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# Chapter

# 4-Hydroxynonenal Is Linked to Sleep and Cognitive Disturbances in Children: Once upon the Time of COVID-19

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#### **Abstract**

The better prognosis of COVID-19 in children conferred a higher survival rate, but a higher prevalence of post-COVID sequalae, including insomnia and defective cognition. COVID-19 triggered oxidative stress, with hyperlipidemia correlated with susceptibility to severe COVID-19. Consequently, lipids peroxidation could be a likely candidate for disease progression and sequalae. Hence, this overview explored one of the commonly studied lipid peroxides, 4-hydroxynonenal (4-HNE), in terms of gamma-amino butyric acid (GABA) and glutamate. Higher glutamate and lower glutamine, a GABA substrate, triggered severe COVID-19. Increased glutamate and inflammatory cytokines induced GABA endocytosis, reducing the anti-inflammatory and antioxidant effects of GABA. Defective glutathione antioxidant was detected in Down syndrome, the latter was associated with severe COVID-19. Increased 4-HNE, due to consumption of electronic devices and flavors containing 1-bromopropane, was increased in inflammatory neurologic disorders. A higher hippocampal 4-HNE triggered excitotoxicity and cognitive deficits. Hippocampal inflammation and loss were also evident in COVID-19. 4-HNE might play role in disturbing sleep and cognition in children during COVID-19, a hypothesis that could be verified in future research by redeeming 4-HNE in the sputum and urine of children. Currently, supplying children with optimum dietary antioxidants, while rationalizing the use of flavors is to be encouraged.

**Keywords:** COVID-19, insomnia, 4-Hydroxynonenal, cognition, GABA, lipid peroxidation

#### 1. Introduction

Until early January 2023, over 660 million cases were diagnosed with coronavirus disease (COVID-19), most cases residing in Europe with much less cases in Africa, and the United States being the most affected country [1]. When first recognized in late 2019 and early 2020, COVID-19 was thought of as an 'adult and older' disease, exempting the younger population. Later, this revelation was falsified by positive infected cases found among neonates and children. Despite lower death rates at

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younger ages, COVID-19 survivors are mainly those in the pediatric age group. The milder disease was related to lower immune responses in children [2].

Knowing that COVID-19 has affected 6.3% of children aged 5-14 years old from December 30, 2019 till September 13, 2022 [3] would give us clues about the magnitude of post-COVID in children, even if definite evidence is still missing [4]. Reports outlining the possibility of asymptomatic disease occurring in children [5] suggest that the global prevalence of COVID in children may be much higher than the registered cases. In contrast, other studies highlighted that children were especially afflicted by hyperinflammatory multisystem syndrome [6–8]. The issue arose when some studies detected that recurrent infection is likely to occur in school children, compared to pre-school age, and that some of the affected children tested negative for viral antigen and antibodies and they did not shed the virus [9-11], which can lead us to infinite unexplored areas of research targeting whether the virus remains dormant or not, for how long and where, in the neurological system and/or elsewhere. Are there late-onset sequalae that could affect the future quality of life of young generations or even be transmitted to their offspring after decades? The anonymous fate of viral infection in children who survived but did not shed the virus should draw the attention of investigators toward the outcomes of COVID-19 on various aspects, including cognition as one crucial vector in childhood determining the ability to learn, develop new skills, and have a future fruitful life.

During the post-COVID period, survivors suffered neuropsychiatric symptoms, including anxiety and mood disturbances [12]. Such neuropsychiatric sequalae were attributed to the viral invasion of the brain [13] and neural control over the immune system [14]. In their retrospective cohort study, Taquet, Geddes, et al. [15] noticed a higher liability for insomnia during 6 months post-COVID, one plausible explanation was the impact of inflammatory cytokines over neuroendocrine sleep mediators [16].

Although a direct link between insomnia and liability for a more severe COVID-19 was not conclusive, yet inferences could be made based on previous studies showing that persons who had less than 7 hours of daily sleeping were three times more vulnerable to getting a flu attack [17]. An association between disturbed sleep or even prolonged sleep and a state of low-grade systemic inflammation was suggested [18, 19], the latter impaired the immune defenses against the respiratory pathogens [16, 20, 21], added to a higher risk of developing pneumonia [22]. The deleterious effects of disturbed sleep over immunity were also emphasized in Module 2 of The National Institute for Occupational Safety and Health (NIOSH) [23] declaring more than 50% decline in the production of antibodies following influenza vaccination in presence of sleep shifting, compared to regular sleep.

As the susceptibility to COVID-19 is higher with cardiovascular diseases, diabetes mellitus (DM), and obesity, and as dyslipidemia is common among these vulnerable groups [24–26], a causal relationship might exist between lipid metabolism and COVID-19 morbidity. Apart from the structural, non-structural, and accessory proteins identified for severe acute respiratory syndrome virus (SARS-CoV-2), lipid-based structures remain to be identified, especially when knowing their pivotal role in viral fusion, entry, and replication and that the host lipid profile is altered following COVID-19 [27, 28]. The involvement of lipids in promoting the creation of severe acute respiratory syndrome (SARS-CoV-2) progeny is becoming increasingly an interesting entity that awaits further exploration. Interestingly, insomnia has been reported to alter lipid metabolism and trigger lipid peroxidation [29]. Both insomnia and lipid peroxidation were associated with cognitive decline [30, 31]. In this context, this overview will focus on the relationship between COVID-19, insomnia, and

4-hydroxynonenal (4-HNE), as the most studied among lipid peroxides, on one side, and cognitive defects, on the other side.

#### 2. COVID-19 in children: insomnia and cognitive defects

The American Academy of Sleep Medicine [32] has quoted from the Centers of Disease Control (CDC) that sleep disturbances among middle- and high-school students were highly prevalent. This prevalence was also noticed in survivors of COVID-19 who experienced long-term insomnia [33] with younger age being more vulnerable [34]. During COVID-19, higher liability to insomnia was also reported in students, compared to workers, and in undergraduates, compared to postgraduates [35, 36].

Novel stressors were superimposed with the emergence of COVID-19, including locking down at home, studying in an isolated environment with no social interactions, lacking friends, missing both physical activities and teamwork-based learning, having one or more of beloved family members affected, added to dealing with stressed parents [37]. Learning at home has posed a greater stressful challenge to parents whose anxiety was transferred to their children [38]. All these stressors contributed to higher anxiety in children, and subsequent sleep issues [39], including, inability to fall asleep, insufficient duration of sleep, excessive sleep duration, nightmares, and unstable sleep timings. In turn, disturbed sleep, by triggering mood swings, caused a further reduction in social interactions [40], and impaired psychological and physical well-being [41]. Interestingly, being home alone, using electronic devices during studying, playing, or chatting, were associated with poor sleep quality in children with autism spectrum disorder (ASD) [42].

Attention, as one of the cognitive domains, tended to decline with insomnia. Focused attention, detected by responding to a specific stimulus while overlooking other stimuli, was reduced with insomnia [43]. Vigilance (sustained attention) or the ability to keep alertness over time [44] was negatively affected by insomnia with reduced accuracy and prolonged time needed to perform vigilance-related tasks [45]. Similarly, shifting attention or the ability to adapt and modify the focus of attention, requiring a higher level of cognition [46], was defective in cases with insomnia [47]. However, some other studies did not prove these correlations, especially for the simplest form of attention, focused attention [48, 49].

Another cognitive domain, memory, was negatively impacted by insomnia [50, 51], whether working memory or that of the implicit (procedural) or explicit (declarative) categories. These three memory categories correspond to the inability of keeping information for a short period [46], learning new skills, and recalling a new learned material after a delay, respectively [52]. In a meta-analysis, there was a mild correlation between insomnia and working memory, yet the authors declared that results could be biased by the heterogenicity between studied groups in different studies. Other studies denied such an insomnia-memory relationship [48, 53].

Whatever is the magnitude of controversy regarding the correlation between insomnia and cognitive defects, most studies agreed about the correlation between stress and both cognition [54, 55] and sleep, especially, at a young age [56].

What links cognition to sleep at the neuronal level? In terms of memory, the role of glutamate, the main excitatory neurotransmitter in the brain, in the encoding and consolidation of memory through binding to its ionotropic receptors, N-methyl-D-aspartate (NMDA), and its metabotropic receptors (mGLuRs), respectively, has been established [57, 58]. Li et al. [59] in a meta-analysis of the African population

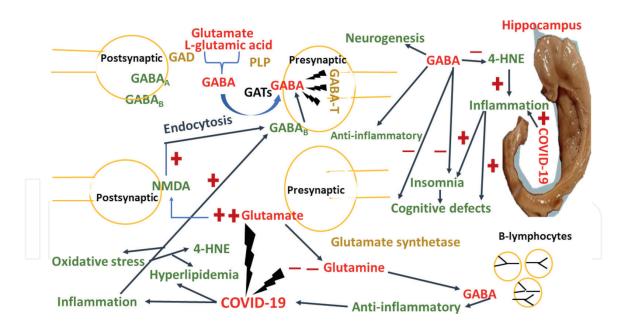
concluded that the higher glutamate, and the lower its byproduct, glutamine, the more severe would be COVID-19 and related cognitive defects.

In astrocytes, glutamate is converted by glutamine synthetase to glutamine, the substrate of both gamma-aminobutyric acid (GABA) and glutamate [60]. GABA, the main inhibitory neurotransmitter in the mammalian brain, is also modulated, through NMDA activation when glutamate is released, then presynaptic auto-receptors GABA<sub>B</sub> stimulation, mediating GABA endocytosis [61]. In the brain, more than 50% of synapses are GABAergic [62], signifying the pleiotropic effects of GABA.

### 3. Gamma-aminobutyric acid linking sleep and cognition

GABA is an amino acid present in plants, bacteria, fungi, animals, and humans [63–65]. GABA in vertebrates is synthesized and metabolized (as shown in **Figure 1**) [66, 67].

GABA acts on two types of receptors, the fast ionotropic or ligand-gated ion channel,  $GABA_A$ , and the slow metabotropic or G protein-coupled receptor,  $GABA_B$ . The binding of GABA to  $GABA_A$  results in chloride influx and a fast hyperpolarization of postsynaptic neurons. While  $GABA_B$  receptors are present in presynaptic and postsynaptic [68]. Postsynaptic  $GABA_B$  stimulation produces a slow, but long-term hyperpolarization. Presynaptic  $GABA_B$  activation reduces the release of many neurotransmitters, including GABA itself, yielding either an excitatory or inhibitory brain signaling, depending on whether the suppressed neurotransmitter was



GABA, glutamate, 4-HNE in COVID-19-related insomnia and cognitive defects. GABA in vertebrates is derived from L-glutamic acid or its salts, glutamate, by a decarboxylation reaction, catalyzed by GAD, and using PLP as a cofactor. After its release, GABA is uptaken by GATs 1, 2, and 3 as well as BGT-1 and metabolized by GABA-transaminases (GABA-T). Upon glutamate release, it simulates NMDA, with subsequent presynaptic GABAB activation, mediating GABA endocytosis. Increased glutamate, along with reduced glutamine, aggravates COVID severity. GABA is secreted from B-lymphocytes to exert anti-inflammatory effects. GABA promotes antioxidants, reducing lipid peroxides, including 4-HNE. COVID-19 triggers inflammation and oxidative stress. Both COVID-19 and increased hippocampal 4-HNE cause inflammation and neurodegeneration, precipitating insomnia and cognitive decline, which could be antagonized by GABA anti-inflammatory and neurogenesis effects. GABA: Gamma-aminobutyric acid; GAD: Glutamic acid decarboxylase; PLP: Pyridoxal 5'-phosphate; GATs: GABA transporters; BGT Betaine GABA transporter; GABA-T: GABA-transaminases; Fe<sup>3+:</sup> Ferric; GSH: Glutathione;

NMDA: N-methyl-D-aspartate; 4-HNE: 4-Hydroxynonenal; COVID-19: Coronavirus disease.

inhibitory or excitatory. This means that if auto-receptors' presynaptic  $GABA_B$  is stimulated, GABA release is dampened leading to a depolarizing postsynaptic current, or disinhibition. If a heteroreceptor  $GABA_B$  was activated, glutamate release could be suppressed, which would favor an inhibitory status [69].

