Figure 7. WI-38 cell viability was not affected by the QSA6 sample at concentrations below 15.6 µg/mL – no inhibitory action was observed in this sample below this concentration. The cytotoxicity gradually increased by increasing sample concentration up to 500 µg/mL. Moreover, the concentration of QSA6 that caused a toxic impact in half of the intact cells (IC<sub>50</sub>) was 122 µg/mL. From Figure 6, the QSA6 sample showed strong activity against the tested bacteria and fungi, with minimum inhibitory concentration (MIC) values (ranging from 0.98 to 7.81 µg/mL), *i.e.* smaller than the cytotoxic concentration (15.6 µg/mL). This showed that the use of such materials on the human cells is safe and they could be recommended as antimicrobial agents in biomedical and food applications.

### **CONCLUSION**

Quaternization of sodium alginate (SA) proceeded in alkaline medium using 3-chloro-2-hydroxypropyltrimethylammonium chloride (Quat 188) in different time intervals: 1, 3, and 6 h, to prepare quaternized sodium alginate (QSA) composites with high degree of quaternization. The grafting of the quaternized moiety onto SA through chemical covalent bond was elucidated *via* FTIR, H-NMR, XRD, SEM, and TGA techniques. Three different QSAs were successfully fabricated with various degrees of quaternization (DQ)%, calculated from H-NMR images, as 63.8% (QSA1), 70.6% (QSA3), and 85.0% (QSA6). Moreover, the data of antimicrobial activity for the three different QSA composites and unmodified SA against four pathogenic foodborne pathogens: *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Salmonella enteritidis* and *Aspergillus niger*, exhibited microbial attack against SA from all the microorganisms, while the three prepared QSAs showed high broad spectrum antimicrobial activity against the four examined microorganisms. Also, the results illustrated the sustained increased antimicrobial activity with a boost in DQ%. Thus, the QSA6 sample demonstrated the highest inhibition of all the microorganisms' growth, and its MIC (100% inhibition) values were

listed as 0.98, 1.95 and 7.81 µg/mL against bacteria: *S. enteritidis*, *S. aureus* and *P. aeruginosa*, respectively, as well as highly effective activity against *A. niger* fungus as 7.81 µg/mL. Additionally, the cytotoxic activity of QSA6 was investigated against normal human cells and the data confirmed that the prepared grafted composites can be used as safe and promising antimicrobial agents in medicine and food industries.

## REFERENCES

- <sup>1</sup> A Abbasi, Z. Emam-Djomeh, M. A. E. Mousavi and D. Davoodi, *Food Chem.*, **143**, 379 (2014), <https://doi.org/10.1016/j.foodchem.2013.08.018>
- <sup>2</sup> H. Jolink, R. de Boer, P. Hombrink, R. E. Jonkers, J. T. van Dissel *et al.*, *J. Infect.*, **74**, 81 (2017), <https://doi.org/10.1016/j.jinf.2016.10.010>
- <sup>3</sup> N. A. Mohamed, N. A. Abd El-Ghany, Mona M. Fahmy and M. M. Abdel-Aziz, *Polym. Adv. Technol.*, **1** (2020), <https://doi.org/10.1002/pat.5179>
- <sup>4</sup> Y. Chang, H. Yoon, D. H. Kang, P. S. Chang and S. Ryu, *Int. J. Food Microbiol.*, **244**, 19 (2017), <https://doi.org/10.1016/j.ijfoodmicro.2016.12.007>
- <sup>5</sup> M. H. Abu Elella, M. Sabaa, D. H. Hanna, M. M. Abdel-Aziz and R. R. Mohamed, *J. Drug. Deliv. Sci. Technol.*, **57**, 101673 (2020), <https://doi.org/10.1016/j.jddst.2020.101673>
- <sup>6</sup> H. Xing, M. Lu, T. Yang, H. Liu, Y. Sun *et al.*, *Acta Biomater.*, **86**, 15 (2019), <https://doi.org/10.1016/j.actbio.2018.12.041>
- <sup>7</sup> A. Muñoz-Bonilla, C. Echeverria, A. Sonseca, M. P. Arrieta and M. Fernández-García, *Materials*, **12**, 641 (2019), <https://doi.org/10.3390/ma12040641>
- <sup>8</sup> M. M. Konai, B. Bhattacharjee, S. Ghosh and J. Haldar, *Biomacromolecules*, **19**, 1888 (2018), <https://doi.org/10.1021/acs.biomac.8b00458>
- <sup>9</sup> C. Ergene, K. Yasuhara and E. F. Palermo, *Polym. Chem.*, **9**, 2407 (2018), <https://doi.org/10.1039/C8PY00012C>
- <sup>10</sup> M. Mujtaba, R. E. Morsi, G. Kerch, M. Z. Elsabee, M. Kaya *et al.*, *Int. J. Biol. Macromol.*, **121**, 889 (2019), <https://doi.org/10.1016/j.ijbiomac.2018.10.109>
- <sup>11</sup> C. Vasile, *Materials*, **11**, 1834 (2018), <https://doi.org/10.3390/ma11101834>
- <sup>12</sup> P. Yadav, H. Yadav, V. G. Shah, G. Shah and G. Dhaka, *J. Clin. Diagn.*, **9**, ZE21 (2015), <https://doi.org/10.7860/JCDR/2015/13907.6565>
- <sup>13</sup> A. S. Veiga and J. P. Schneider, *J. Pept. Sci.*, **100**, 637 (2013), <https://doi.org/10.1002/bip.22412>
- <sup>14</sup> D. S. Cha and M. S. Chinnan, *Crit. Rev. Food Sci. Nutr.*, **44**, 223 (2004), <https://doi.org/10.1080/10408690490464276>
- <sup>15</sup> M. Shahid and F. Mohammad, *Ind. Eng. Chem.*, **52**, 5245 (2013), <https://doi.org/10.1021/ie303627x>
- <sup>16</sup> M. U. Sankar, S. Aigal, S. M. Maliyekkal, A. Chaudhary, A. A. Kumar *et al.*, *PNAS*, **110**, 8459 (2013), <https://doi.org/10.1073/pnas.1220222110>
- <sup>17</sup> S. Kondaveeti, P. V. de Assis Bueno, A. M. Carmona-Ribeiro, F. Esposito, N. Lincopan *et al.*, *Carbohydr. Polym.*, **186**, 159 (2018), <https://doi.org/10.1016/j.carbpol.2018.01.044>
- <sup>18</sup> O. Jeon, D. S. Alt, S. M. Ahmed and E. Alsberg, *Biomaterials*, **33**, 3503 (2012), <https://doi.org/10.1016/j.biomaterials.2012.01.041>
- <sup>19</sup> N. A. Abd El-Ghany, M. S. A. Aziz, M. M. Abdel-Aziz and Z. Mahmoud, *Int. J. Biol. Macromol.*, **134**, 912 (2019), <https://doi.org/10.1016/j.ijbiomac.2019.05.078>
- <sup>20</sup> E. Obłąk, A. Piecuch, J. Rewak-Soroczyńska and E. Paluch, *Appl. Microbiol. Biotechnol.*, **103**, 625 (2019), <https://doi.org/10.1007/s00253-018-9523-2>
- <sup>21</sup> M. E. Fait, L. Bakas, G. L. Garrote, S. R. Morcelle and M. C. Saparrat, *Appl. Microbiol. Biotechnol.*, **103**, 97 (2019), <https://doi.org/10.1007/s00253-018-9467-6>
- <sup>22</sup> P. Makvandi, R. Jamaledin, M. Jabbari, N. Nikfarjam and A. Borzacchiello, *Dent. Mater.*, **34**, 851 (2018), <https://doi.org/10.1016/j.dental.2018.03.014>
- <sup>23</sup> I. Mulder, J. Siemens, V. Sentek, W. Amelung, K. Smalla *et al.*, *Rev. Environ. Sci. Biotechnol.*, **17**, 159 (2018), <https://doi.org/10.1007/s11157-017-9457-7>
- <sup>24</sup> H. Tan, R. Ma, C. Lin, Z. Liu and T. Tang, *Int. J. Mol. Sci.*, **14**, 1854 (2013), <https://doi.org/10.3390/ijms14011854>
- <sup>25</sup> Y. Song, Y. Sun, X. Zhang, J. Zhou and L. Zhang, *Biomacromolecules*, **9**, 2259 (2008), <https://doi.org/10.1021/bm800429a>
- <sup>26</sup> N. A. Abd El-Ghany and Z. M. Mahmoud, *Polym. Bull.*, **1** (2020), <https://doi.org/10.1007/s00289-020-03417-8>
- <sup>27</sup> R. C. Goy, S. T. Morais and O. B. Assis, *Rev. Bras. Farmacogn.*, **26**, 122 (2016), <https://doi.org/10.1016/j.bjpp.2015.09.010>
- <sup>28</sup> D. Hu, T. Qiang and L. Wang, *Wound Med.*, **16**, 15 (2017), <https://doi.org/10.1016/j.wndm.2016.12.003>
- <sup>29</sup> S. Geresh and R. Dawadi, *Carbohydr. Polym.*, **43**, 75 (2000), [https://doi.org/10.1016/S0144-8617\(99\)00194-0](https://doi.org/10.1016/S0144-8617(99)00194-0)