From functional perspective, GABA is implicated in sleep regulation and memory enhancement [70]. GABA deficiency can lead to insomnia, anxiety, and impaired stress responses [71–73]. The established role of GABA in sleep and sedation led to the wide use of benzodiazepines (BZs) as hypnotics and anxiolytics, acting by enhancing the binding of GABA to its receptors, GABA<sub>A</sub> [74, 75]. Unfortunately, BZs are associated with a high risk of tolerance and dependence [76] which mitigated their long-term use.

Despite the crucial role of GABA in the processes of sleep and cognition, it is not the only one, as inflammatory factors seem to contribute as well. In COVID-19, during the cytokine storm, excessive amounts of pro-inflammatory cytokines are produced, of which the tumor necrosis factor-alpha (TNF- $\alpha$ ) induced the endocytosis of GABA<sub>A</sub>, possibly rationalizing the associated sleep disturbances.

In the cognition domain, fast-spike GABAergic interneurons play a crucial role in the generation of electroencephalographic gamma rhythms [77], as well as hippocampal theta rhythm, corresponding to exploratory behavior [78]. Inhibitory postsynaptic potentials (IPSPs), generated by GABA, assist memory acquisition in rodents and humans [79, 80], and the progression to memory consolidation requires GABA<sub>B</sub> activation [81]. This GABA-cognitive function was experimentally verified when the passive avoidance learning of mice and rats was inhibited after the blockade of GABA<sub>B</sub> using baclofen [82, 83].

Furthermore, by promoting neurogenesis, GABA enriches long-term memory and learning processing [84]. This was emphasized in stressful conditions when mice with depressive-like symptoms exhibited defective neurogenesis and reduced microglia [85], along with reduced survival in neural stem progenitor cell culture [86]. In ASD, decreased glutamic acid decarboxylase (GAD), GABA<sub>A</sub>, and GABA<sub>B</sub> were observed in postmortem specimens [87], with GABA<sub>A</sub> reduction possibly underlying the co-existing delayed linguistic abilities [88], along with behavioral deficits; the latter being also demonstrated in transgenic animal models [89].

Although multiple sclerosis (MS) is mainly a disease of young adults, it is the most common neurologic disorder due to immunologic dysfunction in children and adolescents [90]. In MS, where 65% of patients have disturbed memory and attention, low plasma GABA was detected [91]. Recent reports revealed aggravated or de novo symptoms of MS associated with COVID-19 [92]. Hence, GABA might be a likely candidate for COVID-related cognitive derangement.

## 4. Inflammation, oxidative stress, and GABA: key targets in COVID

A growing body of evidence supports the secretion of GABA and its precursors, glutamine, and glutamate, from murine and human B-lymphocytes, [93]. While GABA<sub>A</sub> reduces T-cell response to antigens [94] and dampens inflammation, it endorses regulatory T-cells [95]. In turn, T-cells enhance the expression of GABA receptor subunits [96]. Additionally, GABA transporter-1 (GAT-1), found only on antigen-primed T-cells, arrested the proliferation of CD4+ and CD8+ T-cells [97]. Such GABA immunomodulatory effect could prevent the tissue damage elicited by inflammatory responses in cases of autoimmune diseases, as inferred from rodent models of

DM and MS [98–100]. In patients with DM, the secretion of TNF- $\alpha$  and interleukin (IL)-6 (IL-6) from T-cells was successfully inhibited using GABA [101, 102].

Such systemic anti-inflammatory potentiality of GABA was detected also in macrophages and dendritic cells of rodents and humans, expressing the respective fast GABA<sub>A</sub> and slow GABA<sub>B</sub> receptors [103]. GATs dampen the functions and release of proinflammatory cytokines as demonstrated in a mouse model of autoimmune encephalomyelitis (EAE) [97]. Thus, it was not surprising to find that the most common subtype of GABA<sub>A</sub> in the brain, ( $\alpha$ 1 $\beta$ 2 $\gamma$ 2), was also expressed in immune cells [104]. Conversely, immuno-stimulation and cytokines release promoted the neuronal sequestration of extracellular GABA [67]. In the brain, neuroinflammation, vascular insufficiency, and the pro-inflammatory cytokines, such as TNF- $\alpha$ , interferon-gamma (IFN- $\gamma$ ), IL-6 and IL-1 $\beta$  enhanced GAT expression, favoring GABA degradation [105–108].

In the context of lung diseases, GABA, along with enhanced GABA<sub>A</sub> and GABA<sub>B</sub> activities could limit acute lung injury in rodent models and ameliorate clinical outcomes in humans on ventilation [109, 110]. As an inhibitor of platelet aggregation, GABA, by inhibiting the formation of the thromboxane A2 [111], might have an additional clinical privilege in patients whose pulmonary thrombosis is attributed to the severe COVID-19 [112, 113]. These assumed benefits of early treatment with GABA in COVID-19 were verified in mice infected with mouse hepatitis virus (MHV-1) [114], another coronavirus whose symptoms mimic those of COVID-19 [115].

The COVID-associated anxiety and stress could have resulted in lowered immunity [116], which could be reversed using GABA as was emphasized in human volunteers when oral GABA administration resulted in electroencephalographic evidence of relaxed alertness and anti-stress effects (higher alpha-to-lower beta) [117], while increasing salivary IgA [118], as a non-invasive index of enhanced upper respiratory immunity against bacteria and viruses [119].

Interestingly, extracellular glutamine, the GABA precursor, was implicated in viral replication of both DNA and RNA viruses to which SARS-CoV-2 belongs, by incorporation into the Kreb's cycle after conversion by glutaminase (GLS) to alphaketoglutarate ( $\alpha$ -KG), so that the lack of glutamine hampered rhinoviruses replication [120]. Presumably, if GABA synthesis is inhibited, glutamine would be redirected to promote viral replication and, in case of viral infection, defective GABA synthesis would be anticipated secondary to the incorporation of glutamine in the viral replication cycle.

Knowing that COVID-19 can precipitate oxidative stress [121, 122] while GABAergic neurons are especially susceptible to the neuro-damaging effects of the reactive oxygen species (ROS), generated during oxidative stress [123], makes both GABA and oxidative stress likely candidates for aggravating the sequalae of COVID-19.

Hypercholesterolemia might perpetuate viral infections as was the case in mice infected with lymphocyte choriomeningitis virus (LCMV) [124]. Some viral infections and related treatments can induce long-term changes in lipid metabolism as well. After 12 years of SARS-CoV, survivors had higher cell membrane phospholipids, namely, phosphatidylinositol and lysophosphatidylinositol, attributed to corticosteroid administration during the infection [125]. In the post-infection period of SARS-CoV, lysocardiolipin acetyltransferase (LCLAT), phosphoinositide phosphatase (PIP), and diacylglycerol (DG) kinase, enzymes involved in lipid metabolism, were upregulated [126].

Coronaviruses consume the intracellular membranes of host cells to build their own replication nests called "double-membrane vesicles (DMVs)," where it preserves

its own viral proteins and robbed host factors, to ensure a suitable lipid bedding for a successful viral replication [127].

Of interest to our discussion is the increased total cholesterol (TC) in patients with COVID-19, favoring viral invasion, with a positive correlation to the severity of symptoms [93]. The lipid changes might be attributed to hypoxia and were also shared with patients having a chronic obstructive pulmonary disease (COPD) [128]. On the other hand, normal lipid metabolism seems preemptive in the context of pulmonary and neuronal disorders as sphingolipids were implicated in protection from a lung injury, added to their anti-inflammatory, anti-coagulant, along with their neuroprotective effects [129, 130]. Hyperlipidemia and oxidative stress during COVID make lipids peroxidation likely candidates for post-COVID syndrome.

# 5. Lipid peroxidation

Oxidative stress is conceived as an imbalance between oxidants and antioxidants, in favor of oxidation. In physiology, such oxidative stress is minimal and well-equilibrated in a process known as "the redox potential." It is noteworthy to mention that an imbalance in the antioxidant direction is deleterious and causes "reductive stress" [131].

Conversely, when the antioxidant mechanisms are overwhelmed, oxidative stress occurs. The consecutive reversible oxidative stress and irreversible oxidative damage are to be blamed for many pathologic conditions [132, 133]. With defective antioxidant mechanisms such as in the case of vitamin E (alpha-tocopherol) or vitamin C deficiency, excess reactive oxygen and nitrogen species are produced. The issue is that a propagation chain reaction perpetuates lipid peroxidation [131] as shown in (**Figure 2**). The interruption of chain reaction occurs when two free radicals are conjugated or when antioxidants break the chain.

Lipid peroxidation was formerly known for oils and fats in our diet. It involves oxidative damage to cellular structures, including cell membranes in plants and animals, causing cellular death. This destructive process includes the generation of lipid radicals, the uptake of oxygen, the re-organization of double bonds in unsaturated lipids, and the production of breakdown products such as alcohols, ketones, alkanes, aldehydes, and ethers. Lipid peroxidation results in an easily breakable cell membrane with plenty of polyunsaturated fatty acids (PUFAs) and transition metals. Lipid peroxidation reduces membrane fluidity and makes it more permissible and easily invaded. Apart from the loss of cell membrane integrity, protein synthesis is disrupted, as well as macrophage function, along with derangement of chemotactic signals and altered enzyme activity [134]. All membranes of cellular structures are damaged, including those of mitochondria, microsomes, peroxisomes, and cell membranes [135]. Lipid peroxidation toxicity affects the liver, kidneys, and to our interest, neurological structures, where it takes part in neurodegenerative, inflammatory, and infectious diseases [136].

Considering the brain as a susceptible organ to oxidative stress, the intracellular antioxidant, free glutathione (GSH) plays a crucial role by eliminating peroxides [137] in a reaction catalyzed by glutathione peroxidase (GSH-Px), oxidizing GSH to GSH disulfide (GSSG) [138]. Thus, the GSH/GSSG can be used as a determinant of the redox status of cells [139]. Defective GSH was previously correlated to Down syndrome in children [140]. Recently, Down syndrome was correlated to severe COVID-19 [141].

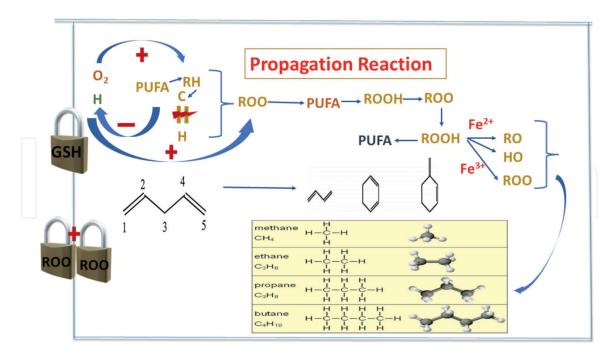


Figure 2.