- <sup>30</sup> Y. Zhang, Y. Zhou and C. Liu, *Polym. Bull.*, **71**, 1909 (2014), <https://doi.org/10.1007/s00289-014-1163-9>
- <sup>31</sup> B. Liu, J. Luo, X. Wang, J. Lu, H. Deng *et al.*, *J. Biomater. Sci. Polym.*, **24**, 589 (2013), <https://doi.org/10.1080/09205063.2012.701160>
- <sup>32</sup> Y. Wang and W. Xie, *Carbohydr. Polym.*, **80**, 1172 (2010), <https://doi.org/10.1016/j.carbpol.2010.01.042>
- <sup>33</sup> M. M. Tunney, G. Ramage, T. R. Field, T. F. Moriarty and D. G. Storey, *AAC*, **48**, 1879 (2004), <https://doi.org/10.1128/AAC.48.5.1879-1881.2004>
- <sup>34</sup> H. Itagaki, S. Hagino, S. Kato, T. Kobayashi and M. Umeda, *Toxicol. In Vitro*, **5**, 139 (1991), [https://doi.org/10.1016/0887-2333\(91\)90034-B](https://doi.org/10.1016/0887-2333(91)90034-B)
- <sup>35</sup> H. Daemi and M. Barikani, *Sci. Iran.*, **19**, 2023 (2012), <https://doi.org/10.1016/j.scient.2012.10.005>
- <sup>36</sup> M. Taha, K. Aiedeh, Y. Al-Hiari and H. Al-Khatib, *Pharmazie*, **60**, 736 (2005), <https://www.ingentaconnect.com/content/govi/pharmaz/2005/00000060/00000010/art00003#expand/collapse>
- <sup>37</sup> O. Kostyk, O. Budishevskaya, V. Vostres, Z. Nadashkevych and S. Voronov, *Catalysis*, **150**, 1 (2019), <https://doi.org/10.32434/0321-4095-2019-127-6-113-120>
- <sup>38</sup> L. Marci, M. C. Meloni, A. M. Maccioni, C. Sinico, F. Lai *et al.*, *Chem. Select.*, **1**, 669 (2016), <https://doi.org/10.1002/slct.201600145>
- <sup>39</sup> Q. Wang, X. Hu, Y. Du and J. F. Kennedy, *Carbohydr. Polym.*, **82**, 842 (2010), <https://doi.org/10.1016/j.carbpol.2010.06.004>
- <sup>40</sup> G. Yang, L. Zhang, T. Peng and W. Zhong, *J. Membrane Sci.*, **175**, 53 (2000), [https://doi.org/10.1016/S0376-7388\(00\)00407-5](https://doi.org/10.1016/S0376-7388(00)00407-5)
- <sup>41</sup> H. E. Salama, M. S. A. Aziz and M. W. Sabaa, *Int. J. Biol. Macromol.*, **116**, 443 (2018), <https://doi.org/10.1016/j.ijbiomac.2018.04.183>
- <sup>42</sup> T. Xu, M. Xin, M. Li, H. Huang, S. Zhou *et al.*, *Carbohydr. Res.*, **346**, 2445 (2011), <https://doi.org/10.1016/j.carres.2011.08.002>
- <sup>43</sup> M. H. Abu Elella, E. S. Goda, H. M. Abdallah, A. E. Shalan, H. Gamal *et al.*, *Int. J. Biol. Macromol.*, **167**, 1113 (2021), <https://doi.org/10.1016/j.ijbiomac.2020.11.065>
- <sup>44</sup> Z. Guo, R. Xing, S. Liu, Z. Zhong, X. Ji *et al.*, *Carbohydr. Polym.*, **71**, 694 (2008), <https://doi.org/10.1016/j.carbpol.2007.10.021>