Propagation reaction of lipid peroxidation. Lipid peroxidation is initiated with hydrogen subtraction and oxygen addition. Hydrogen subtraction is promoted in PUFAs by the presence of a double bond of the RH group, leaving the carbon with an unpaired electron. When combined with oxygen, ROO is produced, generating ROOH, capable of repeating the hydrogen subtraction from another PUFA, perpetuating the chain reaction. When lipid peroxides interact with Fe<sup>+2</sup>, RO radicals are produced, when the interaction involves Fe<sup>+3</sup>, ROO radicals are generated. These reactions will end up with cytotoxic aldehydes and hydrocarbon gases as ethane. The interruption of chain reaction occurs when two free radicals are conjugated or when antioxidants, such as GSH, break the chain. PUFA: Polyunsaturated fatty acids; RH: methylene; ROO: Peroxyl; ROOH: Hydroperoxide; Fe<sup>2+:</sup> Ferrous; RO: Alcoxyl; Fe<sup>3+:</sup> Ferric; GSH: Glutathione.

Most of brain GSH is derived from the reducing action of GSH reductase (GR) over GSSG to get back GSH. Another less amount of GSH can be synthesized *de novo* from glutamate, cysteine, and glycine [142].

In contrast to the antioxidant GSH, one of the lipid peroxides, 4-hydroxynonenal (4-HNE), an  $\alpha$ ,  $\beta$ -unsaturated aldehyde, is a potent neurotoxin, derived from the oxidation of  $\omega$ -6 PUFA of cell membranes [143], such as arachidonic acid, linoleic and linolenic acid.

# 6. 4-Hydroxynonenal

4-Hydroxynonenal (4-HNE) is described as a short-chain reactive carbonyl compound [144], having amphiphilic properties yet with lipophilic tendency [143]. Its high electrophilicity makes it reactive to the amino acid residues, namely, cysteine (Cys), histidine (His), and lysine (Lys), in a decrescendo order. 4-HNE can adduct to the cysteine residue of the "flippase" enzyme (amino phospholipid-translocase), an enzyme that maintains lipid bilayer asymmetry by an ATP-dependent process [145]. Forming Michael adducts with nucleophilic sites, 4-HNE can interact with cellular DNA, lipids, and proteins [146]. The destiny of 4-HNE protein adducts is either proteolysis or covalent cross-linking. Additionally, 4-HNE can inactivate GR, reducing the antioxidant ability of GSH [147]. In turn, physiological concentrations of GSH can revert 4-HNE protein adducts to their unadducted condition [148].

The metabolism of 4-HNE occurs by oxidative and reductive processes, employing enzymatic and non-enzymatic pathways [144], in addition to conjugation to GSH catalyzed by the glutathione-S transferases (GST), which contributes to a major part in the detoxification process [149]. Although all these detoxifying processes are present in the mitochondria [150], yet it seems that the mitochondria play little role in 4-HNE degradation in intact tissue.

In the lungs, GST is more active than the liver, then comes the brain in the third place, however, the respiratory capacity to metabolize 4-HNE is limited by slow oxidative-reductive pathways, unlike the liver [151, 152]. 4-HNE can be detected in human breath and sputum [153] and its metabolites can be recovered in urine [154]. A slow metabolism of 4-HNE was previously reported when dealing with rat hearts and kidneys, along with other tissues [155].

To our knowledge, HNE concentration at or below 1  $\mu$ M might be physiological, with in vitro toxicity at 10  $\mu$ M–1 mM [156]. The physiological roles of HNE include, but are not limited to [see **Table 1**] [157–164]. Dianzani [156] mentioned that, in pathologic conditions, the high concentrations of 4-HNE suppress mitochondrial oxidation, lysosomal enzyme activity, adenyl cyclase, sodium pump, protein synthesis, and cell proliferation. Also, while physiological 4-HNE concentrations can affect proteins, favoring proteolysis of the deformed proteins [165], only extra-physiologic concentrations of 4-HNE can increase membrane fluidity [166].

In inflammatory disorders such as osteoarthritis, 20  $\mu$ M HNE suppressed the high nuclear factor-kappa beta (NF- $\kappa$ B) induced by TNF- $\alpha$  overexpression in human

Targets of HNE	Role
Neutrophils chemotactic factor [157]	Increased inflammatory response to invading pathogens
AC [158]	Catalyze the breakdown of ATP to yield cAMP
PLC [159]	Hydrolysis of inositol phospholipids in cell membranes, yielding the intracellular second messengers: ${\rm IP}_3$ and DAG
Caspases [160]	Protease enzymes that mediate programmed cell death, leading, for example, to tumor suppression and axonal degeneration
Hsp 70 [161]	Increased antigens delivery to APCs Suppression of inflammation
Aldose reductase [162]	Cytosolic NADPH-dependent oxidoreductase that catalyzes the reduction of monosaccharides, for example, the reduction of glucose to sorbitol, the first step in glucose metabolism
Hem oxygenases [163]	The degradation of heme to CO, biliverdin and heme iron, mediating anti-inflammatory, anti-apoptotic, and potential anti-viral functions
γ-GCS [164]	Catalyzes the production of γ-glutamylcysteine from both glutamate and cysteine, and other glutamylpeptides and can be used as predictor of defective GSH redox

AC: Adenyl cyclase; ATP: Adenosine triphosphate; cAMP: Cyclic adenosine monophosphate; PLC: Phospholipase C;  $IP_3$ : Inositol 1,4, 5-triphosphate; DAG: diacyl glycerol; Hsp 70: Heat shock proteins 70; APCs: Antigen-presenting cells; NADPH: Nicotinamide adenine dinucleotide phosphate hydrogen; CO: Carbon monoxide;  $\gamma$ -GCS:  $\gamma$ -glutamyl cys synthetase; GSH: Glutathione.

At physiologic concentration, HNE seems to exert immunostimulatory activity by enhancing neutrophils' chemotactic factor, increasing the production of multiple intracellular second messengers, such as cAMP, IP<sub>3</sub>, DAG, mediate tumor suppression, and might promote axonal degeneration and aging, along with immune-stimulatory, anti-inflammatory, anti-apoptotic, and possibly anti-viral functions. HNE catalyzes glucose metabolism and interestingly, can increase antioxidant activity.

**Table 1.** Physiological targets stimulated by hydroxynonenal (HNE).

osteoblasts [167]. While most studies focused on the link between 4-HNE and hepatic insult, few of them found that 4-HNE was also implicated in multiple respiratory and neurological disorders, such as bronchial asthma, COPD [168], Alzheimer's disease (AD), and Parkinson's disease (PD) [169]. The ability of 4-HNE to diffuse from one organ to another [170] might indicate the accumulation of HNE in the lungs, for instance, can affect the brain, and vice versa. Fortunately, GSH was able to suppress 4-HNE protein adducts in the liver, lungs, and brain [152].

In COPD, HNE adducts were increased in bronchial, bronchiolar, alveolar, and endothelial cells as well as macrophages and neutrophils. In alveolar epithelium, HNE adducts were inversely correlated to forced expiratory volume in 1 sec and positively linked to the pro-fibrotic cytokine, transforming growth factor-beta (TGF- $\beta$ ) [171]. In rat alveolar epithelial cells, HNE induced glutamylcysteinyl glycine (GCS), the rate-limiting enzyme in GSH synthesis [172], and enhanced the expression of antioxidants by recruiting nuclear factor erythroid 2-related factor-2 (Nrf2) [173, 174].

*In vitro* exposure to mild stress assisted the accelerated GSH-mediated removal of HNE and enhanced resistance to oxidative stress [175], which might not apply to chronic stress when antioxidants are consumed.

Measuring HNE in umbilical cord plasma, it was increased in full-term newborns exposed to acidosis and in full—as well as pre-term neonates experiencing asphyxia when compared to healthy controls [176]. A suggested role in autoimmunity was reported in children with systemic lupus erythematosus (SLE) when plasma HNE was increased, especially during the active disease stage [177].

The brain is a vulnerable organ that can be affected by oxidative stress owing to its relatively lower antioxidant capacity against a higher oxygen consumption rate, added to the abundance of PUFAs in neuronal cell membranes [178]. Upon 12-day exposure of rats to oral 1- bromopropane (1-BP), a cleaning agent for electronic and optical instruments and an intermediate in the synthesis of pharmaceuticals and flavors, the animals demonstrated behavioral evidence of impaired cognition with underlying oxidative stress as shown by the reduced level of GSH versus increased GSSG, owing to the inhibitory effect of 1-BP over GR, with subsequently increased 4-hydroxynonenal (4-HNE) and malondialdehyde (MDA) [179]. The increased 4-HNE was also replicated in patients with AD showing mild cognitive dysfunction [180]. It is to be noted that while human exposure to 1-BP is by inhalation, yet, in experimental animals, the inhalation route might not yield similar neurological effects as the oral route.

4-HNE can form adducts with glutamate transporter, excitatory amino acid transporter 2 (EAAT2) [181], dopamine transporter, sodium pump [182], dopamine 1 (D1)-like transporter [183], and immunoglobulins [184]. In cultured rat cerebrocortical neurons, HNE uncoupled cholinergic and glutamatergic receptors from the GTP-binding proteins [185]. In patients with ischemia–reperfusion and stroke, plasma HNE was elevated [186]. Immunohistochemical assay of the brain lesions in patients with the progressive demyelinating disease, multiple sclerosis, and the dominant autosomal disorder, Huntington's disease (HD), detected increased HNE [187, 188], along with elevation of the inflammatory marker C-reactive protein in serum of patients with advanced HD. In rat hippocampal cell culture, 10  $\mu$ M HNE hampered sodium pump activity, resulting in increased intracellular free Ca<sup>2+</sup> and predisposition to excitotoxicity [189]. The hippocampus is well known for its relevance to both cognition [190, 191] and insomnia-related cognitive issues at all ages, including children [192, 193], and inflammation and loss were recently reported in COVID-19 [194].

#### 7. Hydroxynonenal- and GABA-targeted therapies

Based on the presumptive involvement of HNE in COVID-related insomnia and subsequent cognitive dysfunction, HNE-targeted therapy might offer an exit doorway that might rescue the young generation. For instance, carnosine, a dipeptide ( $\beta$ -alanyl-L-histidine) abundant in mammalian skeletal muscle, can inhibit the crosslinking of HNE protein adducts [195], and its analogs showed a similar neuroprotective effect as emphasized in rats [196].

Nutritional support seems crucial to sustaining the growth and development of childhood processes, including those related to their emotional, cognitive, and behavioral aspects. Above all, supplying dietary antioxidants, including vitamin E, vitamin C, and glutathione, could be helpful. The consumption of wheat germ oil, sunflower oil and seeds, hazelnuts, and peanut butter, as sources of vitamin E, and red and green pepper, orange, kiwi, and broccoli, providing vitamin C, with recommended daily dietary allowances at 4–13 years old of 7–11 and 25–45 mg, respectively [197].

Despite the controversies regarding the extent of systemic GABA to cross the BBB [198–200] as quoted by Tian et al. [114], supplying dietary GABA could add some benefit as an adjuvant to COVID treatment, especially since GABA has antioxidant properties [201, 202], GABA can be obtained from tomatoes, rice, soybean, and fermented food [70]. It is worthwhile that this policy can be adopted in the context of cognitive affection in children whose anxiety and insomnia could be the major contributing factors to the post-COVID syndrome.

#### 8. Conclusion

Lipid peroxidation, along with inflammatory crisis, plays a crucial role, not only in the prognosis of COVID-19 but also in neurological sequalae, namely, sleep and cognitive issues, by affecting both GABA and glutamate neurotransmission.

4-HNE might have some role in both COVID-19 and its neurological sequalae, triggering hippocampal inflammation and neurodegeneration, by disturbing glutamate/ GABA neurotransmission.

Perhaps a nutritional supply of antioxidants and abstaining from the consumption of flavors could support our children to maintain optimal sleep and develop cognitive skills. The rationale use of electronic devices is also recommended. A more vigorous investigation is still needed to verify the hypothesis of 4-HNE involvement and to explore the feasibility of GABA—and HNE-targeted therapy in children who survived COVID-19 with residual issues regarding sleep and cognition.

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#### **Conflict of interest**

The author declares no conflict of interest.

#### Notes

- COVID-19 triggers insomnia and cognitive defects.
- Higher glutamate, with subsequent low GABA, was associated with severe COVID-19.
- Electronic Devices and flavors could lead to increased 4-HNE.
- Increased 4-HNE caused hippocampal inflammation, an area implicated in sleep and cognition.
- Supplying pediatric nutrition with antioxidants and abstaining from flavors consumption and overuse of electronic devices might prove preemptive in COVID-19, and related sleep and cognitive issues.



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#### References

- [1] The World Health Organization (WHO). WHO coronavirus (COVID-19) dashboard. WHO Coronavirus (COVID-19) Dashboard | WHO Coronavirus (COVID-19) Dashboard with Vaccination Data. 2023 [Accessed 10 January 2023]
- [2] Dong Y, Mo X, Hu Y, Qi X, Jiang F, Jiang Z, et al. Epidemiology of COVID-19 among children in China. Pediatrics. 2020;**145**(6):e20200702. DOI:10.1542/peds.2020-0702
- [3] Hoang A, Chorath K, Moreira M, Evans M, Burmeister-Morton F, Burmeister F, et al. COVID-19 in 7780 pediatric patients: A systematic review. EClinicalMedicine. 2020;24:100433. DOI:10.1016/j.eclinm.2020.100433
- [4] Buonsenso D, Munblit D, De Rose C, Sinatti D, Ricchiuto A, Carfi A, et al. Preliminary evidence on long COVID in children. Acta Paediatrica. 2021;**110**(7):2208-2211. DOI:10.1111/ apa.15870
- [5] Buitrago-Garcia D, Egli-Gany D, Counotte MJ, Hossmann S, Imeri H, Ipekci A, et al. Occurrence and transmission potential of asymptomatic and presymptomatic SARS-CoV-2 infections: A living systematic review and meta-analysis. PLoS Medicine. 2020;17(9):e1003346. DOI:10.1371/journal.pmed.1003346
- [6] Jiang L, Tang K, Levin M, Irfan O, Morris SK, Wilson K, et al. COVID-19 and multisystem inflammatory syndrome in children and adolescents. The Lancet Infectious Diseases. 2020;**20**(11):e276-e288. DOI:10.1016/S1473-3099(20)30651-4
- [7] Dufort EM, Koumans EH, Chow EJ, Rosenthal EM, Muse A, Rowlands J, et al.

- Multisystem inflammatory syndrome in children in New York State. The New England Journal of Medicine. 2020;383(4):347-358. DOI:10.1056/NEJMoa2021756
- [8] Riphagen S, Gomez X, Gonzalez-Martinez C, Wilkinson N, Theocharis P. Hyperinflammatory shock in children during COVID-19 pandemic. Lancet. 2020;**395**(10237):1607-1608. DOI:10.1016/S0140-6736(20)31094-1
- [9] Somekh E, Gleyzer A, Heller E, Lopian M, Kashani-Ligumski L, Czeiger S, et al. The role of children in the dynamics of intra family coronavirus 2019 spread in densely populated area. The Pediatric Infectious Disease Journal. 2020;39(8):e202-e204. DOI:10.1097/INF.00000000000000002783
- [10] Yung CF, Kam KQ, Chong CY, Nadua KD, Li J, Tan NWH, et al. Household transmission of severe acute respiratory syndrome coronavirus 2 from adults to children. The Journal of Pediatrics. 2020;225:249-251. DOI:10.1016/j.jpeds.2020.07.009
- [11] Tosif S, Neeland MR, Sutton P, Licciardi PV, Sarkar S, Selva KJ, et al. Immune responses to SARS-CoV-2 in three children of parents with symptomatic COVID-19. Nature Communications. 2020;**11**(1):5703. DOI:10.1038/s41467-020-19545-8
- [12] Taquet M, Luciano S, Geddes JR, Harrison PJ. Bidirectional associations between COVID-19 and psychiatric disorder: Retrospective cohort studies of 62 354 COVID-19 cases in the USA. Lancet Psychiatry. 2021;8(2):130-140. DOI:10.1016/S2215-0366(20)30462-4
- [13] Meinhardt J, Radke J, Dittmayer C, Franz J, Thomas C, Mothes R, et al.

- Olfactory transmucosal SARS-CoV-2 invasion as a port of central nervous system entry in individuals with COVID-19. Nature Neuroscience. 2020;24(2):168-175. DOI:10.1038/s41593-020-00758-5
- [14] Kreye J, Reincke SM, Kornau HC, Sanchez-Sendin E, Corman VM, Liu H, et al. A therapeutic non-self-reactive SARS-CoV-2 antibody protects from lung pathology in a COVID-19 Hamster model. Cell. 2020;**183**(4):1058-1069. DOI:10.1016/j.cell.2020.09.049
- [15] Taquet M, Geddes JR, Husain M, Luciano S, Harrison PJ. 6-month neurological and psychiatric outcomes in 236 379 survivors of COVID-19: A retrospective cohort study using electronic health records. Lancet Psychiatry. 2021;8(5):416-427. DOI:10.1016/S2215-0366(21)00084-5
- [16] Ibarra-Coronado EG, Velazquez-Moctezuma J, Diaz D, Becerril-Villanueva LE, Pavon L, et al. Sleep deprivation induces changes in immunity in Trichinella spiralisinfected rats. International Journal of Biological Sciences. 2015;**11**(8):901-912. DOI:10.7150/ijbs.11907
- [17] Cohen S, Doyle WJ, Alper CM, Janicki-Deverts D, Turner RB. Sleep habits and susceptibility to the common cold. Archives of Internal Medicine. 2009;**169**(1):62-67. DOI:10.1001/archinternmed.2008.505
- [18] Lasselin J, Rehman JU, Akerstedt T, Lekander M, Axelsson J. Effect of long-term sleep restriction and subsequent recovery sleep on the diurnal rhythms of white blood cell subpopulations. Brain, Behavior, and Immunity. 2015;47:93-99. DOI:10.1016/j.bbi.2014.10.004
- [19] Irwin MR, Olmstead R, Carroll JE. Sleep disturbance, sleep duration, and inflammation: A systematic review

- and Meta-analysis of cohort studies and experimental sleep deprivation. Biological Psychiatry. 2016;**80**(1):40-52. DOI:10.1016/j.biopsych.2015.05.014
- [20] Spiegel K, Leproult R, Van Cauter E. Impact of sleep debt on physiological rhythms. Revue Neurologique (Paris). 2003;159(11 Suppl):6S11-6S20 ISSN (Linking): 0035-3787
- [21] Ferrie JE, Kivimaki M, Akbaraly TN, Singh-Manoux A, Miller MA, Gimeno D, et al. Associations between change in sleep duration and inflammation: Findings on C-reactive protein and interleukin 6 in the Whitehall II study. American Journal of Epidemiology. 2013;178(6):956-961. DOI:10.1093/aje/kwt072
- [22] Patel SR, Malhotra A, Gao X, Hu FB, Neuman MI, Fawzi WW. A prospective study of sleep duration and pneumonia risk in women. Sleep. 2012;35(1):97-101. DOI:10.5665/sleep.1594
- [23] The National Institute of Occupational Safety and Health (NIOSH). Centers for Disease Control AND Prevention (CDC). NIOSH Training for Nurses on Shift Work and Long Work Hours. Part 1. The Risks and Why These Occur. Module 2. How shift work and long work hours increase health and safety risks. Module 2. Sleep and the Immune System | NIOSH | CDC [Accessed 10 January 2023]
- [24] Klop B, Elte JW, Cabezas MC. Dyslipidemia in obesity: Mechanisms and potential targets. Nutrients. 2013;5:1218-1240. DOI:10.3390/nu5041218
- [25] Krist AH, Davidson KW, Mangione CM, Barry MJ, Cabana M, Caughey AB, et al. Behavioral counseling interventions to promote a healthy diet and physical activity for cardiovascular disease prevention in adults with

cardiovascular risk factors: US preventive services task force recommendation statement. JAMA. 2020;**324**:2069-2075. DOI:10.1001/jama.2020.21749

- [26] Savelieff MG, Callaghan BC, Feldman EL. The emerging role of dyslipidemia in diabetic microvascular complications. Current Opinion in Endocrinology Diabetes Obesity. 2020;27:115-123. DOI:10.1097/MED.00000000000000033
- [27] Hu X, Chen D, Wu L, He G, Ye W. Declined serum high density lipoprotein cholesterol is associated with the severity of COVID-19 infection. Clinical Chimica Acta. 2020;**510**:105-110. DOI:10.1016/j. cca.2020.07.015
- [28] Peng Y, Wan L, Fan C, Zhang P, Wang X, Sun J, et al. Cholesterol metabolism-impacts on SARS-CoV-2 infection prognosis. medRxiv. Preprint posted online August 13, 2020. Doi:10.1101/2020.04.16.20068528
- [29] Semenova NV, Madaeva IM, Kolesnikov SI, Solodova EI, Kolesnikova LI. Insomnia in peri- and postmenopausal women: Plasma lipids, lipid peroxidation and some antioxidant system parameters. Neuropsychiatry (London). 2018;8(4):1452-1460. DOI:10.4172/Neuropsychiatry.1000477
- [30] Reed TT. Lipid peroxidation and neurodegenerative disease. Free Radical Biology & Medicine. 2011;51(7):1302-1319. DOI:10.1016/j. freeradbiomed.2011.06.027
- [31] Wardle-Pinkston S, Slavish DC, Taylor DJ. Insomnia and cognitive performance: A systematic review and meta-analysis. Sleep Medicine Reviews. 2019;48:101205. DOI:10.1016/j. smrv.2019.07.008
- [32] The American Academy of Sleep Medicine. CDC reports that insufficient

- sleep is common on school nights (aasm. org). 2018 [Accessed 13 January 2023]
- [33] Choudhry AA, Shahzeen F, Choudhry SA, Batool N, Murtaza F, Dilip A, et al. Impact of COVID-19 infection on quality of sleep. Cureus. 2021;**13**(9):e18182. DOI:10.7759/cureus.18182
- [34] Mandelkorn U, Genzer S, Choshen-Hillel S, Reiter J, Meira E, Cruz M, et al. Escalation of sleep disturbances amid the COVID-19 pandemic: A cross-sectional international study. Journal of Clinical Sleep Medicine. 2021;17(1):45-53. DOI:10.5664/jcsm.8800
- [35] Marelli S, Castelnuovo A, Somma A, Castronovo V, Mombelli S, Bottoni D, et al. Impact of COVID-19 lockdown on sleep quality in university students and administration staff. Journal of Neurology. 2021;268(1):8-15. DOI:10.1007/s00415-020-10056-6
- [36] Kokou-Kpolou CK, Megalakaki O, Laimou D, Kousouri M. Insomnia during COVID-19 pandemic and lockdown: Prevalence, severity, and associated risk factors in French population. Psychiatry Research. 2020;**290**:113128. DOI:10.1016/j.psychres.2020.113128
- [37] Prime H, Andrews K, McTavish J, Harris M, Janus M, Bennett T, et al. The application of positive parenting interventions to academic school readiness: A scoping review. Child: Care, Health and Development. 2021;47(1):1-14. DOI:10.1111/cch.12810
- [38] Douglas KD, Smith KK, Stewart MW, Walker J, Mena L, Zhang L. Exploring Parents' intentions to monitor and mediate adolescent social media use and implications for school nurses. The Journal of School Nursing. 2020;2020:105. DOI:10.1177/1059840520983286

- [39] Altena E, Baglioni C, Espie CA, Ellis J, Gavriloff D, Holzinger B, et al. Dealing with sleep problems during home confinement due to the COVID-19 outbreak: Practical recommendations from a task force of the European CBT-I academy. Journal of Sleep Research. 2020;29(4):e13052. DOI:10.1111/jsr.13052
- [40] Wakefield JRH, Bowe M, Kellezi B, Butcher A, Groeger JA, Longitudinal associations between family identification. Loneliness, depression, and sleep quality. British Journal of Health Psychology. 2020;25(1):1-16. DOI:10.1111/bjhp.12391
- [41] Grossman ES, Hoffman YSG, Palgi Y, Shrira A. COVID-19 related loneliness and sleep problems in older adults: Worries and resilience as potential moderators. Personality and Individual Differences. 2021;**168**:110371. DOI:10.1016/j.paid.2020.110371
- [42] Brooks JD, Bronskill SE, Fu L, Saxena FE, Arneja J, Pinzaru VB, et al. Identifying children and youth with autism Spectrum disorder in electronic medical records: Examining health system utilization and comorbidities. Autism Research. 2021;**14**(2):400-410. DOI:10.1002/aur.2419
- [43] Vignola A, Lamoureux C, Bastien CH, Morin CM. Effects of chronic insomnia and use of benzodiazepines on daytime performance in older adults. The Journals of Gerontology. Series B, Psychological Sciences and Social Sciences. 2000;55(1):P54-P62. DOI:10.1093/geronb/55.1.p54
- [44] Mirsky AF, Anthony BJ, Duncan CC, Ahearn MB, Kellam SG. Analysis of the elements of attention: A neuropsychological approach. Neuropsychology Review. 1991;2(2):109-145. DOI:10.1007/BF01109051

- [45] Altena E, Van Der Werf YD, Strijers RL, Van Someren EJ. Sleep loss affects vigilance: Effects of chronic insomnia and sleep therapy. Journal of Sleep Research. 2008;17(3):335-343. DOI:10.1111/j.1365-2869.2008.00671.x
- [46] Shekleton JA, Rogers NL, Rajaratnam SM. Searching for the daytime impairments of primary insomnia. Sleep Medicine Reviews. 2010;14(1):47-60. DOI:10.1016/j. smrv.2009.06.001
- [47] Edinger JD, Means MK, Carney CE, Krystal AD. Psychomotor performance deficits and their relation to prior nights' sleep among individuals with primary insomnia. Sleep. 2008;**31**(5):599-607. DOI:10.1093/sleep/31.5.599
- [48] Orff HJ, Drummond SP, Nowakowski S, Perils ML. Discrepancy between subjective symptomatology and objective neuropsychological performance in insomnia. Sleep. 2007;**30**(9):1205-1211. DOI:10.1093/ sleep/30.9.1205
- [49] Boyle J, Trick L, Johnsen S, Roach J, Rubens R. Next-day cognition, psychomotor function, and drivingrelated skills following nighttime administration of eszopiclone. Human Psychopharmacology. 2008;**23**(5):385-397. DOI:10.1002/hup.936
- [50] Haimov I, Hanuka E, Horowitz Y. Chronic insomnia and cognitive functioning among older adults. Behavioral Sleep Medicine. 2008;**6**(1):32-54. DOI:10.1080/15402000701796080
- [51] Shekleton JA, Flynn-Evans EE, Miller B, Epstein LJ, Kirsch D, Brogna LA, et al. Neurobehavioral performance impairment in insomnia: Relationships with self-reported sleep and daytime functioning. Sleep. 2014;37(1):107-116. DOI:10.5665/sleep.3318

- 4-Hydroxynonenal Is Linked to Sleep and Cognitive Disturbances in Children: Once upon the... DOI:http://dx.doi.org/10.5772/intechopen.110285
- [52] Fortier-BrochuE, Beaulieu-BonneauS, Ivers H, Morin CM. Insomnia and daytime cognitive performance: A meta-analysis. Sleep Medicine Reviews. 2012;**16**(1):83-94. DOI:10.1016/j. smrv.2011.03.008
- [53] Cellini N, de Zambotti M, Covassin N, Sarlo M, Stegagno L. Impaired off-line motor skills consolidation in young primary insomniacs. Neurobiology of Learning and Memory. 2014;**114**:141-147. DOI:10.1016/j.nlm.2014.06.006
- [54] Sandi C. Stress and cognition. Wiley Interdisciplinary Reviews: Cognitive Science. 2013;4(3):245-261. DOI:10.1002/wcs.1222
- [55] Calvo MG, Gutierrez-Garcia A. Cognition and stress. In: Fink G, editor. Stress: Concepts, Cognition, Emotion, and Behavior. San Diego, CA, USA: Academic Press; 2016. pp. 139-144. Available online: https://www.sciencedirect.com/science/article/pii/B9780128009512000169
- [56] The American Physiological Association (APA). Stress in America™: Are Teens Adopting Adults' Stress Habits? was developed, reviewed and produced by the following team of experts. 2014. Accessed online on 13 January 2023
- [57] Tang YP, Shimizu E, Dube GR, Rampon C, Kerchner GA, Zhuo M, et al. Genetic enhancement of learning and memory in mice. Nature. 1999;**401**(6748):63-69.DOI:10.1038/43432
- [58] Riedel A, Stober F, Richter K, Fischer KD, Miettinen R, Budinger E. VGLUT3-immunoreactive afferents of the lateral septum: Ultrastructural evidence for a modulatory role of glutamate. Brain Structure & Function. 2013;218(1):295-301. DOI:10.1007/s00429-012-0395-4

- [59] Li XK, Tu B, Zhang XA, Xu W, Chen JH, Zhao GY, et al. Dysregulation of glutamine/glutamate metabolism in COVID-19 patients: A metabolism study in African population and mini meta-analysis. Journal of Medical Virology. 2023;95(1):e28150. DOI:10.1002/jmv.28150
- [60] Sonnewald U, Westergaard N, Schousboe A, Svendsen JS, Unsgard G, Petersen SB. Direct demonstration by [13C] NMR spectroscopy that glutamine from astrocytes is a precursor for GABA synthesis in neurons. Neurochemistry International. 1993;22(1):19-29. DOI:10.1016/0197-0186(93)90064-c
- [61] Petroff OA. GABA and glutamate in the human brain. The Neuroscientist. 2002;8(6):562-573. DOI:10.1177/1073858402238515
- [62] Schwartz RD, Mindlin MC. Inhibition of the GABA receptorgated chloride ion channel in brain by noncompetitive inhibitors of the nicotinic receptor-gated cation channel. The Journal of Pharmacology and Experimental Therapeutics. 1988;244(3):963-970 ISSN (Linking): 0022-3565
- [63] Roberts E, Frankel S. Gammaaminobutyric acid in brain: Its formation from glutamic acid. The Journal of Biological Chemistry. 1950;**187**(1):55-63 ISSN (Linking): 0021-9258
- [64] Roberts E, Eidelberg E. Metabolic and neurophysiological roles of gamma-aminobutyric acid. International Review of Neurobiology. 1960;2:279-332. DOI:10.1016/s0074-7742(08)60125-7
- [65] Bouche N, Lacombe B, Fromm H. GABA signaling: A conserved and ubiquitous mechanism. Trends in Cell Biology. 2003;**13**(12):607-610. DOI:10.1016/j.tcb.2003.10.001

- [66] Martin DL, Rimvall K. Regulation of gamma-aminobutyric acid synthesis in the brain. Journal of Neurochemistry. 1993;**60**(2):395-407. DOI:10.1111/j.1471-4159.1993.tb03165.x
- [67] Dionisio L, Jose De Rosa M, Bouzat C, Esandi MC. An intrinsic GABAergic system in human lymphocytes. Neuropharmacology. 2011;**60**(2-3):513-519. DOI:10.1016/j. neuropharm.2010.11.007
- [68] Foster JD, Kitchen I, Bettler B, Chen Y. GABAB receptor subtypesdifferentially modulate synaptic inhibition in the dentate gyrus to enhancegranule cell output. British Journal of Pharmacology. 2013;**168**:1808-1819. DOI:10.1111/bph.12073
- [69] Haeney CF, Kinney JW. Role of GABAB receptors in learning and memory and neurological disorders. Neurosci Behav Rev. 2016;**63**:1-28. DOI:10.1016/j.neubiorev.2016.01.007
- [70] Rashmi D, Zanan R, John S, Khandagale K, Nadaf A. Chapter 13 - g-aminobutyric acid (GABA): Biosynthesis, role, commercial production, and applications. Studies in Natural Products Chemistry. 2018;57:413-452. DOI:10.1016/ B978-0-444-64057-4.00013-2
- [71] Gottesmann C. GABA mechanisms and sleep. Neuroscience. 2002;**111**:231-239. DOI:10.1016/S0306-4522(02)00034-9
- [72] Nemeroff CB. The Role of GABA in the Pathophysiology and Treatment of anxiety disorders. Psychopharmacology Bulletin. 2003;**37**:133-146
- [73] Jie F, Yin G, Yang W, Yang M, Gao S, Lv J, et al. Stress in regulation of GABA amygdala system and relevance to neuropsychiatric diseases. Frontiers in

- Neuroscience. 2018;**12**:562. DOI:10.3389/fnins.2018.00562
- [74] Nuss P. Anxiety disorders and GABA neurotransmission: A disturbance of modulation. Neuropsychiatric Disease and Treatment. 2015;**11**:165-175. DOI:10.2147/NDT.S58841
- [75] Riemann D, Nissen C, Palagini L, Otte A, Perlis ML, Spiegelhalder K. The neurobiology, investigation, and treatment of chronic insomnia. Lancet Neurology. 2015;14:547-558. DOI:10.1016/S1474-4422(15)00021-6
- [76] Lichstein KL, Nau SD, Wilson NM, Aguillard RN, Lester KW, Bush AJ, et al. Psychological treatment of hypnotic-dependent insomnia in a primarily older adult sample. Behaviour Research and Therapy. 2013;51:787-796
- [77] Lozano-Soldevilla D, ter Huurne N, Cools R, Jensen O. GABAergic modulation of visual gamma and alpha oscillations and its consequences for working memory performance. Current Biology. 2014;**24**(24):2878-2887. DOI:10.1016/j.cub.2014.10.017
- [78] Schmidt-Wilcke T, Fuchs E, Funke K, Vlachos A, Muller-Dahlhaus F, Puts NAJ. GABA-from inhibition to cognition: Emerging concepts. The Neuroscientist. 2018;24(5):501-515. DOI:10.1177/1073858417734530
- [79] Torta DM, Castelli L, Zibetti M, Lopiano L, Geminiani G. On the role of dopamine replacement therapy in decision-making, working memory, and reward in Parkinson's disease: Does the therapy-dose matter? Brain and Cognition. 2009;71(2):84-91. DOI:10.1016/j.bandc.2009.04.003
- [80] Jutras MJ, Buffalo EA. Synchronous neural activity and memory formation. Current Opinion in Neurobiology.

2010;**20**(2):150-155. DOI:10.1016/j. conb.2010.02.006

- [81] Hollnagel JO, Maslarova A, Haq RU, Heinemann U. GABAB receptor dependent modulation of sharp wave-ripple complexes in the rat hippocampus in vitro. Neuroscience Letters. 2014;574:15-20. DOI:10.1016/j. neulet.2014.04.045
- [82] Dubrovina NI, Zinov'ev DR. Contribution of GABA receptors to extinction of memory traces in normal conditions and in a depression-like state. Neuroscience and Behavioral Physiology. 2008;38(8):775-779. DOI:10.1007/s11055-008-9045-y
- [83] Nakagawa Y, Iwasaki T, Ishima T, Kimura K. Interaction between benzodiazepine and GABA-A receptors in state-dependent learning. Life Sciences. 1993;52(24):1935-1945. DOI:10.1016/0024-3205(93)90634-f
- [84] Pontes A, Zhang Y, Hu W. Novel functions of GABA signaling in adult neurogenesis. Frontiers in Biology (Beijing). 2013;8(5):496-507. DOI:10.1007/s11515-013-1270-2
- [85] Kreisel T, Frank MG, Licht T, et al. Dynamic microglial alterations underlie stress-induced depressive-like behavior and suppressed neurogenesis Molecular Psychiatry. 2014;19(6):699-709. Doi: 10.1038/mp.2013.155
- [86] Nunan R, Sivasathiaseelan H, Khan D, Zaben M, Gray W. Microglial VPAC1R mediates a novel mechanism of neuroimmune-modulation of hippocampal precursor cells via IL-4 release. Glia. 2014;62(8):1313-1327. DOI:10.1002/glia.22682
- [87] Fatemi SH, Reutiman TJ, Folsom TD, Rooney RJ, Patel DH, Thuras PD. mRNA and protein levels for GABAAalpha4,

- alpha5, beta1 and GABABR1 receptors are altered in brains from subjects with autism. Journal of Autism and Developmental Disorders. 2010;**40**(6):743-750. DOI:10.1007/s10803-009-0924-z
- [88] Enticott PG, Kennedy HA, Rinehart NJ, Tonge BJ, Bradshaw JL, Fitzgerald PB. GABAergic activity in autism spectrum disorders: An investigation of cortical inhibition via transcranial magnetic stimulation. Neuropharmacology. 2013;68:202-209. DOI:10.1016/j.neuropharm.2012.06.017
- [89] DeLorey TM, Sahbaie P, Hashemi E, Homanics GE, Clark JD. Gabrb3 gene deficient mice exhibit impaired social and exploratory behaviors, deficits in non-selective attention and hypoplasia of cerebellar vermal lobules: A potential model of autism spectrum disorder. Behavioural Brain Research. 2008;187(2):207-220. DOI:10.1016/j. bbr.2007.09.009
- [90] Alroughani R, Akhtar S, Ahmed SF, Behbehani R, Al-Abkal J, Al-Hashel J. Incidence and prevalence of pediatric onset multiple sclerosis in Kuwait: 1994-2013. Journal of the Neurological Sciences. 2015;353(1-2):107-110. DOI:10.1016/j.jns.2015.04.025
- [91] Demakova EV, Korobov VP, Lemkina LM. Determination of gammaaminobutyric acid concentration and activity of glutamate decarboxylase in blood serum of patients with multiple sclerosis. Klin Lab Diagnostic. 2003;4:15-17
- [92] Garjani A, Middleton RM, Hunter R, Tuite-Dalton KA, Coles A, Dobson R, et al. COVID-19 is associated with new symptoms of multiple sclerosis that are prevented by disease modifying therapies. Multiple Sclerosis and Related Disorders. 2021;52:102939. DOI:10.1016/j. msard.2021.102939

[93] Zhang B, Vogelzang A, Miyajima M, Sugiura Y, Wu Y, Chamoto K, et al. B cell-derived GABA elicits IL-10(+) macrophages to limit anti-tumor immunity. Nature. 2021;**599**(7885):471-476

[94] Tian J, Chau C, Hales TG, Kaufman DL. GABA(A) receptors mediate inhibition of T cell responses. Journal of Neuroimmunology. 1999;**96**(1):21-28. DOI:10.1016/ s0165-5728(98)00264-1

[95] Prud'Homme GJ, Glinka Y, Hasilo C, Paraskevas S, Li X, Wang Q. GABA protects human islet cells against the deleterious effects of immunosuppressive drugs and exerts immunoinhibitory effects alone. Transplantation. 2013;96(7):616-623. DOI:10.1097/TP.0b013e31829c24be

[96] Tian J, Lu Y, Zhang H, Chau CH, Dang HN, Kaufman DL. Gamma-aminobutyric acid inhibits T cell autoimmunity and the development of inflammatory responses in a mouse type 1 diabetes model. Journal of Immunology. 2004;173(8):5298-5304. DOI:10.4049/jimmunol.173.8.5298

[97] Wang Y, Feng D, Liu G, Luo Q, Xu Y, Lin S, et al. Gamma-aminobutyric acid transporter 1 negatively regulates T cell-mediated immune responses and ameliorates autoimmune inflammation in the CNS. Journal of Immunology. 2008;**181**(12):8226-8236. DOI:10.4049/jimmunol.181.12.8226

[98] Beales PE, Hawa M, Williams AJ, Albertini MC, Giorgini A, Pozzilli P. Baclofen, a gamma-aminobutyric acid-b receptor agonist, delays diabetes onset in the non-obese diabetic mouse. Acta Diabetologica. 1995;32(1):53-56. DOI:10.1007/BF00581047

[99] Tian J, Dang H, Wallner M, Olsen R, Kaufman DL. Homotaurine, a safe blood-brain barrier permeable GABAA-R-specific agonist, ameliorates disease in mouse models of multiple sclerosis. Scientific Reports. 2018;8(1):16555. DOI:10.1038/s41598-018-32,733-3

[100] Tian J, Dang H, O'Laco KA, Song M, Tiu BC, Gilles S, et al. Homotaurine Treatment Enhances CD4(+) and CD8(+) Regulatory T Cell Responses and Synergizes with Low-Dose Anti-CD3 to Enhance Diabetes Remission in Type 1 Diabetic Mice. Immunohorizons. 2019;3(10):498-510. DOI:10.4049/immunohorizons.1900019

[101] Bhandage AK, Jin Z, Korol SV, Shen Q, Pei Y, Deng Q, et al. GABA regulates release of inflammatory cytokines from peripheral blood mononuclear cells and CD4(+) T Cells and Is Immunosuppressive in Type 1 Diabetes. Ebio Medicine. 2018;**30**:283-294. DOI:10.1016/j.ebiom.2018.03.019

[102] Vabret N, Britton GJ, Gruber C, Hegde S, Kim J, Kuksin M, et al. Immunology of COVID-19: Current State of the Science. Immunity. 2020;52(6):910-941. DOI:10.1016/j. immuni.2020.05.002

[103] Januzi L, Poirier JW, Maksoud MJE, Xiang YY, Veldhuizen RAW, Gill SE, et al. Autocrine GABA signaling distinctively regulates phenotypic activation of mouse pulmonary macrophages.

Cellular Immunology. 2018;332:7-23.

DOI:10.1016/j.cellimm.2018.07.001

[104] Forstera B, Castro PA, Moraga-Cid G, Aguayo LG. Potentiation of Gamma Aminobutyric Acid Receptors (GABAAR) by Ethanol: How are inhibitory receptors affected? Frontiers in Cellular Neuroscience. 2016;**10**:114. DOI:10.3389/fncel.2016.00114

[105] Paul AM, Branton WG, Walsh JG, Polyak MJ, Lu JQ, et al. GABA transport

and neuroinflammation are coupled in multiple sclerosis: regulation of the GABA transporter-2 by Ganaxolone. Neuroscience. 2014;273:24-38. DOI:10.1016/j.neuroscience.2014.04.037

[106] Fu CY, He XY, Li XF, Zhang X, Huang ZW, Li J, et al. NefiracetamAttenuatesPro-Inflammatory Cytokines and GABA Transporter in Specific Brain Regions of Rats with Post-Ischemic Seizures. Cellular Physiology and Biochemistry. 2015;37(5):2023-2031. DOI:10.1159/000438562

[107] Su J, Yin J, Qin W, Sha S, Xu J, Jiang C. Role for pro-inflammatory cytokines in regulating expression of GABA transporter type 1 and 3 in specific brain regions of kainic acid-induced status epilepticus. Neurochemical Research. 2015;40(3):621-627. DOI:10.1007/s11064-014-1504-y

[108] Hernandez-Rabaza V,
Cabrera-Pastor A, Taoro-Gonzalez L,
Gonzalez-Usano A, Agusti A, Balzano T,
et al. Neuroinflammation increases
GABAergic tone and impairs cognitive
and motor function in hyperammonemia
by increasing GAT-3 membrane
expression. Reversal by sulforaphane by
promoting M2 polarization of microglia.
Journal of Neuroinflammation.
2016;13(1):83. DOI:10.1186/
s12974-016-0549-z

[109] Huang T, Zhang Y, Wang C, Gao J. Propofol reduces acute lung injury by up-regulating gamma-aminobutyric acid type a receptors. Experimental and Molecular Pathology. 2019;**110**:104295. DOI:10.1016/j.yexmp.2019.104295

[110] Mahmoud K, Ammar A. Immunomodulatory effects of anesthetics during thoracic surgery. Anesthesiology Research Practise. 2011;**2011**:317410. DOI:10.1155/2011/317410

[111] Pan CF, Shen MY, Wu CJ, Hsiao G, Chou DS, Sheu JR. Inhibitory mechanisms of gabapentin, an antiseizure drug, on platelet aggregation. The Journal of Pharmacy and Pharmacology. 2007;59(9):1255-1261. DOI:10.1211/jpp.59.9.0010

[112] Lin KH, Lu WJ, Wang SH, Fong TH, Chou DS, Chang CC, et al. Characteristics of endogenous gamma-aminobutyric acid (GABA) in human platelets: Functional studies of a novel collagen glycoprotein VI inhibitor. Journal of Molecular Medicine (Berlin, Germany). 2014;92(6):603-614. DOI:10.1007/s00109-014-1140-7

[113] Hottz ED, Azevedo-Quintanilha IG, Palhinha L, Teixeira L, Barreto EA, Pao CRR, et al. Platelet activation and platelet-monocyte aggregate formation trigger tissue factor expression in patients with severe COVID-19. Blood. 2020;136(11):1330-1341. DOI:10.1182/blood.2020007252

[114] Tian J, Middleton B, Kaufman DL. GABA(A)-Receptor Agonists Limit Pneumonitis and Death in Murine Coronavirus-Infected Mice. Viruses. 2021;**13**(6):966. DOI:10.3390/v13060966

[115] Khanolkar A, Fulton RB, Epping LL, Pham NL, Tifrea D, Varga SM, et al. T cell epitope specificity and pathogenesis of mouse hepatitis virus-1-induced disease in susceptible and resistant hosts. Journal of Immunology. 2010;185(2):1132-1141. DOI:10.4049/jimmunol.0902749

[116] Krystal JH, D'Souza DC, Sanacora G, Goddard AW, Charney DS. Current perspectives on the pathophysiology of schizophrenia, depression, and anxiety disorders. The Medical Clinics of North America. 2001;85(3):559-577. DOI:10.1016/ s0025-7125(05)70329-1 [117] Morinushi T, Masumoto Y, Kawasaki H, Takigawa M. Effect on electroencephalogram of chewing flavored gum. Psychiatry and Clinical Neurosciences. 2000;54(6):645-651. DOI:10.1046/j.1440-1819.2000.00772.x

[118] Abdou AM, Higashiguchi S, Horie K, Kim M, Hatta H, Yokogoshi H. Relaxation and immunity enhancement effects of gamma-aminobutyric acid (GABA) administration in humans. BioFactors. 2006;**26**(3):201-208. DOI:10.1002/biof.5520260305

[119] Tomasi TB, Gray HM. Structure and function of immunoglobulin A. Progress in Allergy. 1972;**16**:81-213

[120] Asim M, Jiang S, Yi L, Chen W, Sun L, Zhao L. Glutamine is required for red-spotted grouper nervous necrosis virus replication via replenishing the tricarboxylic acid cycle. Virus Research. 2017;227:245-248. DOI:10.1016/j. virusres.2016.11.007

[121] Kalyanaraman B. Reactive oxygen species, proinflammatory and immunosuppressive mediators induced in COVID-19: Overlapping biology with cancer. RSC Chemical Biology. 2021;2(5):1402-1414. DOI:10.1039/d1cb00042j

[122] Kumar P, Osahon O, Vides DB, Hanania N, Minard CG, Rajagopal V, et al. Severe glutathione deficiency, oxidative stress and oxidant, damage in adults hospitalized with COVID-19: Implication, for GlyNAC (Glycine and N-Acetylcysteine) supplementation. Antioxidants. 2022;11:50. DOI:10.3390/antiox11010050

[123] Yowtak J, Wang J, Kim HY, Lu Y, Chung K, Chung JM. Effect of antioxidant treatment on spinal GABA neurons in a neuropathic pain model in the mouse. Pain. 2013;154(11):2469-2476. DOI:10.1016/j.pain.2013.07.024 [124] Ludewig B, Jaggi M, Dumrese T, Brduscha-Riem K, Odermatt B, Hengartner H. Hypercholesterolemia exacerbates virus-induced immunopathologic liver disease via suppression of antiviral cytotoxic T cell responses. Journal of Immunology. 2001;**166**(5):3369-3376. DOI:10.4049/jimmunol.166.5.3369

[125] Wu Q, Zhou L, Sun X, Yan Z, Hu C, Wu J, et al. Altered lipid metabolism in recovered SARS patients twelve years after infection. Scientific Reports. 2017;7:9110. DOI:10.1038/s41598-017-09536-z

[126] Nguyen A, Guedan A, Mousnier A, Swieboda D, Zhang Q, Horkai D, et al. Host lipidome analysis during rhinovirus replication in HBECs identifies potential therapeutic targets. Journal of Lipid Research. 2018;59(9):1671-1684. DOI:10.1194/jlr.M085910

[127] Xu K, Nagy PD. RNA virus replication depends on enrichment of phosphatidylethanolamine at replication sites in subcellular membranes. Proceedings of the National Academy Science USA. 2015;**112**:E1782-E1791

[128] Grieb P, Swiatkiewicz M, Prus K, Rejdak K. Hypoxia may be a determinative factor in COVID-19 progression. Current Research Pharmacological Drug Discovery. 2021;2:100030. DOI:10.1016/j. crphar.2021.100030

[129] Hannun YA, Obeid LM. Sphingolipids and their metabolism in physiology and disease. Nature Reviews. Molecular Cell Biology. 2018;**19**(3):175-191. DOI:10.1038/nrm.2017.107

[130] Chakinala RC, Khatri A, Gupta K, Koike K, Epelbaum O. Sphingolipids in COPD. European Respiratory Review. 2019;**28**(154). DOI:10.1183/16000617.0047-2019

[131] Yin Y, Xu L, Porter NA. Free radical lipid peroxidation: Mechanisms and analysis. Chemical Reviews. 2011;**111**(10):5944-5972. DOI:10.1021/cr200084z

[132] Fruhwirth GO, Loidl A, Hermetter A. Oxidized phospholipids: From molecular properties to disease. Biochimica et Biophysica Acta, Molecular Basis of Disease. 2007;**1772**(7):718-736. DOI:10.1016/j.bbadis.2007.04.009

[133] Kinnunen PKJ, Kaarniranta K, Mahalka A. Protein oxidized phospholipid interactions in cellular signaling for cell death: From biophysics to clinical correlations. Biochimica et Biophysica Acta. 2012;1818(10):2446-2455. DOI:10.1016/j. bbamem.2012.04.008

[134] Gracanin M, Hawkins CL, Pattison DI, Davies MJ. Singlet-oxygenmediated amino acid and protein oxidation: Formation of tryptophan peroxides and decomposition products. Free Radical Biology & Medicine. 2009;47(1):92-102. DOI:10.1016/j. freeradbiomed.2009.04.015

[135] Zarkovic N, Cipak A, Jaganjac M, Borovic S, Zarkovic K. Pathophysiological relevance of aldehydic protein modifications. Journal of Proteomics. 2013;**92**:239-247. DOI:10.1016/j. jprot.2013.02.004

[136] Ayala A, Munoz MF, Arguelles S. Lipid peroxidation: Production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. Oxidative Medical Cell Longevity. 2014;**2014**:360438. DOI:10.1155/2014/360438

[137] Orioli M, Aldini G, Beretta G, Facino RM, Carini M. LC-ESI-MS/ MS determination of 4-hydroxy-trans-2-nonenal Michael adducts with

cysteine and histidine-containing peptides as early markers of oxidative stress in excitable tissues. Journal of Chromatography. B, Analytical Technologies in the Biomedical and Life Sciences. 2005;827(1):109-118. DOI:10.1016/j.jchromb.2005.04.025

[138] Dringen R. Metabolism and functions of glutathione in brain. Progress in Neurobiology. 2000;**62**(6):649-671. DOI:10.1016/s0301-0082(99)00060-x

[139] Pastore A, Federici G, Bertini E, Piemonte F. Analysis of glutathione: Implication in redox and detoxification. Clinica Chimica Acta. 2003;333(1):19-39. DOI:10.1016/s0009-8981(03)00200-6

[140] Pastore A, Tozzi G, Gaeta LM, Giannotti A, Bertini E, Federici G, et al. Glutathione metabolism and antioxidant enzymes in children with Down syndrome. Journal of Peadiatric. 2003;142(5):583-585. DOI:10.1067/mpd.2003.203

[141] Espinosa JM. Down Syndrome and COVID-19: A Perfect Storm? Cell Reports in Medicine. 2020;**1**(2):100019. DOI:10.1016/j.xcrm.2020.100019

[142] Zhu Y, Carvey PM, Ling Z. Agerelated changes in glutathione and glutathione-related enzymes in rat brain. Brain Research. 2006;**1090**(1):35-44. DOI:10.1016/j.brainres.2006.03.063

[143] Poli G, Schaur RJ, Siems WG, Leonarduzzi G. 4-hydroxynonenal: A membrane lipid oxidation product of medicinal interest. Medicinal Research Reviews. 2008;**28**(4):569-631. DOI:10.1002/med.20117

[144] Alary J, Gueraud F, Cravedi JP. Fate of 4-hydroxynonenal in vivo: Disposition and metabolic pathways. Molecular Aspects of Medicine. 2003;**24**(4-5):177-187. DOI:10.1016/s0098-2997(03)00012-8

[145] Castegna A, Lauderback CM, Mohmmad-Abdul H, Butterfield DA. Modulation of phospholipid asymmetry in synaptosomal membranes by the lipid peroxidation products, 4-hydroxynonenal and acrolein: Implications for Alzheimer's disease. Brain Research. 2004;1004(1-2):193-197. DOI:10.1016/j.brainres.2004.01.036

[146] LoPachin RM, Gavin T, Geohagen BC, Das S. Neurotoxic mechanisms of electrophilic type-2 alkenes: Soft interactions described by quantum mechanical parameters. Toxicological Sciences. 2007;98(2):561-570. DOI:10.1093/toxsci/kfm127

[147] Vander Jagt DL, Hunsaker LA, Vander Jagt TJ, Gomez MS, Gonzales DM, Deck LM, et al. Inactivation of glutathione reductase by 4-hydroxynonenal and other endogenous aldehydes. Biochemical Pharmacology. 1997;53(8):1133-1140

[148] Carbone DL, Doorn JA, Kiebler Z, Ickes BR, Petersen DR. Modification of heat shock protein 90 by 4-hydroxynonenal in a rat model of chronic alcoholic liver disease. The Journal of Pharmacology and Experimental Therapeutics. 2005;**315**(1):8-15. DOI:10.1124/jpet.105.088088

[149] Roede JR, Jones DP. Reactive species and mitochondrial dysfunction: Mechanistic significance of 4-hydroxynonenal. Environmental and Molecular Mutagenesis. 2010;**51**(5):380-390. DOI:10.1002/em.20553

[150] Honzatko A, Brichac J, Murphy TC, Reberg A, Kubatova A, Smoliakova IP. Enantioselective metabolism of trans-4hydroxy-2-nonenalbybrainmitochondria. Free Radical Biology & Medicine. 2005;**39**(7):913-924. DOI:10.1016/j. freeradbiomed.2005.05.010

[151] Crabb DW, Galli A, Fischer M, You M, et al. Molecular mechanisms of alcoholic fatty liver: Role of peroxisome proliferator-activated receptor alpha. Alcohol. 2004;**34**(1):35-38. DOI:10.1016/j.alcohol.2004.07.005

[152] Zheng R, Dragomir AC, Mishin V, Richardson JR, Heck DE, Laskin DL, et al. Differential metabolism of 4-hydroxynonenal in liver, lung and brain of mice and rats. Toxicology and Applied Pharmacology. 2014;**279**(1):43-52. DOI:10.1016/j.taap.2014.04.026

[153] Corradi M, Pignatti P, Manini P, Andreoli R, Goldoni M, Poppa M, et al. Comparison between exhaled and sputum oxidative stress biomarkers in chronic airway inflammation.

The European Respiratory
Journal. 2004;24(6):1011-1017.

DOI:10.1183/09031936.04.00002404

[154] Stopforth A, Burger BV, Crouch AM, Sandra P. Urinalysis of 4-hydroxynonenal, a marker of oxidative stress, using stir bar sorptive extractionthermal desorption-gas chromatography/ mass spectrometry. Journal of Chromatography. B, Analytical Technologies in the Biomedical and Life Sciences. 2006;834(1-2):134-140. DOI:10.1016/j.jchromb.2006.02.038

[155] Esterbauer H, Zollner H, Lang J. Metabolism of the lipid peroxidation product 4-hydroxynonenal by isolated hepatocytes and by liver cytosolic fractions. The Biochemical Journal. 1985;228(2):363-373. DOI:10.1042/bj2280363

[156] Dianzani MU. 4-hydroxynonenal from pathology to physiology. Molecular Aspects of Medicine. 2003;**24**(4-5):263-272. DOI:10.1016/s0098-2997(03)00021-9

[157] Curzio M, Torrielli MV, Giroud JP, Esterbauer H, Dianzani MU. Neutrophil chemotactic responses to aldehydes. Research Communication in Chemical Pathology and Pharmacology. 1982;36(3):463-476

[158] Paradisi L, Panagini C, Parola M, Barrera G, Dianzani MU. Effects of 4-hydroxynonenal on adenylate cyclase and 5'-nucleotidase activities in rat liver plasma membranes. Chemico-Biological Interactions. 1985;53(1-2):209-217. DOI:10.1016/s0009-2797(85)80097-1

[159] Rossi MA, Garramone A, Dianzani MU. Stimulation of phospholipase C activity by 4-hydroxynonenal; influence of GTP and calcium concentration. International Journal of Tissue Reactions. 1988;**10**(5):321-325

[160] Camandola S, Poli G, Mattson MP. The lipid peroxidation product 4-hydroxy-2,3-nonenal increases AP-1-binding activity through caspase activation in neurons. Journal of Neurochemistry. 2000;74(1):159-168. DOI:10.1046/j.1471-4159.2000.0740159.x

[161] Cajone F, Bernelli-Zazzera A. The action of 4-hydroxynonenal on heat shock gene expression in cultured hepatoma cells. Free Radical Research Communications. 1989;7(3-6):189-194. DOI:10.3109/10715768909087941

[162] Spycher S, Tabataba-Vakili S, O'Donnell VB, Palomba L, Azzi A. 4-hydroxy-2,3-trans-nonenal induces transcription and expression of aldose reductase. Biochemical and Biophysical Research Communications. 1996;**226**(2):512-516. DOI:10.1006/bbrc.1996.1386

[163] Basu-Modak S, Luscher P, Tyrrell RM. Lipid metabolite involvement in the activation of the human heme oxygenase-1 gene. Free Radical Biology & Medicine. 1996;**20**(7):887-897. DOI:10.1016/0891-5849(95)02182-5

[164] Liu RM, Shi MM, Giulivi C, Forman HJ. Quinones increase gamma-glutamyl transpeptidase expression by multiple mechanisms in rat lung epithelial cells. The American Journal of Physiology. 1998;274(3):L330-L336. DOI:10.1152/ajplung.1998.274.3.L330

[165] Siems W, Grune T. Intracellular metabolism of 4-hydroxynonenal. Molecular Aspects of Medicine. 2003;24(4-5):167-175. DOI:10.1016/s0098-2997(03)00011-6

[166] Subramaniam R, Roediger F, Jordan B, Mattson MP, Keller JN, Waeg G, et al. The lipid peroxidation product, 4-hydroxy-2-trans-nonenal, alters the conformation of cortical synaptosomal membrane proteins. Journal of Neurochemistry. 1997;69(3):1161-1169. DOI:10.1046/j.1471-4159.1997.69031161.x

[167] Shi Q, Vaillancourt F, Cote V, Fahmi H, Lavigne P, Afif H, et al. Alterations of metabolic activity in human osteoarthritic osteoblasts by lipid peroxidation end product 4-hydroxynonenal. Arthritis Research & Therapy. 2006;8(6):R159. DOI:10.1186/ ar2066

[168] Arunachalam G, Sundar IK, Hwang JW, Yao H, Rahman I. Emphysema is associated with increased inflammation in lungs of atherosclerosis-prone mice by cigarette smoke: Implications in comorbidities of COPD. Journal of Inflammation. 2010;7:34. DOI:10.1186/1476-9255-7-34

[169] Zarkovic K. 4-hydroxynonenal and neurodegenerative diseases. Molecular Aspects of Medicine. 2003;24(4-5):293-303. DOI:10.1016/s0098-2997(03)00024-4

[170] Bennaars-Eiden A, Higgins L, Hertzel AV, Kapphahn RJ, Ferrington DA, Bernlohr DA. Covalent modification of epithelial fatty acid-binding protein by 4-hydroxynonenal in vitro and in vivo. Evidence for a role in antioxidant biology. The Journal of Biological Chemistry. 2002;277(52):50693-50,702. DOI:10.1074/jbc.M209493200

[171] Rahman I, van Schadewijk AA, Crowther AJ, Hiemstra PS, Stolk J, MacNee W, et al. 4-Hydroxy-2-nonenal, a specific lipid peroxidation product, is elevated in lungs of patients with chronic obstructive pulmonary disease. American Journal of Respiratory and Critical Care Medicine. 2002;**166**(4):490-495. DOI:10.1164/rccm.2110101

[172] Dickinson DA, Forman HJ. Glutathione in defense and signaling: Lessons from a small thiol. Annals of the New York Academy of Sciences. 2002;973:488-504. DOI:10.1111/j.1749-6632.2002.tb04690.x

[173] Zhang H, Liu H, Dickinson DA, Liu RM, Postlethwait EM, Laperche Y, et al. gamma-Glutamyl transpeptidase is induced by 4-hydroxynonenal via EpRE/Nrf2 signaling in rat epithelial type II cells. Free Radical Biology & Medicine. 2006;40(8):1281-1292. DOI:10.1016/j. freeradbiomed.2005.11.005

[174] Malone PE, Hernandez MR. 4-Hydroxynonenal, a product of oxidative stress, leads to an antioxidant response in optic nerve head astrocytes. Experimental Eye Research. 2007;84(3):444-454. DOI:10.1016/j. exer.2006.10.020

[175] Yang Y, Sharma R, Sharma A, Awasthi S, Awasthi YC. Lipid peroxidation and cell cycle signaling: 4-hydroxynonenal, a key molecule in stress mediated signaling. Acta Biochemica Pol. 2003;50(2):319-336

[176] Schmidt H, Grune T, Muller R, Siems WG, Wauer RR. Increased levels of lipid peroxidation products malondialdehyde and 4-hydroxynonenal after perinatal hypoxia. Pediatric Research. 1996;40(1):15-20. DOI:10.1203/00006450-199,607,000-00003

[177] Grune T et al. Increased levels of 4-hydroxynonenal modified proteins in plasma of children with autoimmune diseases. Free Radical Biology & Medicine 1997;23(3):357-360. DOI: 10.1016/s0891-5849(96)00586-2.

[178] Lovell MA, Markesbery WR. Oxidative damage in mild cognitive impairment and early Alzheimer's disease. Journal of Neuroscience Research. 2007;85(14):3036-3040. DOI:10.1002/jnr.21346

[179] Zhong Z, Zeng T, Xie K, Zhang C, Chen J, Bi Y, et al. Elevation of 4-hydroxynonenal and malondialdehyde modified protein levels in cerebral cortex with cognitive dysfunction in rats exposed to 1-bromopropane. Toxicology. 2013;306:16-23. DOI:10.1016/j. tox.2013.01.022

[180] Bradley MA, Markesbery WR, Lovell MA. Increased levels of 4-hydroxynonenal and acrolein in the brain in preclinical Alzheimer disease. Free Radical Biology & Medicine. 2010;48(12):1570-1576. DOI:10.1016/j. freeradbiomed.2010.02.016

[181] Pedersen WA, Fu W, Keller JN, Markesbery WR, Appel S, Smith RG, et al. Protein modification by the lipid peroxidation product 4-hydroxynonenal in the spinal cords of amyotrophic lateral sclerosis patients. Annals of Neurology. 1998;44(5):819-824. DOI:10.1002/ana.410440518

[182] Fleuranceau-Morel P, Barrier L, Fauconneau B, Piriou A, Huguet F. Origin

of 4-hydroxynonenal incubationinduced inhibition of dopamine transporter and Na+/K+ adenosine triphosphate in rat striatal synaptosomes. Neursoci Lett. 1999;**277**(2):91-94. DOI:10.1016/s0304-3940(99)00652-7

[183] Shin Y, White BH, Uh M, Sidhu A. Modulation of D1-like dopamine receptor function by aldehydic products of lipid peroxidation. Brain Research. 2003;968(1):102-113. DOI:10.1016/s0006-8993(02)04279-8

[184] Moreau C, Devos D, Brunaud-Danel V, Defebvre L, Perez T, Destee A, et al. Elevated IL-6 and TNFalpha levels in patients with ALS: Inflammation or hypoxia? Neurology. 2005;**6**(12):1958-1960. DOI:10.1212/01. wnl.0000188907.97339.76

[185] Blanc EM, Kelly JF, Mark RJ, Waeg G, Mattson MP. 4-Hydroxynonenal, an aldehydic product of lipid peroxidation, impairs signal transduction associated with muscarinic acetylcholine and metabotropic glutamate receptors: Possible action on G alpha(q/11). Journal of Neurochemistry. 1997;69(2):570-578. DOI:10.1046/j.1471-4159.1997.69020570.x

[186] Re G, Azzimondi G, Lanzarini C, Bassein L, Vaona I, Guarnieri C. Plasma lipoperoxidative markers in ischaemic stroke suggest brain embolism. European Journal of Emergency Medicine. 1997;4(1):5-9

[187] Newcombe J, Li H, Cuzner ML. Low density lipoprotein uptake by macrophages in multiple sclerosis plaques: Implications for pathogenesis. Neuropathology and Applied Neurobiology. 1994;20(2):152-162. DOI:10.1111/j.1365-2990.1994.tb01174.x

[188] Stoy N, Mackay GM, Forrest CM, Christofides J, Egerton M, Stone TW, et al. Tryptophan metabolism and

oxidative stress in patients with Huntington's disease. Journal of Neurochemistry. 2005;**93**(3):611-623. DOI:10.1111/j.1471-4159.2005.03070.x

[189] Mark RJ, Keller JN, Kruman I, Mattson MP. Basic FGF attenuates amyloid beta-peptide-induced oxidative stress, mitochondrial dysfunction, and impairment of Na+/K + -ATPase activity in hippocampal neurons. Brain Research. 1997;756(1-2):205-214. DOI:10.1016/s0006-8993(97)00196-0

[190] Eichenbaum H. Hippocampus: Cognitive processes and neural representations that underlie declarative memory. Neuron. 2004;44(1):109-120. DOI:10.1016/j.neuron.2004.08.028

[191] Lisman J, Buzsaki G, Eichenbaum H, Nadel L, Ranganath C, Redish AD. Viewpoints: How the hippocampus contributes to memory, navigation and cognition. Nature Neuroscience. 2017;20(11):1434-1447. DOI:10.1038/nn.4661

[192] Taki Y, Hashizume H, Thyreau B, Sassa Y, Takeuchi H, Wu K, et al. Sleep duration during weekdays affects hippocampal gray matter volume in healthy children. NeuroImage. 2012;**60**(1):471-475. DOI:10.1016/j. neuroimage.2011.11.072

[193] Kreutzmann JC, Havekes R, Abel T, Meerlo P. Sleep deprivation and hippocampal vulnerability: Changes in neuronal plasticity, neurogenesis and cognitive function. Neuroscience. 2015;309:173-190. DOI:10.1016/j. neuroscience.2015.04.053

[194] Klein R, Soung A, Sissoko C, Nordvig A, Canoll P, Mariani M, et al. COVID-19 induces neuroinflammation and loss of hippocampal neurogenesis. Research Sequence. 2021;**2021**:1031 [195] Liu Y, Xu G, Sayre LM. Carnosine inhibits (E)-4-hydroxy-2-nonenal-induced protein cross-linking: Structural characterization of carnosine-HNE adducts. Chemical Research in Toxicology. 2003;**16**(12):1589-1597. DOI:10.1021/tx034160a

[196] Guiotto A, Calderan A, Ruzza P, Borin G. Carnosine and carnosine-related antioxidants: A review. Current Medicinal Chemistry. 2005;**12**(20):2293-2315. DOI:10.2174/0929867054864796

[197] The National Institutes of Health (NIH). Vitamin C. Fact Sheet for Health Porfessionals. Office of Dietary Supplements (ODS). Vitamin C - Health Professional Fact Sheet (nih.gov). 2021. Accessed online on 17 January 2023.

[198] National Institutes of Health (NIH). Vitamin E. Fact Sheet for Health Porfessionals. Office of Dietary Supplements (ODS). Vitamin E - Health Professional Fact Sheet (nih.gov). 2021. Accessed online on 17 January 2023.

[199] Bassett IB, Pannowitz DL.
Barnetson RSA comparative study of tea-tree oil versus benzoylperoxide in the treatment of acne. The Medical Journal of Australia. 1990;153(8):455-458.
DOI:10.5694/j.1326-5377.1990.tb126150.x

[200] Al-Sarraf H. Transport of 14C-gamma-aminobutyric acid into brain, cerebrospinal fluid and choroid plexus in neonatal and adult rats. Brain Research Development. 2002;139(2):121-129

[201] Yoto A, Murao S, Motoki M, Yokoyama Y, Horie N, Takeshima K, et al. Oral intake of gamma-aminobutyric acid affects mood and activities of central nervous system during stressed condition induced by mental tasks. Amino Acids. 2012;43(3):1331-1337. DOI:10.1007/s00726-011-1206-6

[202] Tu J, Jin Y, Zhuo J, Cao X, Liu G, Du H, et al. Exogenous GABA improves the antioxidant and anti-aging ability of silkworm (Bombyx mori). Food Chemistry. 2022;383:132400. DOI:10.1016/j.foodchem.2022.132400

